



UNIVERSITY OF TM
KWAZULU-NATAL
—
INYUVESI
YAKWAZULU-NATALI

**THE ASSOCIATION BETWEEN INCRETIN HORMONES CONCENTRATION
AND THE DEVELOPMENT OF DIET-INDUCED PREDIABETES.**

**Submitted as a dissertation component in fulfilment for the degree of Master of Medical
Science in the School of Laboratory Medicine and Medical Sciences, University of
KwaZulu- Natal**

By
Mzimela Nhlakanipho
218006756

2023

Supervisor: Dr A Khathi
Co- Supervisor: Mr A.M Sosibo

Bachelor of Medical Science Honors in Human Physiology School of Laboratory Medicine
and Medical Science: College of Health Sciences

PREFACE

Prolonged consumption of a diet high in carbohydrates and fats has been shown to lead to the development of insulin resistance and the onset of type 2 diabetes mellitus (T2DM). T2DM has been shown to be preceded by an intermediate hyperglycemic condition known as prediabetes. Prediabetes is characterised by glucose levels that are above normal but below the diabetes threshold, and studies have demonstrated that the complications associated with T2DM begin in the prediabetic state. Impaired incretin peptides in T2DM have been shown to maintain the abnormal concentrations of blood glucose and insulin resistance. However, the changes and association of incretin peptides in the development of prediabetes have not been characterised. A diet-induced prediabetes animal model that mimics the human condition of prediabetes was established in our laboratory. In the development of this animal model, we investigated the changes and the association of incretin peptides. When the animal model was developed and prediabetes had been induced, we further investigated the effect of a low carbohydrate, high fat diet on incretin peptides levels and observed their role in the reversal of prediabetes. Incretin hormone levels could potentially serve as biomarkers for early detection and risk stratification for prediabetic individuals. Identifying the biomarkers could lead to the development of targeted drugs or treatments that aim to modulate these hormone levels to prevent or treat prediabetes effectively. This knowledge can be particularly useful for individuals with high-risk dietary habits or those seeking to make dietary modifications to reduce their prediabetes risk.

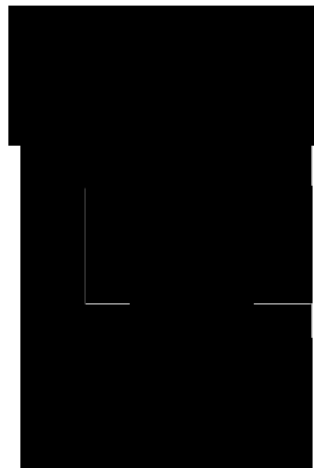
The experimental work described in this dissertation was conducted at the University of KwaZulu Natal, Westville Campus, Durban, South Africa. All work was conducted under the supervision of Dr. Andile Khathi and co-supervised by Mr. Aubrey Sosibo.

DECLARATION

I, Mr. Nhlakanipho Mzimela declare.

1. That the work described in this thesis has not been submitted to UKZN or other tertiary institutions for purposes of obtaining an academic qualification, whether by myself or any other party.
2. The dissertation is my own investing and work from other researchers is rephrased and referenced, or if their exact words are used, they are quoted and referenced.
3. Dr. Khathi and Mr. AM Sosibo supervised the research done in this study.

Student: Nhlakanipho Mzimela
Date: 15/04/2023



Signature:

Supervisor: Dr A. Khathi
Date: 19 /04/ 2023

Signature:

Co-Supervisor: Mr. A.M Sosibo

Signature: _____ **Date:** 19/04/ 2023

PLAGIARISM DECLARATION

School of Laboratory Medicine and Medical Sciences, College of Health Sciences

MASTER'S DEGREE IN MEDICAL SCIENCES 2023

1. I know that plagiarism is wrong. Plagiarism is to use another's work and pretend that it is one's
2. I have used the Vancouver convention for citation and referencing. Each contribution to, and quotation in, this dissertation from the works of other people has been attributed and has been cited and referenced.
3. This dissertation is my own work.
4. I have not allowed and will not allow anyone to copy my work with the intention of passing it off as his or her own work.

Signature: .

ACKNOWLEDGEMENTS

Words cannot express my gratitude to my supervisor Dr. Andile Khathi and my co-supervisor Mr Aubrey Sosibo for their patience and guidance throughout this journey. It was a pleasure working with you, and I hope this is our first of many studies together. I would also like to thank Mr. Dennis Makhubela (Bab' Dennis) for his technical assistance.

To Dr Mabuza-M, Dr Mxinwa and Dr Luvuno, thank you for generously providing your knowledge and expertise.

I'm also grateful to my colleagues and friends from the endocrinology group, especially Nosipho Dimba, Mathuli Ngema, Siphesihle Mkhize, Mbuyiselo Mzila, Ayanda Nkosi, Nombuso Xulu, and Palesa Mosili, thank you guys for your assistance in the lab and your support throughout. Additionally, this dissertation would not have been possible without the generous support from the National Research Foundation and UKZN College of Health Science who provided financial assistance.

Table of Contents

PREFACE.....	II
DECLARATION.....	III
PLAGIARISM DECLARATION	IV
ACKNOWLEDGEMENTS.....	V
LIST OF FIGURES	IX
LIST OF TABLES.....	XI
LIST OF ABBREVIATIONS.....	XII
LIST OF APPENDICES.....	XIII
STUDY OUTLINE.....	XIV
ABSTRACT	XV
Background.....	XV
Methods	XV
Results	XVI
Conclusion.....	XVI
Chapter 1: Literature Review.....	1
1. Introduction	1
2. Incretin hormones	2
2.1. The Physiological Role of GIP on Glucose Homeostasis	2
2.2. The Physiological Role of GLP-1 on Glucose Homeostasis	3
2.3. The Physiological Role of Ghrelin on Glucose Homeostasis.....	3
2.4. The Physiological Role of Leptin on Glucose Homeostasis	4
3. Type 2 Diabetes Mellitus.....	4
3.1 Changes in GIP levels in T2DM.....	5
3.2. Changes in GLP-1 levels in T2DM	5
3.3. Ghrelin and Leptin levels in T2DM	6
4. Prediabetes.....	6
5. Management of T2DM and prediabetes	7
6. HFHC animal model of prediabetes	7
7. Aim	7
8. Objectives	8
8.1. Manuscript 1.....	8
8.2. Manuscript 2.....	8
References	9
Chapter 2: Manuscript 1	17
Prologue 1.....	17
Abstract.....	19
1. Introduction	20

2. Materials and Methods	21
2.1 Chemicals and drugs.....	21
2.2 Animals.....	21
2.3.1. Induction of prediabetes	21
2.3.2. Experimental design	21
2.3.3. Blood collection and Termination	22
2.4. Biochemical Analysis	22
2.5. HOMA2-IR index.....	22
2.6. Statistical analysis.....	23
3. Results	23
3.1. Plasma Glucose.....	23
3.2. Plasma Insulin.....	24
3.4. Plasma GIP	25
3.5. Plasma Leptin	26
3.6. Plasma GLP-1	27
3.7. Plasma Ghrelin.....	28
3.8. Plasma Glycated Hemoglobin (HbA1c)	29
4. Discussion.....	30
Conclusion.....	33
Acknowledgments	33
Author Contributions	34
Conflict of Interest.....	34
Availability of Data and Materials	34
Ethics Approval and Consent to Participate	34
References	35
Chapter 3: Manuscript 2	40
Prologue 2.....	40
Abstract.....	42
1. Introduction	43
2. Methods and Materials	44
2.1. Chemicals and drugs.....	44
2.2. Animals and housing	44
2.3.1. Induction of pre-diabetes	44
2.3.2. Experimental design	44
2.3.3. Blood collection and tissue harvesting	45
2.4 Biochemical analysis	45
2.5. HOMA2-IR index.....	46
2.6 Statistical analysis.....	46

3. Results	47
3.1. Plasma GIP	47
3.2. Plasma GLP-1	48
3.3. Plasma Insulin.....	49
3.4. Plasma Leptin	50
3.5. Plasma Ghrelin.....	51
3.6. Plasma Glucose.....	52
3.7. Plasma Glycated Hemoglobin (HbA1c).	53
3.8. HOMA2-IR Index.....	54
4. Discussion.....	55
Conclusion.....	57
Acknowledgments	57
Author Contributions	57
Conflict of Interest.....	57
Availability of Data and Materials	57
Ethics Approval and Consent to Participate	57
References	58
Chapter 4: Synthesis and Conclusion	61
Conclusion.....	65
Shortfalls and Future Studies.....	65
References	66
APPENDICES	77
Appendix 1: AREC Ethics Approval Letter	77
Appendix 2: Manuscript 1 journal guide: Canadian Journal of Physiology and Pharmacology	78
Appendix 3: Manuscript 2 Journal guide: Endocrinología, Diabetes y Nutrición.....	90
Appendix 4: Abstract of CHS symposium 2022 oral presentation.....	109

LIST OF FIGURES

Figure	Legend	Page no.
	Chapter 2: Manuscript 1	
Figure 1	Plasma glucose concentrations in the standard and HFHC diet-fed animals for 10 and 20 weeks. Values are presented as mean $\pm$ SEM (n=6). ★ $p < 0.05$ denotes comparison with the standard diet group.	21
Figure 2	Plasma insulin concentrations in the standard and HFHC diet-fed animals for 10 and 20 weeks. Values are presented as mean $\pm$ SEM (n=6). ★ $p < 0.05$ denotes comparison with the standard diet group.	22
Figure 3	Plasma GIP concentrations in the standard and HFHC diet-fed animals for 10 and 20 weeks. Values are presented as mean $\pm$ SEM (n=6). ★ $p < 0.05$ denotes comparison with the standard diet group.	23
Figure 4	Plasma leptin concentrations in the standard and HFHC diet-fed animals for 10 and 20 weeks. Values are presented as mean $\pm$ SEM (n=6). ★ $p < 0.05$ denotes comparison with the standard diet group.	24
Figure 5	Plasma GLP-1 concentrations in the standard and HFHC diet-fed animals for 10 and 20 weeks. Values are presented as mean $\pm$ SEM (n=6). ★ $p < 0.05$ denotes comparison with the standard diet group.	25
Figure 6	Plasma ghrelin concentrations in the standard and HFHC diet-fed animals for 10 and 20 weeks. Values are presented as mean $\pm$ SEM (n=6). ★ $p < 0.05$ denotes comparison with the standard diet group.	26
Figure 7	Plasma HbA _{1c} concentrations in the standard and HFHC diet-fed animals for 10 and 20 weeks. Values are presented as mean $\pm$ SEM (n=6). ★ $p < 0.05$ denotes comparison with the standard diet group.	27
	Chapter 3: Manuscript 2	
Figure 1	Schematic diagram showing the experimental design.	42
Figure 2	Plasma GIP concentrations in non-prediabetic control animals (NPDC), untreated prediabetic control animals (PD), as well as pre-diabetic animals that had a dietary intervention (PD +DI) for 12 weeks. Values are presented as mean $\pm$ SEM (n=6). ★ $p < 0.05$ denotes comparison with NPDC group; # $p < 0.05$ denotes comparison with PD group.	44
Figure 3	Plasma GLP-1 concentrations in non-prediabetic control animals (NPDC), untreated pre-diabetic control animals (PD), as well as pre-diabetic animals that had a dietary intervention (PD +DI) for 12 weeks. Values are presented as mean $\pm$ SEM (n=6). ★ $p < 0.05$ denotes comparison with NPDC group; # $p < 0.05$ denotes comparison with PD group.	45

Figure 4	Plasma insulin concentrations in non-prediabetic control animals (NPDC), untreated prediabetic control animals (PD), as well as pre-diabetic animals that had a dietary intervention (PD +DI) for 12 weeks. Values are presented as mean $\pm$ SEM (n=6). ★ $p < 0.05$ denotes comparison with NPDC group; # $p < 0.05$ denotes comparison with PD group.	46
Figure 5	Plasma leptin concentrations in non-prediabetic control animals (ND), untreated pre-diabetic control animals (PD), as well as pre-diabetic animals that had a dietary intervention (PD +DI) for 12 weeks. The Values are presented as mean $\pm$ SEM (n=6). ★ $p < 0.05$ denotes comparison with NPDC group; # $p < 0.05$ denotes comparison with PD group.	47
Figure 6	Plasma GLP-1 concentrations in non-prediabetic control animals (NPDC), untreated prediabetic control animals (PD), as well as pre-diabetic animals that had a dietary intervention (PD +DI) for 12 weeks. Values are presented as mean $\pm$ SEM (n=6). ★ $p < 0.05$ denotes comparison with NPDC group; # $p < 0.05$ denotes comparison with PD group.	48
Figure 7	Plasma glucose concentrations in non-prediabetic control animals (NPDC), untreated prediabetic control animals (PD), as well as pre-diabetic animals that had a dietary intervention (PD +DI) for 12 weeks. Values are presented as mean $\pm$ SEM (n=6). ★ $p < 0.05$ denotes comparison with NPD group; # $p < 0.05$ denotes comparison with PD group.	49
Figure 8	Plasma HbA1c concentrations in non-prediabetic control animals (NPDC), untreated prediabetic control animals (PD), as well as pre-diabetic animals that had a dietary intervention (PD +DI) for 12 weeks. Values are presented as mean $\pm$ SEM (n=6). ★ $p < 0.05$ denotes comparison with NPD group; # $p < 0.05$ denotes comparison with PD group.	50
	Chapter 4: Synthesis	
Figure 1	Diagram illustrating the role of incretins in developing and reversing pre-diabetes.	62

LIST OF TABLES

Figure	Legend	Page no.
	Chapter 2: Manuscript 1	
Table 1	Displays the HOMA2-IR index calculated at weeks 10 and 20 using the HOMA2 calculator in the NPD and PD groups (n=6 per group).	23
	Chapter 3: Manuscript 2	
Table 1	Displays the HOMA2-IR index calculated at week 32 using the HOMA2 calculator in the NPDC, PD and PD + DI groups (n=6 per group).	51
Table 2	Shows Pearson's correlation between incretins and the HbA1c after 32 weeks.	51

LIST OF ABBREVIATIONS

Abbreviation	Name
ADA	American Diabetes Association
AgRP	Agouti-related protein
BBB	Blood Brain Barrier
BRU	Biomedical Research Unit
cAMP	Cyclic adenosine monophosphate
CART	Cocaine and amphetamine-regulated transcript
CCK	Cholecystokinin
DPP-4	Dipeptidyl-peptidase 4
ELIZA	Enzyme linked immunosorbent assay
GH	Growth hormone
GHS-R	Growth hormone secretagogues receptor
GHSs	Growth hormone secretagogues
GIP	Glucose-dependent insulintropic polypeptide
GLP-1	Glucagon-Like Peptide 1
GLUT-4	Glucose transporter type 4
HbA _{1c}	Glycated hemoglobin
HFHC	High fat and high carbohydrate
HOMA	Homeostatic model assessment
IR	Insulin resistance
LEP-R	Leptin receptor
LH	Lateral hypothalamus
ND	Normal diet (standard diet)
NPD	Non-prediabetes
NPDC	Non-prediabetes control
NPY	Neuropeptide Y
PD	Prediabetes
POMC	Pro-opiomelanocortin
PYY	Peptide YY
SEM	Standard error of mean
T2DM	Type 2 diabetes Mellitus.
UKZN	University of KwaZulu-Natal.
WHO	World Health Organization

LIST OF APPENDICES

Appendix	Name	Page no.
1	AREC Ethics Approval Letter	71
2	Manuscript 1 journal guide: Canadian Journal of Physiology and Pharmacology	72
3	Manuscript 2 Journal guide: Endocrinología, Diabetes y Nutrición	84
4	Abstract of CHS symposium 2022 oral presentation	103

STUDY OUTLINE

This dissertation is presented in a manuscript format consisting of 4 chapters; chapter 1: a literature review. Chapter 2: prologue, abstract and manuscript 1. Chapter 3: prologue, abstract and manuscript 2, chapter 4: synthesis and conclusion. Chapter 1 details the study's introduction in the form of a literature review, the key aims and objectives as well as the justification of the study. Chapter 2 comprises the prologue and abstract of manuscript 1, which further details the link between incretins and the development of prediabetes. In addition, the work in chapter 2 has been submitted for publication and is currently under review in the "Canadian Journal of Physiology and Pharmacology". Chapter 3 presents manuscript 2, which looked at the effect of a low carbohydrate, high unsaturated fat diet on incretin levels on diet-induced prediabetic rats. This work is currently under review in the journal "Endocrinologia, Diabetes y Nutricion". Chapter 4 consists of the synthesis and conclusion. Mzimela N authored the work presented in this study, supervised by Dr A Khathi and Mr. A Sosibo.

ABSTRACT

Background

The increase in the prevalence of type two diabetes mellitus (T2DM) is attributed to unhealthy lifestyles and high-calorie diets. T2DM is a chronic metabolic condition characterised by impaired insulin function and high blood glucose concentration. Prediabetes is an intermediate hyperglycemic condition that frequently occurs before the onset of T2DM. This condition is characterised by a gradual reduction of insulin sensitivity by insulin receptors in insulin-dependent cells, frequently followed by significantly high plasma glucose levels. In this condition, the blood glucose concentration is insufficient to diagnose T2DM. Studies have looked at how incretin peptides affect the pathology of T2DM. However, the link between incretin peptide levels and the onset of prediabetes remains unknown. Additionally, the effect of a low carbohydrate, high unsaturated fat diet on incretin levels during the reversal of prediabetes has not been established. Thus, this study aimed to assess the role of incretin levels in the emergence of prediabetes and the effect of a low carbohydrate, high unsaturated fat diet on incretin levels during the reversal of prediabetes.

Methods

The first study was conducted using 24 male Sprague-Dawley rats, divided into two groups given a standard rat diet (NPD) (n=12), while the other group was given a high-fat high carbohydrate (HFHC) (n=12) diet. Six animals from each group were sacrificed at week 10 and week 20, and blood was collected for biochemical analysis at each time interval. After 20 weeks, the HFHC fed group was found to be prediabetic and were therefore named the prediabetic group (PD). At week 10, the NPD group had the following mean measurements for the NPD and HFHC groups respectively: Glucose (4.3mmol/L and 5.9mmol/L), Insulin (40.26 and 118.32pmol/L), HbAc1 (4.9 and 5.15%), GIP (9.308 and 12.91pmol/L), GLP-1 (18.53 and 15.73pmol/L), Leptin (1.92 and 1.08mmol/L), and Ghrelin (122.1 and 186.5pmol/L). At week 20, the PD group had the following mean measurements for the NPD and PD groups: Glucose (4.4mmol/L and 7.35mmol/L), Insulin (41.18 and 159.42pmol/L), HbAc1 (4.7 and 6.65%), GIP (10.03 and 15.1pmol/L), GLP-1 (21.52 and 6.73pmol/L), Leptin (2.16 and 0.78mmol/L), and Ghrelin (124.2 and 210.63pmol/L).

After 20 weeks of pre-diabetes induction, the second study began with 18 male Sprague-Dawley rats. Group A continued with the standard diet and was used as a non-prediabetic control (NPDC) (n=6). The pre-diabetic group B (n=12) was split into two experimental groups. One of the groups continued a HFHC diet and served as the pre-diabetic control group (PD) (n=6). In contrast, the other group had a diet intervention where the diet was changed to a low carbohydrate-high unsaturated fats diet (PD+DI) (n=6). All groups were then maintained on their respective diets for a further 12 weeks. At week 32, the PD+DI group had the following mean measurements: Glucose (5.367mmol/L), Insulin (188.5ng/ml), HbAc1 (4.62%), GIP (24.08pg/ml), GLP-1, Leptin (1.267ng/ml), and Ghrelin (17.09pg/ml).

Results

In the first study, after 20 weeks, the HFHC diet resulted in moderate hyperglycaemia, elevated plasma insulin, elevated HbA1c and insulin resistance in the PD group compared to the NPD group. There were also significantly increased GIP and ghrelin concentrations with significantly low GLP-1 and leptin concentrations in the PD group compared to the NPD group. Interestingly, at week 10, there was moderate hyperglycaemia, elevated plasma insulin, elevated HbA1c and insulin resistance in the HFHC group. There were also significant GIP and ghrelin levels with significantly low GLP-1 and leptin concentrations, but there was no prediabetes.

In the second study, there were significantly reduced blood glucose levels, plasma insulin levels, HOMA-IR index, and HbA1c in the PD+DI group compared to the PD group. In the PD+DI group, there are significantly reduced GIP and ghrelin levels with significantly increased GLP-1 and leptin concentrations compared to the PD group. However, when the PD-DI group is compared to the NPDC group, there is no significant difference in all measured parameters.

Conclusion

The first study's findings show that chronic ingestion of a HFHC diet causes dysregulation of incretin hormones from week 10, while prediabetes was only diagnosed at week 20. This dysregulation of incretin hormones precedes the onset of prediabetes and may trigger chronic insulin stimulation, leading to prediabetes development. In the second study, we observed the effect of diet on incretins as they play a significant role in developing and reversing pre-diabetes. Chronic consumption of a HFHC diet led to elevated blood glucose and insulin concentration, resulting in abnormal concentrations of incretins. The abnormal incretins then maintained this state of hyperglycemia and hyperinsulinemia, resulting in prediabetes. Chronic consumption of a LCHF by the pre-diabetic rats led to reduced concentrations of incretins, which could have led to a reduced HbA1c and eventually to the reversal of pre-diabetes. The results of this study suggest that incretin concentrations preceded the development of prediabetes and may even have a role in its development as well as its reversal.

Chapter 1: Literature Review

1. Introduction

Glucose is a carbohydrate we get from the food we eat and is used for energy generation (1). Blood glucose concentration is maintained within a homeostatic range via a process known as glucose homeostasis (2). Insulin and glucagon are hormones secreted by islet cells of the pancreas to help maintain the balance of blood glucose concentration within the homeostatic range of 3.5-5.5mmol/L (2).

In the case of increased glucose concentration in a healthy individual above the homeostatic range, insulin is secreted shortly after the ingestion of food (2). It stimulates the uptake of glucose as well as the conversion of glucose into glycogen by the liver for storage (3). Insulin facilitates this process by binding to the insulin receptors on membranes of skeletal muscle and adipose tissue, triggering the exocytosis of glucose transporter type 4 (GLUT4) to the cell membrane (4). The binding of insulin prevents the mobilization of GLUT4; this allows for the removal of glucose from the blood into the cell, which is used for energy generation through glycolysis (4). This exocytosis of GLUT4 in muscle cells is also triggered during exercise resulting in reduced blood glucose levels (5). When the blood glucose concentration falls below the homeostatic range, glucagon is secreted. Once secreted, it binds to the glucagon receptors in the cell membrane (6). This triggers a cascade of reactions that eventually shuts down glycolysis in the adipose tissues and promotes glycogenolysis and gluconeogenesis in the liver cells (7). Glycogen is then converted into glucose and released into the bloodstream, increasing the blood glucose concentration (7).

There are other hormones involved in regulating glucose homeostasis known as incretin hormones. Incretins are hormones secreted in the gut that enhance insulin secretion and contribute significantly to postprandial insulin release (8). The incretin peptides account for up to 60% of postprandial insulin release. The "incretin effect" is a significant factor in maintaining postprandial glucose homeostasis (9). The incretin hormones glucose-dependent insulintropic polypeptide (GIP) and glucagon-like peptide 1 (GLP-1) are associated with appetite-regulating hormones ghrelin and leptin (10, 11). Leptin and ghrelin work closely related to insulin and glucagon to maintain glucose homeostasis (8, 12). A disturbance in glucose homeostasis results in many health problems over time, such as type 2 diabetes mellitus (T2DM) (13). During T2DM, glucose remains in the blood and doesn't enter the cells causing health problems such as fatigue, hyperphagia, and excessive thirst (13). An intermediate hyperglycemic condition known as prediabetes was found to precede the onset of T2DM (14). The disturbances related to insulin during prediabetes are well defined, however, changes in incretin homeostasis are unknown. This raised the question of whether the disturbances in incretin hormones also present and what effect a low carbohydrate, high unsaturated fat diet has on the incretin levels of a prediabetic animal model.

2. Incretin hormones

Incretins are hormones secreted in the gut and aid in maintaining glucose homeostasis by enhancing insulin secretion as they contribute significantly to overall postprandial insulin release (8). The incretin hormones work together to produce the incretin effect, which is a two- to three-fold increase in insulin secretion in response to oral glucose administration compared to intravenous glucose administration (8). Incretin peptides include glucose-dependent insulintropic polypeptide (GIP) and glucagon-like peptide 1 (GLP-1), associated with ghrelin and leptin.

2.1. The Physiological Role of GIP on Glucose Homeostasis

GIP is a 42-amino-acid peptide secreted by the enteroendocrine K-cells in the duodenum and proximal jejunum in response to the presence of glucose in the stomach (15). GIP is an inhibitory peptide that plays a role in insulin secretion via binding to the beta-pancreatic cells (16). The release of insulin into the blood occurs before glucose absorption to prepare for the incoming blood glucose concentration elevation (17).

In response to an empty stomach of a healthy individual, the hormone ghrelin is secreted from the stomach (18). Ghrelin is a gut hormone that induces food ingestion leading to the entrance of glucose in the stomach (18). The enteroendocrine K-cells are stimulated by glucose in the stomach and secrete the glucose-dependent peptide GIP into the blood (17). GIP acts on the G-protein coupled receptors in the beta cells of the pancreatic islets (19). The binding of GIP to its receptor increases the intracellular cyclic adenosine monophosphate (cAMP) concentration, leading to the activation of adenylate cyclase, protein kinase A, and phospholipase C/protein C cascades, respectively (19). This promotes an influx of calcium ions and insulin exocytosis (19). The production of insulin leads to an inhibition of ghrelin and leptin secretion (18) (20). Leptin is a hormone secreted in the gut that increases insulin sensitivity, which aids in energy balance; it suppresses hunger and reduces fat storage in adipocytes (21).

When glucose enters the bloodstream, the beta-pancreatic cells detect increased blood glucose concentration above the homeostatic range and secrete even more insulin to lower blood glucose within the homeostatic range (3). Increasing GIP concentration moderately inhibits gastric acid secretion and gastrointestinal movement, this inhibitory feedback mechanism controls the transit of a meal to optimize nutrient digestion and absorption and is known as the "ileal break" (22). The incretins are rapidly degraded by the ubiquitous enzyme dipeptidyl peptidase 4 (DPP-4) (22). DPP-4 works by cleaving alanine and proline residues in the second position of the N-terminus in peptide chains (19). GIP has a substitution of L-alanine for D-alanine residue at position 2; this makes GIP resistant to the action of DPP-4, enhancing its incretin effect (19). Although GIP is still inactivated, its inactivation occurs slower than GLP-1, giving GIP a half-life of 5 to 7 minutes (19).

2.2. The Physiological Role of GLP-1 on Glucose Homeostasis

Glucagon-like peptide 1 is a 30-amino acid peptide hormone produced by the intestinal epithelial endocrine L-cells (23). GLP-1 is rapidly degraded and inactivated by the enzyme DPP-4 before it leaves the stomach, supporting the notion that GLP-1 activities are communicated via GLP-1 receptor expressing sensory neurons in the intestine and liver (24). The primary functions of GLP-1s are to stimulate insulin secretion (acting as an incretin hormone) and inhibit glucagon secretion, thereby reducing postprandial blood glucose concentration (25). Leptin's interaction with other incretin hormones is needed to stimulate short-term satiation. Leptin has been shown to cause satiation by interacting with GLP-1 and its receptor antagonist (26). Leptin receptors have been discovered in GLP-1-secreting endocrine L cells and neurons, and leptin has been shown to promote GLP-1 release in L cells. GLP-1 plays a vital role in regulating satiety and food intake (26).

The lateral hypothalamus (LH) contains the glucagon-like peptide-1 receptor (GLP-1R), which binds the GLP-1 activating a series of reactions that are required for average body weight, food intake, and food reinforcement management (25). Reduced GLP-1R expression in the LH enhanced weight gain, hyperphagia, and minor glucose tolerance deficits. Like GIP, GLP-1 also functions as an enterogastrone and part of the ileal brake mechanism by inhibiting gastrointestinal motility and secretion (25). GLP-1 secretion is thought to play a role in developing obesity, while exaggerated secretion is believed to cause postprandial reactive hypoglycemia (27). GLP-1 receptor agonists are currently being studied for T2DM due to these activities (27). The stimuli and molecular mechanisms that cause GLP-1 to be produced in response to meal ingestion are still being explored.

2.3. The Physiological Role of Ghrelin on Glucose Homeostasis

Ghrelin is a 28-amino-acid peptide that was discovered to be the first circulating hunger hormone (28, 29). The ghrelin receptor, also known as the growth hormone secretagogue receptor (GHS-R), is a G protein-coupled receptor that binds growth hormone secretagogues (GHSs), such as ghrelin (30). Ghrelin acts in the brain to regulate food intake, body weight, adiposity, and glucose metabolism. GHSR is thought to play a role in maintaining energy homeostasis and body weight (31). Ghrelin concentration in the blood rises during fasting and falls during feeding. Ghrelin is primarily synthesized in the stomach and increases growth hormone production (GH) (32). A-like cells in the stomach's oxyntic glands generate and secrete this peptide hormone's octanoylated form (Acyl ghrelin) (32). GHSR1a and 1b receptors are widely distributed and can be detected in various body and brain regions, including tumours and metastasis (33). This wide distribution of ghrelin and its receptors means that ghrelin plays many physiological roles.

In the hypothalamus, particularly in the arcuate nucleus, there are appetite stimulator neurons, namely neuropeptide Y (NPY), that promote food intake and Agouti-related protein (AgRP); which prolongs feeding (32). During a hypoglycaemic state, ghrelin is secreted from the stomach, increasing the circulating peripheral ghrelin that directly crosses the brain-blood barriers as a saturable transport

system and acts on the hypothalamic arcuate nucleus to activate and increase the synthesis of NPY and AgRP neurons, thus promoting an increased appetite and food intake and overall increase in blood glucose (31).

An increased insulin secretion inactivates the AgRP neurons due to increased blood glucose simultaneously with an increased leptin secretion (34). The capacity of ghrelin-producing cells to perceive foods directly or gut hormones (insulin, glucagon-like-peptide 1 (GLP-1), peptide YY (PYY), and cholecystokinin (CCK) released following a meal may be one proposed mechanism for suppressing ghrelin production by food (33). Ghrelin has been found to inhibit glucose-stimulated insulin release, while insulin has been shown to reduce ghrelin secretion (18).

2.4. The Physiological Role of Leptin on Glucose Homeostasis

Leptin is a 167-amino-acid protein produced predominantly by white adipose tissue and absorbed into circulation. Leptin is secreted after an increased insulin secretion in a hyperglycaemic state and increases insulin sensitivity by activating the insulin receptor substrate-1 found in the brain (20, 35, 36). Leptin aids in energy balance by suppressing hunger and reducing fat storage in adipocytes (21). Longform leptin receptor-b (LEPR-b) is found in both the peripheral and central nervous systems, with the hypothalamus on the arcuate nucleus (ARC) being the primary site of expression (32). The ARC is the first-order centre that senses peripheral metabolic signals and other brain areas (32).

After increasing insulin secretion, leptin is secreted by white adipose tissues and absorbed into the circulation (37). In the hypothalamus, it crosses the brain-blood barriers as a saturable transport to the hypothalamic arcuate nucleus, where it binds to LEPR-b (37). The binding of leptin stimulates neurons in the ARC, and the stimulated neurons, such as pro-opiomelanocortin (POMC), cocaine and amphetamine-regulated transcript (CART) neurons, operate as appetite inhibitors that increase satiety (20, 35, 36). When leptin acts on the hypothalamic neurons, it has been found to deactivate the NPY and AgRP appetite stimulator neurons and therefore suppress appetite and food intake, inducing satiety (37). The incretin hormones' mechanisms of action are disturbed in conditions such as T2DM (20, 38) (39). The changes to incretins in T2DM are well-defined; however, the changes during prediabetes are still unknown.

3. Type 2 Diabetes Mellitus

Type 2 diabetes mellitus is a metabolic disorder characterised by hyperglycemia due to insulin resistance (40). This can be due to resistance of insulin receptors developed over time, mainly from an inactive lifestyle and diet high in fats and carbohydrates (41). Insulin resistance refers to a condition whereby insulin is secreted into the blood but cannot effectively bind to its receptors on the cell membranes.

The ineffective binding of insulin means glucose cannot be allowed inside cells because the GLUT4, which allows glucose in the cell, will not be embedded in the plasma membrane (42). An oral glucose tolerance test (OGTT), a fasting glucose test, a postprandial glucose test, or glycated haemoglobin (HbA1c) test can all be used to diagnose T2DM (43). When an individual's fasting blood glucose concentration is greater than 7 mmol/L with postprandial glucose concentration greater than 11.1 mmol/L and glycated haemoglobin concentration greater than 6.5 percent, that individual is diagnosed with T2DM (44). T2DM is associated with several microvascular and macrovascular complications, such as renal failure, heart disease and bone weakness (45, 46).

3.1 Changes in GIP levels in T2DM

Due to hyperglycemia, there are high concentrations of the hormone gastric inhibitory peptide since it's a glucose-dependent peptide (38). GIP secretion is triggered after a meal because of the increase in blood glucose concentration, but it is secreted constantly in T2DM due to a high blood glucose concentration (38). If a person with diabetes mellitus consumes food high in carbohydrate and fats content, their blood glucose, together with GIP and glucagon-like peptides, will further increase, inhibiting gastric acid secretion and absorption in the small intestines and reducing the gut motility (47). Due to the Inhibition of gastric emptying and gastric acid secretions, there are low motilin and ghrelin concentration. These hormones help induce muscle peristalsis in the digestive tract and signal hunger, which increases gut motility (48, 49). This blockage might lead to bloating and constipation.

The reduced gut motility results in a build-up of food in the small intestines, and the Inhibition of absorption results in an overgrowth of bacteria in the small intestines (49). The excess bacteria trigger an increase in zonulin concentration, a protein that regulates the permeability of the tight junctions between cells in the digestive tract wall and increases the washout of bacteria (50). The washout of the excess bacteria causes pain and leads to diarrhea. The increase in zonulin secretion increases the size and permeability of the tight junctions, allowing foreign entities like bacteria and digested food particles to enter the extracellular membrane, causing inflammation, and leading to the immune changes observed in T2DM (50). The Increased concentration of GIP is responsible for the irregularities observed in T2DM.

3.2. Changes in GLP-1 levels in T2DM

GLP-1 (glucagon-like peptide-1) is an incretin hormone that causes glucose-dependent insulin production, glucagon suppression, delayed stomach emptying, and reduced caloric intake (25). In T2DM, DPP4 expression is elevated during adipose tissue differentiation and is linked to an increased secretion rate from adipose tissue in obesity (BMI) (39). The pathway of GLP-1 is disturbed in T2DM; GLP-1 is being actively degraded by the increased circulating DPP-4 enzyme as fast as it is secreted, meaning it brings about no significant changes (39).

Oral glucose stimulates the release of GLP-1, whereas intravenously delivered glucose does not affect plasma GLP-1 concentrations (51). Glucagon-like peptide-1 is a glucose-lowering peptide that provides the basis for two new types of glucose-lowering drugs for type 2 diabetic individuals: incretin mimetics and inhibitors of protease DPP-4 (27).

3.3. Ghrelin and Leptin levels in T2DM

During a hypoglycaemic state or when the stomach is empty, ghrelin is secreted into the blood, increasing the circulating peripheral ghrelin that directly crosses the brain-blood barriers as a saturable transport system (52). Ghrelin binds to the GHS-R and therefore activates the hypothalamic arcuate nucleus, which increases the synthesis of NPY and AgRP neurons, promoting an increased appetite and food intake (31). After a meal, increased blood glucose stimulates insulin secretion from the pancreatic beta cells (53). As mentioned above, leptin increases after insulin secretion, secreted by white adipose tissues, increases circulation, and crosses the brain-blood barriers as a saturable transport to the hypothalamic arcuate nucleus, where it binds leptin receptor-b (LEPR-b) (37). Its binding stimulates the synthesis of appetite inhibitors and increases its synthesis, which reduces appetite and food intake by activating the satiety centre (Ventromedial hypothalamus) (20, 54). Giving an individual a feeling of satisfaction by reducing appetite and food intake reduces ghrelin secretion after a meal, thus regulating glucose homeostasis (55).

However, in T2DM, this pathway is impaired due to hyperlipidaemia and hyperinsulinemia; leptin resistance occurs because of increased leptin concentration for an extended period (20). Due to insulin resistance, the cells cannot access glucose from the blood, and they constantly transmit signals to the hypothalamus to increase food intake (56). The excitation of the vagus nerve stimulates ghrelin secretion in the stomach, increasing appetite and food intake (57). Due to leptin resistance, satiety is not induced, and appetite and food intake are not inhibited, so the individual is constantly hungry (20).

A condition known as prediabetes has been discovered to precede T2DM (58). The main difference between T2DM and prediabetes is that T2DM has a higher blood glucose concentration, meaning its effects are more intense than prediabetes (58). This raised whether the complications mentioned above brought upon the incretins, ghrelin and leptin are also present in the prediabetic state.

4. Prediabetes

Prediabetes is a high-risk state characterised by intermediate hyperglycaemia (59). It occurs when the blood glucose is higher than normal but it's not high enough to diagnose type 2 diabetes mellitus (59). According to studies, prolonged consumption of food high in carbohydrates and fat content is linked to an increased risk of developing prediabetes (60). Other risk factors for prediabetes are inactive lifestyles, lack of exercise, obesity, and family history (59, 61). Long durations of inactivity can impair normal blood glucose homeostasis, resulting in obesity, cardiovascular disease, and the risk of prediabetes (61). Individuals at risk of developing T2DM have one or more prediabetic conditions, such as impaired

fasting glucose (IFG) of 5.5- 6.9 mmol/L, impaired glucose tolerance (IGT) of 7.8-11.1 mmol/L, and impaired glycated haemoglobin (Hb1Ac) concentration of 5.7-6.4 percent (62). Glycated haemoglobin determines how much haemoglobin has become bound to glucose in the past months (63).

A study to observe the changes in immune cell concentration during the progression of HFHC diet-induced prediabetes was conducted in our lab, using male rats to mimic the human conditions (64). They were fed a high-fat, high-carbohydrate diet for 20 weeks to induce prediabetes. Results were collected using blood samples after 12 weeks, and proof of inflammation was confirmed by decreasing blood neutrophils and eosinophils (64). However, this study focused solely on changes in immune cell concentration during prediabetes. Pathophysiologies in the small intestine, such as celiac disease and chronic inflammation, undoubtedly contribute to immune changes during prediabetes.

5. Management of T2DM and prediabetes

As the prevalence of T2DM rises worldwide, it's now more important to prevent prediabetes from progressing to T2DM. A large portion of prediabetic individuals progresses toward the development of T2DM because they are unaware that they have it (65). Type 2 diabetes mellitus has no known cure; it can be kept under control with exercise, a change in diet and medication (66). The association between incretins on T2DM has been established, and several studies have been conducted to check their effects on T2DM. However, this is not known for the prediabetic state. This raises the question of whether the complications associated with the incretins experienced in T2DM begin in the prediabetic state. This work is significant because if the incretins are linked to the development of prediabetes, the findings could develop a new marker for detecting prediabetes. The findings of this study could also help reverse prediabetes and prevent T2DM.

6. HFHC animal model of prediabetes

The average person's diet is high in saturated fats and carbohydrates, with little in the way of vegetables and fruits. As a result, the HFHC diet supplemented with 15% fructose is used in animal models to mimic the average person's diet (67). According to studies, the combined effect of an HFHC diet causes the most severe symptoms, including hyperglycaemia, hypercholesterolemia, and increased inflammatory mediators (41). Additionally, daily consumption of sugars, refined high-calorie foods, high saturated fats, and high-sweetened refined sugars has been linked to the development of prediabetes in studies (41). The prediabetes symptoms in individuals are best described using this diet-induced prediabetic animal model (67). Since the concentration of the incretin hormones is affected by diet, changing from a HFHC diet to a healthy diet could reduce the GIP. Therefore, the insulin in the blood could also be reduced, which could reverse prediabetes.

7. Aim

This study sought to investigate the association of incretins, ghrelin & leptin in the development of diet-induced prediabetes as well as look at their role in the reversal of prediabetes.

8. Objectives

8.1. Manuscript 1.

To assess the association between incretin peptides levels and the development of prediabetes by investigating the following:

- Glycated haemoglobin concentration
- Fasting Blood Glucose
- Insulin concentration
- GIP concentration
- GLP-1 concentration
- Ghrelin concentration
- Leptin concentration

8.2. Manuscript 2.

To explore the effects of a low carbohydrate, high unsaturated fat diet in incretin peptides levels, to attempt to reverse prediabetes and its effects by investigating the following:

- GIP concentration
- GLP-1 concentration
- Ghrelin concentration
- Leptin concentration
- Insulin concentration
- Glycated haemoglobin concentration

References

1. Güemes M, Rahman SA, Hussain K. What is a normal blood glucose? Archives of disease in childhood. 2016;101(6):569-74.
2. Holst JJ, Holland W, Gromada J, Lee Y, Unger RH, Yan H, et al. Insulin and glucagon: partners for life. Endocrinology. 2017;158(4):696-701.
3. Qaid MM, Abdelrahman MM. Role of insulin and other related hormones in energy metabolism—A review. Cogent Food & Agriculture. 2016;2(1):1267691.
4. Wang T, Wang J, Hu X, Huang X-J, Chen G-X. Current understanding of glucose transporter 4 expression and functional mechanisms. World journal of biological chemistry. 2020;11(3):76.
5. Klip A, McGraw TE, James DE. Thirty sweet years of GLUT4. Journal of Biological Chemistry. 2019;294(30):11369-81.
6. Tengholm A, Gylfe E. cAMP signalling in insulin and glucagon secretion. Diabetes, obesity and metabolism. 2017;19:42-53.
7. Abdulkader F. Metabolic Pathways and Cell Signaling. Essential Aspects of Immunometabolism in Health and Disease: Springer; 2022. p. 13-30.
8. Wu T, Rayner CK, Horowitz M. Incretins. Metabolic Control. 2015:137-71.
9. Frost F, Jones GH, Dyce P, Jackson V, Nazareth D, Walshaw MJ. Loss of incretin effect contributes to postprandial hyperglycaemia in cystic fibrosis-related diabetes. Diabetic Medicine. 2019;36(11):1367-74.
10. Lim GE, Brubaker PL. Glucagon-Like Peptide 1 Secretion by the L-Cell. Diabetes. 2006;55(Supplement 2):S70.
11. Ronveaux CC, Tomé D, Raybould HE. Glucagon-like peptide 1 interacts with ghrelin and leptin to regulate glucose metabolism and food intake through vagal afferent neuron signaling. J Nutr. 2015;145(4):672-80.
12. Skuratovskaia D, Vulf M, Chasovskikh N, Komar A, Kirienkova E, Shunkin E, et al. The links of ghrelin to incretins, insulin, glucagon, and leptin after bariatric surgery. Frontiers in genetics. 2021;12.
13. Rocha S, Corvo ML, Fernandes E, Freitas M. The Emerging Target Protein Tyrosine Phosphatase 1B (PTP1B) for Type 2 Diabetes Mellitus Management. 2021.
14. Mkhize B, Mosili P, Ngubane P, Sibiyi N, Khathi A. Diet-induced prediabetes: Effects on the systemic and renal renin-angiotensin-aldosterone system. 2020.
15. Lynggaard MB, Gasbjerg LS, Christensen MB, Knop FK. GIP (3-30) NH₂—a tool for the study of GIP physiology. Current Opinion in Pharmacology. 2020;55:31-40.

16. Baggio LL, Drucker DJ. Biology of incretins: GLP-1 and GIP. *Gastroenterology*. 2007;132(6):2131-57.
17. Joo E, Harada N, Yamane S, Fukushima T, Taura D, Iwasaki K, et al. Inhibition of gastric inhibitory polypeptide receptor signaling in adipose tissue reduces insulin resistance and hepatic steatosis in high-fat diet-fed mice. *Diabetes*. 2017;66(4):868-79.
18. Sun Y, Asnicar M, Saha PK, Chan L, Smith RG. Ablation of ghrelin improves the diabetic but not obese phenotype of ob/ob mice. *Cell metabolism*. 2006;3(5):379-86.
19. Gupta K, Raja A. Physiology, Gastric Inhibitory Peptide. *StatPearls [Internet]: StatPearls Publishing*; 2020.
20. Zhou Y, Rui L. Leptin signaling and leptin resistance. *Frontiers of medicine*. 2013;7(2):207-22.
21. Izadi V, Saraf-Bank S, Azadbakht L. Dietary intakes and leptin concentrations. *ARYA Atheroscler*. 2014;10(5):266-72.
22. Kolodziejski PA, Sassek M, Chalupka D, Leciejewska N, Nogowski L, Mackowiak P, et al. GLP1 and GIP are involved in the action of synbiotics in broiler chickens. *Journal of animal science and biotechnology*. 2018;9(1):1-9.
23. Lim GE, Brubaker PL. Glucagon-Like Peptide 1 Secretion by the L-Cell. *The View From Within*. 2006;55(Supplement 2):S70-S7.
24. Zhang Y, Liu Y, Xu J, Sun Q, Yu F, Cheng J, et al. Inhibition of DPP4 enhances inhibitory synaptic transmission through activating the GLP-1/GLP-1R signaling pathway in a rat model of febrile seizures. *Biochemical Pharmacology*. 2018;156:78-85.
25. Müller TD, Finan B, Bloom SR, D'Alessio D, Drucker DJ, Flatt PR, et al. Glucagonlike peptide 1 (GLP-1). *Molecular Metabolism*. 2019;30:72-130.
26. Ronveaux CC, Tomé D, Raybould HE. Glucagon-like peptide 1 interacts with ghrelin and leptin to regulate glucose metabolism and food intake through vagal afferent neuron signaling. *The Journal of nutrition*. 2015;145(4):672-80.
27. Andersen A, Lund A, Knop FK, Vilsbøll T. Glucagon-like peptide 1 in health and disease. *Nature Reviews Endocrinology*. 2018;14(7):390-403.
28. Hosoda H, Kojima M, Matsuo H, Kangawa K. Ghrelin and des-acyl ghrelin: two major forms of rat ghrelin peptide in gastrointestinal tissue. *Biochemical and biophysical research communications*. 2000;279(3):909-13.
29. Lv Y, Liang T, Wang G, Li Z. Ghrelin, a gastrointestinal hormone, regulates energy balance and lipid metabolism. *Bioscience reports*. 2018;38(5).

30. Poher A-L, Tschöp MH, Müller TD. Ghrelin regulation of glucose metabolism. *Peptides*. 2018;100:236-42.
31. Sun Y, Wang P, Zheng H, Smith RG. Ghrelin stimulation of growth hormone release and appetite is mediated through the growth hormone secretagogue receptor. *Proceedings of the National Academy of Sciences*. 2004;101(13):4679-84.
32. Suyama S, Yada T. New insight into GABAergic neurons in the hypothalamic feeding regulation. *The Journal of Physiological Sciences*. 2018;68(6):717-22.
33. Akalu Y, Molla MD, Dessie G, Ayelign B. Physiological effect of ghrelin on body systems. *International journal of endocrinology*. 2020;2020.
34. Nakazato M, Murakami N, Date Y, Kojima M, Matsuo H, Kangawa K, et al. A role for ghrelin in the central regulation of feeding. *Nature*. 2001;409(6817):194-8.
35. Blázquez E, Velázquez E, Hurtado-Carneiro V, Ruiz-Albusac JM. Insulin in the brain: its pathophysiological implications for States related with central insulin resistance, type 2 diabetes and Alzheimer's disease. *Frontiers in endocrinology*. 2014;5:161.
36. Satoh N, Ogawa Y, Katsuura G, Tsuji T, Masuzaki H, Hiraoka J, et al. Pathophysiological significance of the obese gene product, leptin, in ventromedial hypothalamus (VMH)-lesioned rats: evidence for loss of its satiety effect in VMH-lesioned rats. *Endocrinology*. 1997;138(3):947-54.
37. Park H-K, Ahima RS. Leptin signaling. *F1000Prime Rep*. 2014;6:73-.
38. Irwin N, Gault VA, O'Harte FP, Flatt PR. Blockade of gastric inhibitory polypeptide (GIP) action as a novel means of countering insulin resistance in the treatment of obesitydiabetes. *Peptides*. 2020;125:170203.
39. Sarkar J, Nargis T, Tandia O, Ghosh S, Chakrabarti P. Increased Plasma Dipeptidyl Peptidase-4 (DPP4) Activity Is an Obesity-Independent Parameter for Glycemic Deregulation in Type 2 Diabetes Patients. *Front Endocrinol (Lausanne)*. 2019;10:505-.
40. David L. 2019 ESC Guidelines on diabetes, pre-diabetes, and cardiovascular diseases developed in collaboration with the EASD. *European heart journal*. 2020;41(2):255-323.
41. DeFronzo RA, Ferrannini E, Groop L, Henry RR, Herman WH, Holst JJ, et al. Type 2 diabetes mellitus. *Nature reviews Disease primers*. 2015;1(1):1-22.
42. Freeman AM, Pennings N. Insulin resistance. *StatPearls [Internet]*. 2021.
43. Sultan E, Taha I, Saber LM. Altered bone metabolic markers in type 2 diabetes mellitus: impact of glycemic control. *Journal of Taibah University Medical Sciences*. 2008;3(2):104-16.
44. Rydén L, Grant PJ, Anker SD, Berne C, Cosentino F, Danchin N, et al. Erratum: ESC Guidelines on diabetes, pre-diabetes, and cardiovascular diseases

- developed in collaboration with the EASD (European Heart Journal (2013) 34 (3035-3087. European Heart Journal. 2014;35(27):1824.
45. Khanduker S, Ahmed R, Khondker F, Aharama A, Afrose N, Chowdhury MAA. Electrolyte Disturbances in Patients with Diabetes Mellitus. Bangladesh Journal of Medical Biochemistry. 2017;10(1):27-35.
 46. Cosentino F, Grant PJ, Aboyans V, Bailey CJ, Ceriello A, Delgado V, et al. 2019 ESC Guidelines on diabetes, pre-diabetes, and cardiovascular diseases developed in collaboration with the EASD: The Task Force for diabetes, pre-diabetes, and cardiovascular diseases of the European Society of Cardiology (ESC) and the European Association for the Study of Diabetes (EASD). European heart journal. 2020;41(2):255-323.
 47. Pederson RA, McIntosh CH. Discovery of gastric inhibitory polypeptide and its subsequent fate: Personal reflections. J Diabetes Investig. 2016;7 Suppl 1(Suppl 1):4-7.
 48. Deloose E, Verbeure W, Depoortere I, Tack J. Motilin: from gastric motility stimulation to hunger signalling. Nat Rev Endocrinol. 2019;15(4):238-50.
 49. Mani BK, Zigman JM. Ghrelin as a Survival Hormone. Trends Endocrinol Metab. 2017;28(12):843-54.
 50. Fasano A. Zonulin, regulation of tight junctions, and autoimmune diseases. Ann N Y Acad Sci. 2012;1258(1):25-33.
 51. Bistola V, Lambadiari V, Dimitriadis G, Ioannidis I, Makrilakis K, Tentolouris N, et al. Possible mechanisms of direct cardiovascular impact of GLP-1 agonists and DPP4 inhibitors. Heart Failure Reviews. 2018;23(3):377-88.
 52. Korbonits M, Goldstone AP, Gueorguiev M, Grossman AB. Ghrelin—a hormone with multiple functions. Frontiers in Neuroendocrinology. 2004;25(1):27-68.
 53. Yaney G, Corkey B. Fatty acid metabolism and insulin secretion in pancreatic beta cells. Diabetologia. 2003;46(10):1297-312.
 54. Meier U, Gressner AM. Endocrine Regulation of Energy Metabolism: Review of Pathobiochemical and Clinical Chemical Aspects of Leptin, Ghrelin, Adiponectin, and Resistin. Clinical Chemistry. 2004;50(9):1511-25.
 55. Satoh N, Ogawa Y, Katsuura G, Tsuji T, Masuzaki H, Hiraoka J, et al. Pathophysiological Significance of the Obese Gene Product, Leptin, in Ventromedial Hypothalamus (VMH)-Lesioned Rats: Evidence for Loss of Its Satiety Effect in VMH-Lesioned Rats★. Endocrinology. 1997;138(3):947-54.
 56. Adamska E, Ostrowska L, Górska M, Krętowski A. The role of gastrointestinal hormones in the pathogenesis of obesity and type 2 diabetes. Prz Gastroenterol. 2014;9(2):6976.

57. Ibrahim Abdalla MM. Ghrelin - Physiological Functions and Regulation. *Eur Endocrinol.* 2015;11(2):90-5.
58. Tabák AG, Herder C, Rathmann W, Brunner EJ, Kivimäki M. Prediabetes: a high-risk state for diabetes development. *Lancet.* 2012;379(9833):2279-90.
59. Andes LJ, Cheng YJ, Rolka DB, Gregg EW, Imperatore G. Prevalence of prediabetes among adolescents and young adults in the United States, 2005-2016. *JAMA pediatrics.* 2020;174(2):e194498-e.
60. Punthakee Z, Goldenberg R, Katz P. Definition, classification and diagnosis of diabetes, prediabetes and metabolic syndrome. *Canadian journal of diabetes.* 2018;42:S10-S5.
61. Tabák AG, Herder C, Rathmann W, Brunner EJ, Kivimäki M. Prediabetes: a high-risk state for diabetes development. *The Lancet.* 2012;379(9833):2279-90.
62. Stefanaki C, Bacopoulou F, Peppas M. Prediabetes and adolescence—trends, causes, effects, and screening. *US Endocrinol.* 2016;12(2):94-8.
63. Sharma P, Panchal A, Yadav N, Narang J. Analytical techniques for the detection of glycated haemoglobin underlining the sensors. *International Journal of Biological Macromolecules.* 2020;155:685-96.
64. Mzimela NC, Ngubane PS, Khathi A. The changes in immune cell concentration during the progression of pre-diabetes to type 2 diabetes in a high-fat high-carbohydrate diet-induced pre-diabetic rat model. *Autoimmunity.* 2019;52(1):27-36.
65. Khan RMM, Chua ZJY, Tan JC, Yang Y, Liao Z, Zhao Y. From Pre-Diabetes to Diabetes: Diagnosis, Treatments and Translational Research. *Medicina (Kaunas).* 2019;55(9).
66. Marín-Peñalver JJ, Martín-Timón I, Sevillano-Collantes C, Del Cañizo-Gómez FJ. Update on the treatment of type 2 diabetes mellitus. *World J Diabetes.* 2016;7(17):354-95.
67. Khathi A, Luvuno M, Mabandla M. VOLUNTARY INGESTION OF A HIGH-FAT HIGH-CARBOHYDRATE DIET: A MODEL FOR PREDIABETES. *PONTE International Scientific Researchs Journal.* 2018;74.
68. Oguntibeju OO. Type 2 diabetes mellitus, oxidative stress and inflammation: examining the links. *Int J Physiol Pathophysiol Pharmacol.* 2019;11(3):45-63.
69. Zamora-Kapoor A, Fyfe-Johnson A, Omidpanah A, Buchwald D, Sinclair Ki. Risk factors for pre-diabetes and diabetes in adolescence and their variability by race and ethnicity. *Prev Med.* 2018;115:47-52.
70. Hostalek U. Global epidemiology of prediabetes - present and future perspectives. *Clin Diabetes Endocrinol.* 2019;5:5.

71. Joo E, Harada N, Yamane S, Fukushima T, Taura D, Iwasaki K, et al. Inhibition of Gastric Inhibitory Polypeptide Receptor Signaling in Adipose Tissue Reduces Insulin Resistance and Hepatic Steatosis in High-Fat Diet–Fed Mice. *Diabetes*. 2017;66(4):868.
72. Van Doorn C, Macht VA, Grillo CA, Reagan LP. Leptin resistance and hippocampal behavioral deficits. *Physiology & behavior*. 2017;176:207-13.
73. Nauck MA, Quast DR, Wefers J, Meier JJ. GLP-1 receptor agonists in the treatment of type 2 diabetes – state-of-the-art. *Molecular Metabolism*. 2021;46:101102.
74. Fu Y, Kaneko K, Lin H-Y, Mo Q, Xu Y, Suganami T, et al. Gut Hormone GIP Induces Inflammation and Insulin Resistance in the Hypothalamus. *Endocrinology*. 2020;161(9).
75. Shankar K, Takemi S, Gupta D, Varshney S, Mani BK, Osborne-Lawrence S, et al. Ghrelin cell-expressed insulin receptors mediate meal- and obesity-induced declines in plasma ghrelin. *JCI Insight*. 2021;6(18).
76. Omran DM, Alaraji SM, Albayati AH, Essam W. Relationship between Ghrelin and Leptin with Insulin Resistance in Obese Patients and Non-Obese Individuals. *Research Journal of Pharmacy and Technology*. 2018;11(1):281-3.
77. Papatheodorou K, Banach M, Bekiari E, Rizzo M, Edmonds M. Complications of Diabetes 2017. *Journal of Diabetes Research*. 2018;2018:3086167.
78. Abdul-Ghani MA, DeFronzo RA. Pathophysiology of prediabetes. *Current Diabetes Reports*. 2009;9(3):193-9.
79. Ahren B. DPP-4 inhibitors. *Best Practice & Research Clinical Endocrinology & Metabolism*. 2007;21(4):517-33.
80. Ha K, Joung H, Song Y. Inadequate fat or carbohydrate intake was associated with an increased incidence of type 2 diabetes mellitus in Korean adults: a 12-year community-based prospective cohort study. *Diabetes research and clinical practice*. 2019;148:254-61.
81. Gamede M, Mabuza L, Ngubane P, Khathi A. The effects of plant-derived oleanolic acid on selected parameters of glucose homeostasis in a diet-induced pre-diabetic rat model. *Molecules*. 2018;23(4):794.
82. Luvuno M, Khathi A, Mabandla MV. Diet-induced prediabetes: effects of exercise treatment on risk factors for cardiovascular complications. *Nutrition & Metabolism*. 2021;18(1):1-9.
83. Sosibo AM. Investigating changes to the insulin signalling pathway in a diet-induced pre-diabetic rat model: effects on selected markers 2019.

84. Bano G. Glucose homeostasis, obesity and diabetes. *Best Practice & Research Clinical Obstetrics & Gynaecology*. 2013;27(5):715-26.
85. Freeman AM, Pennings N. Insulin resistance. *StatPearls* [Internet]: StatPearls Publishing; 2022.
86. Kostov K, Blazhev A. Use of glycated hemoglobin (A1c) as a biomarker for vascular risk in type 2 diabetes: Its relationship with matrix metalloproteinases-2,-9 and the metabolism of collagen IV and elastin. *Medicina*. 2020;56(5):231.
87. Ahmed MH, Ghatge MS, Safo MK. Hemoglobin: structure, function and allostery. *Subcellular biochemistry*. 2020;94:345.
88. Sacks DB, Bebu I, Lachin JM. Refining measurement of hemoglobin A1c. *Clinical chemistry*. 2017;63(9):1433-5.
89. Wang C, Ye Y, Sun W, Yu J, Wang J, Lawrence DS, et al. Red blood cells for glucoseresponsive insulin delivery. *Advanced Materials*. 2017;29(18):1606617.
90. Sherwani SI, Khan HA, Ekhzaimy A, Masood A, Sakharkar MK. Significance of HbA1c test in diagnosis and prognosis of diabetic patients. *Biomarker insights*. 2016;11:BMI. S38440.
91. Renz PB, Cavagnoli G, Weinert LS, Silveiro SP, Camargo JL. HbA1c test as a tool in the diagnosis of gestational diabetes mellitus. *PLoS One*. 2015;10(8):e0135989.
92. El K, Campbell JE. The role of GIP in α -cells and glucagon secretion. *Peptides*. 2020;125:170213.
93. Samms RJ, Coghlan MP, Sloop KW. How may GIP enhance the therapeutic efficacy of GLP-1? *Trends in Endocrinology & Metabolism*. 2020;31(6):410-21.
94. Petersen MC, Shulman GI. Mechanisms of insulin action and insulin resistance. *Physiological reviews*. 2018;98(4):2133-223.
95. Fu Y, Kaneko K, Lin H-Y, Mo Q, Xu Y, Suganami T, et al. Gut hormone GIP induces inflammation and insulin resistance in the hypothalamus. *Endocrinology*. 2020;161(9):bqaa102.
96. Kanemaru Y, Harada N, Shimazu-Kuwahara S, Yamane S, Ikeguchi E, Murata Y, et al. Absence of GIP secretion alleviates age-related obesity and insulin resistance. *Journal of Endocrinology*. 2020;245(1):13-20.
97. D'souza AM, Neumann UH, Glavas MM, Kieffer TJ. The glucoregulatory actions of leptin. *Molecular metabolism*. 2017;6(9):1052-65.
98. Obradovic M, Sudar-Milovanovic E, Soskic S, Essack M, Arya S, Stewart AJ, et al. Leptin and Obesity: Role and Clinical Implication. *Frontiers in Endocrinology*. 2021;12.

99. Paz-Filho G, Mastronardi C, Wong ML, Licinio J. Leptin therapy, insulin sensitivity, and glucose homeostasis. *Indian J Endocrinol Metab.* 2012;16(Suppl 3):S549-55.
100. Wang G, Liu X, Christoffel KK, Zhang S, Wang B, Liu R, et al. Prediabetes is not all about obesity: association between plasma leptin and prediabetes in lean rural Chinese adults. *European Journal of Endocrinology.* 2010;163(2):243-9.
101. Hira T, Pinyo J, Hara H. What is GLP-1 really doing in obesity? *Trends in Endocrinology & Metabolism.* 2020;31(2):71-80.

Chapter 2: Manuscript 1

Investigating the effect of incretin peptide levels in the development of diet-induced prediabetes.

Prologue 1

Chronic consumption of high-caloric diets has been associated with the development of a chronic metabolic condition known as type 2 diabetes (T2DM). T2DM is preceded by prediabetes which is a state of intermediate hyperglycaemia. Up to 70% of people with prediabetes will, in the long run, develop T2DM. Incretins have been shown to contribute to the progression from prediabetes to T2DM however the role of incretin concentrations in the development of T2DM has not been elucidated. Hence the study sought to assess the link between incretin concentrations and the development of prediabetes using our laboratory developed HFHC diet-induced prediabetes animal model.

The manuscript in chapter 2 titled **“Investigating the effect of incretin peptide levels in the development of diet-induced prediabetes”**. is authored by N. Mzimela, NR Dimba, A.M Sosibo, P.S Ngubane and A Khathi.

The manuscript is currently under review in the Canadian Journal of Physiology and Pharmacology.

Investigating the effect of incretin peptide levels in the development of diet-induced prediabetes.

Nhlakanipho Mzimela¹★, Nosipho R. Dimba¹, Aubrey M. Sosibo¹, Phikelelani S. Ngubane¹, and

Andile Khathi¹

¹ School of Laboratory Medicine and Medical Sciences, Discipline of Human Physiology, University of KwaZulu-Natal, South Africa

★Correspondence: 218006756@stu.ukzn.ac.za (Nhlakanipho Mzimela)

Abstract

Prediabetes is a chronic metabolic condition characterised by a gradual reduction of insulin sensitivity by insulin receptors in insulin-dependent cells, frequently followed by increased plasma glucose levels, which are still below the threshold for a type 2 diabetes mellitus diagnosis. Studies already conducted have looked at how incretin peptides affect the pathology of type 2 diabetes mellitus. On the other hand, the link between incretin peptide levels and the onset of prediabetes remains unknown. Thus, this study aimed to assess the role that incretin peptide levels play in the emergence of prediabetes. This study was conducted using 24 male Sprague-Dawley rats, divided into two groups given a standard rat diet (NPD) (=12), while the other group was given a high-fat high carbohydrate (HFHC) (n=12) diet. Six animals from each group were sacrificed at week 10 and week 20, and blood was collected for biochemical analysis at each time interval. The chronic ingestion of a HFHC diet for 10 weeks resulted in significantly increased GIP ($p=0.0001$), GLP-1 ($p=0.0019$), leptin ($p=0.0001$) and ghrelin ($p=0.0001$) concentrations. After 20 weeks, there were further increased incretin hormone concentrations; there were significantly higher concentrations of GIP, GLP-1, Leptin, and ghrelin. The dysregulation of incretins was first observed at week 10, while prediabetes was only diagnosed at week 20. This dysregulation of incretin hormones precedes the onset of prediabetes and may trigger chronic insulin stimulation which may lead to the development of prediabetes. The results of this study suggest that the dysregulation of incretin concentrations may not only precede the development of prediabetes but may even have a role in its development.

Keywords: Type two diabetes mellitus, incretins, insulin resistance.

1. Introduction

Type 2 diabetes mellitus (T2DM) is a chronic disorder resulting in increased blood glucose concentration over time (68). However, a state of intermediate hyperglycaemia known as prediabetes often precedes the onset of T2DM (69). In this condition, there is elevated blood glucose concentration, but they are not high enough for the diagnosis of T2DM (70). Prediabetes is characterised by a steady decrease in responsiveness to insulin-by-insulin receptors in insulin-dependent cells of the liver, adipose tissue, and skeletal muscle (70).

In reaction to an empty stomach in a healthy individual, A-like cells in the stomach's oxyntic glands secrete a hormone known as ghrelin (32). Ghrelin acts in the hypothalamus to stimulate appetite and induce food intake, leading to the entrance of glucose into the stomach (32). The entrance of glucose in the stomach inhibits ghrelin secretion and triggers the enteroendocrine K cells of the duodenum jejunum and throughout the small intestines to secrete glucose-dependent insulintropic polypeptide (GIP), its release is dependent on the rate of absorption of glucose into the blood (32, 71). GIP is secreted simultaneously with the hormone glucagon-like peptide 1 (GLP-1) produced by the enteroendocrine L cells, and together they are known as the incretin peptides (10). GLP-1's primary functions are stimulating insulin secretion and inhibiting glucagon secretion, reducing postprandial blood glucose concentration, and inducing satiety by binding to its receptors in the hypothalamus (25). Insulin secretion triggers the release of the hormone leptin, it then is transported via a saturable transporter system across the blood-brain barriers (BBB) to leptin or obesity receptors (LEP-R/OB-R) for activation to induce satiety (72). Leptin also stimulates the secretion of GLP-1 from the endocrine L-cells of the small intestines to induce satiety and reduce adipocyte fat storage (11).

In conditions such as T2DM, the efficiency of GLP-1 is impaired as it is actively degraded by the dipeptidyl peptidase 4 enzyme (DPP-4); this enzyme is produced on the surface of most cell types and is linked to immune regulation, signal transduction, and apoptosis (73). However, the secretion of GIP is increased since it is secreted in the presence of glucose (74). Over-secretion of GIP causes a further increase in insulin concentration by constantly promoting insulin secretion further, thus promoting hyperinsulinemia (74). Due to hyperlipidemia and hyperinsulinemia, there is increased inhibition of ghrelin and elevated leptin concentration (75, 76). There is also the development of leptin resistance due to increased plasma leptin concentrations for extended periods (20). This results in impaired satiety, thus resulting in individuals displaying hyperphagia (20). Hyperinsulinemia also promotes lipogenesis in the liver resulting in increased synthesis of lipids (77). This increased lipogenesis plays a significant role in T2DM complications such as heart attacks, stroke, and kidney failure (77). Because of their functions in the regulation of glucose, GIP and GLP-1 have been used as therapeutic targets for the treatment of T2DM (78). DPP4 inhibitors, GIP, and GLP-1 receptor agonists (RA) were discovered because of research into these medicinal targets (78, 79). The dual GIP and GLP-1 RAs show significant

effects on body weight loss and glycemic control, suggesting their therapeutic applications in the treatment of T2DM (78).

Several studies have shown the role of incretin levels on fasting glucose, impaired glucose tolerance, and that complications observed in T2DM often begin during prediabetes (58, 78, 130, 131). While several other studies have demonstrated moderate insulin resistance and hyperinsulinemia in the prediabetic state, there have been no studies showing the role of incretins in the development of prediabetes. Therefore, this study aims to characterize the association between incretin hormone concentration and the development of prediabetes in a diet-induced animal model of prediabetes.

2. Materials and Methods

2.1 Chemicals and drugs

All chemicals and reagents used were purchased from standard pharmaceutical suppliers and were of analytical grade.

2.2 Animals

24 male Sprague-Dawley rats were used in the current study, and the procedure was conducted according to the institutional guidelines of UKZN, Animals Research Ethics Committee (Ethics number: AREC/00003627/2021). The animals were fed and stored at the University of Kwa-Zulu Natal's biomedical research unit (BRU). The animals were maintained in standard laboratory conditions. There was a constant temperature of 25°C, the lighting was cycled between dark and light every 12 hours, the noise was kept below 65 decibels, and they were allowed uncontrolled access to food and water.

2.3.1. Induction of prediabetes

The animals were randomly assigned to the following diet groups; Prediabetes was induced by allowing the animals to feed on the HFHC diet for 20 weeks, as previously described (64, 80). Glucose tolerance was evaluated 5 days after the 20 weeks of induction with a well-known, established laboratory protocol, the oral glucose tolerance test (OGTT), to determine prediabetes according to the American Diabetes Association criteria (81). The rats with fasting blood glucose (FBG) of more than 5.6 mmol/L were considered prediabetic and grouped further for pharmacological studies. The animals fed the standard diet were also tested and found to be normoglycemic without prediabetes.

2.3.2. Experimental design

The twenty-four (24) animals were given one week to acclimatize and separated into two groups. Group 1(n=12) and group 2(n=12). Group 1 and Group 2 were further divided into 2 groups each. They both had a normal diet (ND) group (n=6) which was each given a standard diet with normal drinking water and they both had a high fat high carbohydrates diet group (HFHC) (n=6) which was each given a high fat, high-carbohydrate diet with drinking water supplemented with 15% fructose (AVI Products (Pty

Ltd, Waterfall, South Africa). The composition of the high-fat, high-carbohydrate diet was customized as follows: Carbohydrates (55% KCAL/G), fats (30%KCAL/G), and proteins (15%KCAL/G). At week 10, all the animals in group 1 were sacrificed, while nothing was done to group 2. The animals from group 2 were maintained on their respective diets for another 10 weeks. At week 20, the animals from group 2 were sacrificed.

2.3.3. Blood collection and Termination

At weeks 10 and 20, the animals were anaesthetized with Isofor (100 mg/kg) (Safeline Pharmaceuticals (Pty) Ltd, Roodepoort, South Africa) via a gas anaesthetic chamber (Biomedical Resource Unit, UKZN, Durban, South Africa) for ± 2 minutes. Blood was collected from all the animals using cardiac puncture, and the collected blood was then injected into individual heparinized EDTA containers. Some of this blood was used for glycated haemoglobin measurements. The remainder of the blood was centrifuged (Eppendorf centrifuge 5403, Germany) at 4°C, 503 g for 15 minutes. Plasma was collected and stored at -80 °C in a Bio Ultra freezer (Snijders Scientific, Holland) until ready for biochemical analysis.

2.4. Biochemical Analysis

Plasma GIP, GLP-1, insulin, leptin and ghrelin concentrations were determined using specific ELISA kits in accordance with the manufacturer's instructions (Elabscience and Biotechnology, Wuhan, China). The kits included a micro-ELISA plate that was coated with antibodies specific to each of the parameters measured. The protocols for GLP-1, insulin and leptin were the same as they all required sandwich ELISA. Ghrelin required competitive ELISA, and the procedure was still the same except for the incubation periods, as ghrelin required less time. Standards and samples were pipetted into the appropriate wells of the micro-ELISA plate and incubated for 90 min at 37 °C. This was followed by adding the relevant biotinylated detection antibody (100 μ l). After 60 min incubation at 37 °C, avidin–horseradish peroxidase conjugate (100 μ l) was added to each microplate well. After a further 30 min incubation at 37 °C, the unbound components were washed away using the wash buffer provided.

Substrate solution (100 μ l) was added to each microplate well, and after 15 min incubation at 37 °C, the stop solution (50 μ l) was added. Optical density was measured using a nano spectrophotometer 47 (BMG Labtech, Ortenburg, Germany) at 450 nm. The concentrations of each parameter in the samples were extrapolated from a standard curve.

2.5. HOMA2-IR index

The homeostatic model assessment (HOMA) is a recognized method for measuring insulin resistance (IR) using fasting glucose and insulin. Two variants, known as HOMA1-IR or HOMA2-IR, make up the HOMA. The updated HOMA2-IR index was the choice model since it produced a more accurate index than the original HOMA1-IR that Matthews and Cols released in 1985. An average HOMA-IR value is within the range of 0.5 to 1.4. Insulin sensitivity is at its peak when the index value is less than

1.0. While index values over 1.9 indicate the onset of insulin resistance, and index values over 2.9 indicate substantial insulin resistance, respectively. HOMA-IR was calculated according to the formula: fasting plasma insulin ($\mu\text{U/ml}$) $\times$ fasting plasma glucose (mmol/L)/22.5.

2.6. Statistical analysis

Data expressed as mean $\pm$ S.E.M, using GraphPad Prism Instant software (version 5) to perform the statistical analysis. A student t-test was used to test for statistical significance of the obtained data from the control and the experimental group. P value <0.05 was considered to be significant.

3. Results

3.1. Plasma Glucose

The plasma glucose concentrations were measured at week 10 ($n=12$), halfway to prediabetes induction and at week 20 ($n=12$) when prediabetes is induced. The findings showed that the HFHC group and the PD group had significantly higher plasma glucose concentrations than the ND and NPD groups.

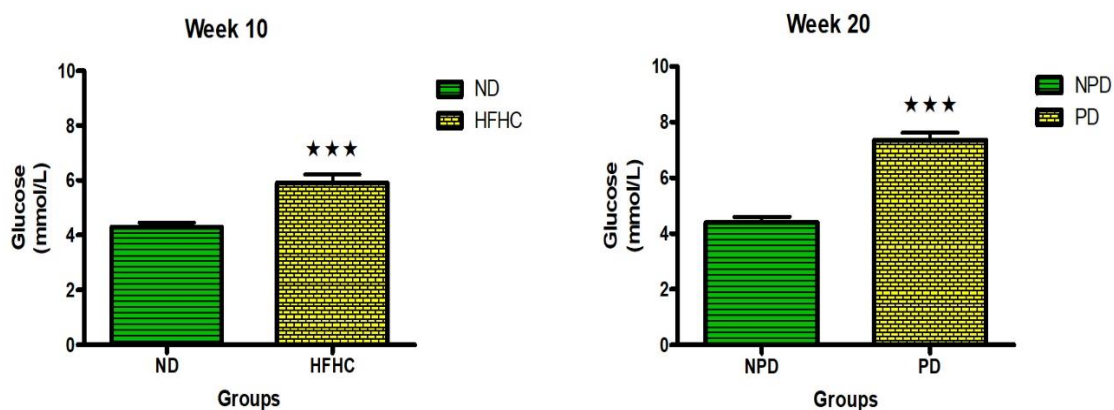


Figure 1: Plasma glucose concentrations in the standard and HFHC diet-fed animals for 10 and 20 weeks. Values are presented as mean $\pm$ SEM ($n=6$). $\star p < 0.05$ denotes comparison with the standard diet group; $\star\star\star p=0.0009$ and $\star\star\star p=0.0001$ respectively.

3.2. Plasma Insulin

The plasma insulin concentrations were measured at week 10 (n=12), halfway to prediabetes induction and at week 20 (n=12) when prediabetes is induced. The findings showed that the HFHC group and the PD group had significantly higher plasma insulin concentrations than the ND and NPD groups.

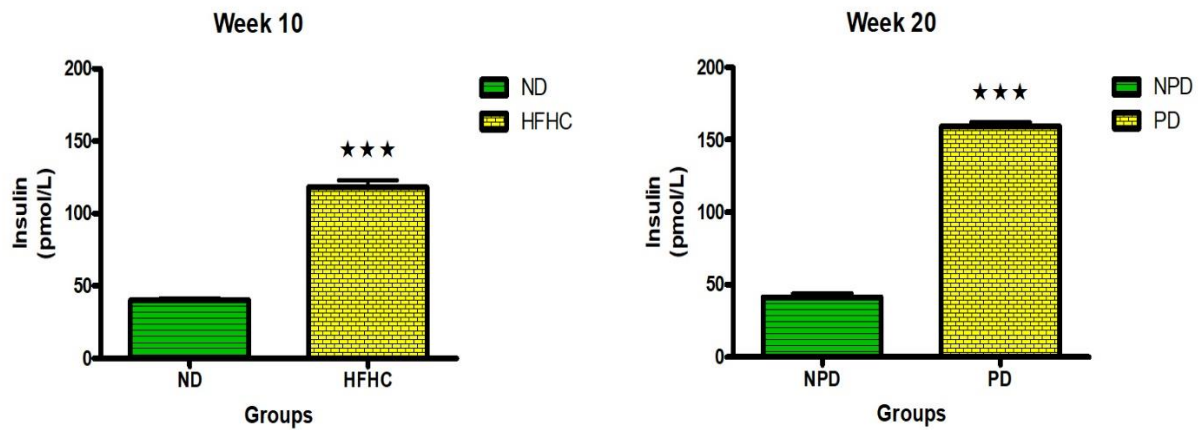


Figure 2: Plasma insulin concentrations in the standard and HFHC diet-fed animals for 10 and 20 weeks. Values are presented as mean $\pm$ SEM (n=6). ★ p < 0.05 denotes comparison with the standard diet group; ★★ p = 0.0001.

3.3. HOMA2-IR Index

The HOMA2-IR index was calculated in the ND and HFHC groups at 10 weeks. The HOMA2-IR value of the ND group at 10 weeks was within the insulin-sensitivity range (<1.0). By comparison with the ND group, the HFHC group showed a higher HOMA2-IR value, which was above the early IR range (>1.9) but lower than the overt IR range (>2.9). (★ = IR range > 1.9). The HOMA2-IR value of the NPD group at 20 weeks was within the insulin-sensitivity range (<1.0). By comparison with the NPD, the PD group showed a higher HOMA2-IR value, which was above the early IR range (>1.9) and the overt IR range (>2.9) (★ = IR range > 1.9).

GROUPS	SEM Glucose	SEM Insulin	HOMA2-IR	p value Glucose	p value Insulin
ND (10 weeks)	0.15	1.41	0.73	0.0001	0.0001
HFHC (10 weeks)	0.31	4.66	2.27★		
NPD (20 weeks)	0.20	2.69	0.75	0.0009	0.0001
PD (20 weeks)	0.28	2.85	3.17★		

Table 1: Displays the SEM, p value and the HOMA2-IR index calculated at weeks 10 and 20 using the HOMA2 calculator in the NPD and PD groups (n=6 per group).

3.4. Plasma GIP

The plasma GIP concentrations were measured at week 10 (n=12), halfway to prediabetes induction and at week 20 (n=12) when prediabetes is induced. The findings showed that the HFHC group and the PD group had significantly higher plasma GIP concentrations than the ND and NPD groups.

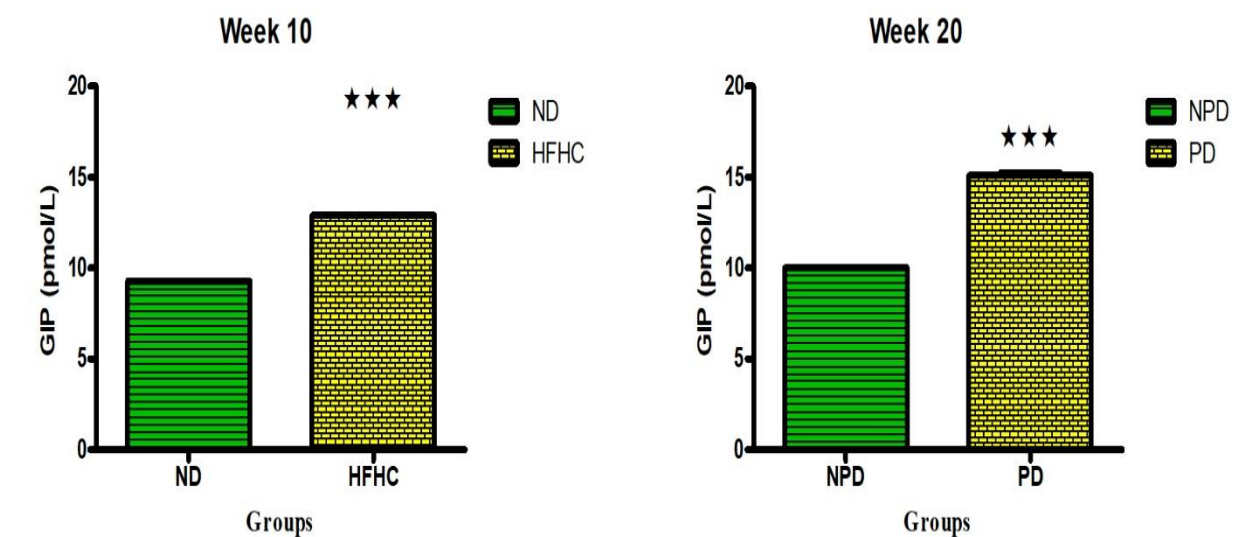


Figure 3: Plasma GIP concentrations in the standard and HFHC diet-fed animals for 10 and 20 weeks. Values are presented as mean $\pm$ SEM (n=6). ★ $p < 0.05$ denotes comparison with the standard diet group; ★★★ $p = 0.0001$.

3.5. Plasma Leptin

The plasma leptin concentrations were measured at week 10 (n=12), halfway to prediabetes induction and at week 20 (n=12) when prediabetes is induced. The findings showed that the HFHC group and the PD group had significantly lower plasma leptin concentrations than the ND and NPD groups.

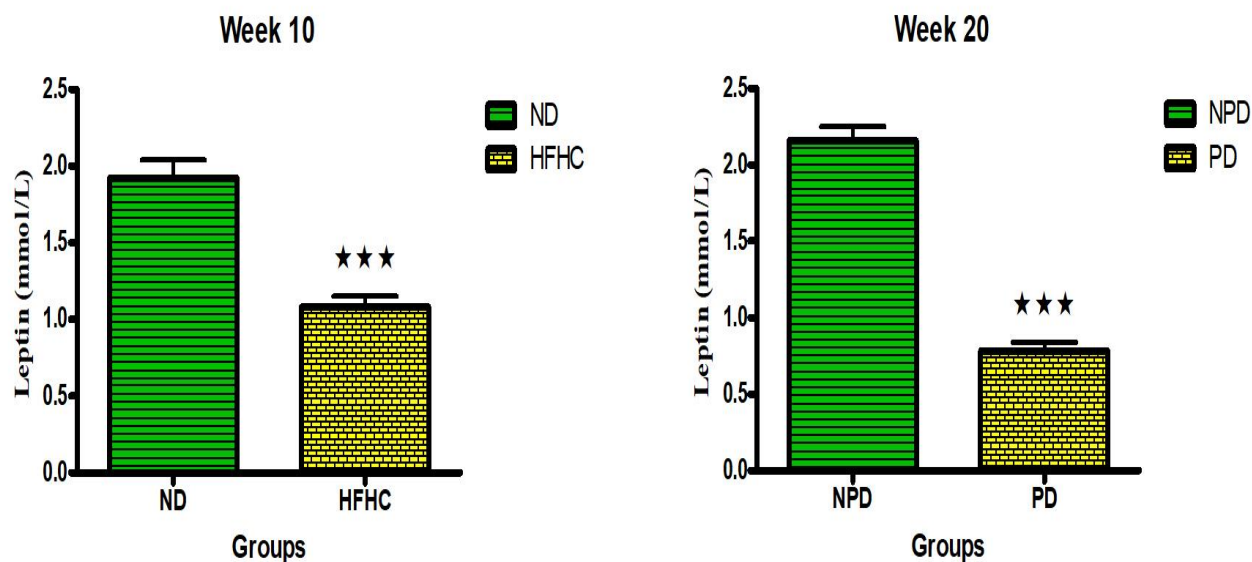


Figure 4: Plasma leptin concentrations in the standard and HFHC diet-fed animals for 10 and 20 weeks.

Values are presented as mean $\pm$ SEM (n=6). ★ $p < 0.05$ denotes comparison with the standard diet group;

★★★ $p = 0.0001$.

3.6. Plasma GLP-1

The plasma GLP-1 concentrations were measured at week 10 (n=12), halfway to prediabetes induction and at week 20 (n=12) when prediabetes is induced. The findings showed that the HFHC group and the PD group had significantly lower plasma GLP-1 concentrations than the ND and NPD groups.

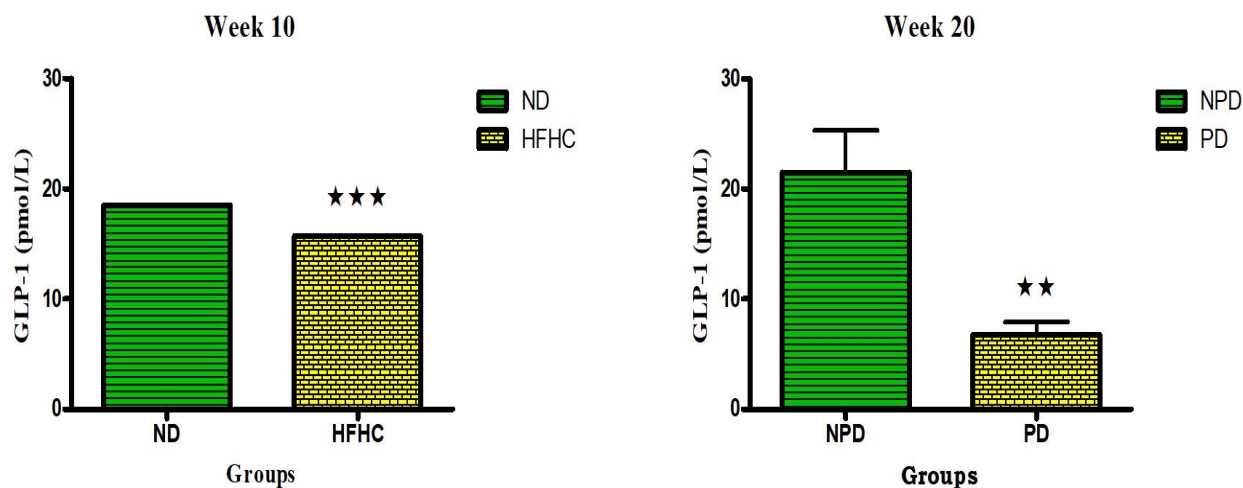


Figure 5: Plasma GLP-1 concentrations in the standard and HFHC diet-fed animals for 10 and 20 weeks. Values are presented as mean $\pm$ SEM (n=6). ★ $p < 0.05$ denotes comparison with the standard diet group; ★★★ $p = 0.0001$ and ★★ $p = 0.0019$.

3.7. Plasma Ghrelin

The plasma ghrelin concentrations were measured at week 10 (n=12), halfway to prediabetes induction and at week 20 (n=12) when prediabetes is induced. The findings showed that the HFHC group and the PD group had significantly higher plasma ghrelin concentrations than the ND and NPD groups.

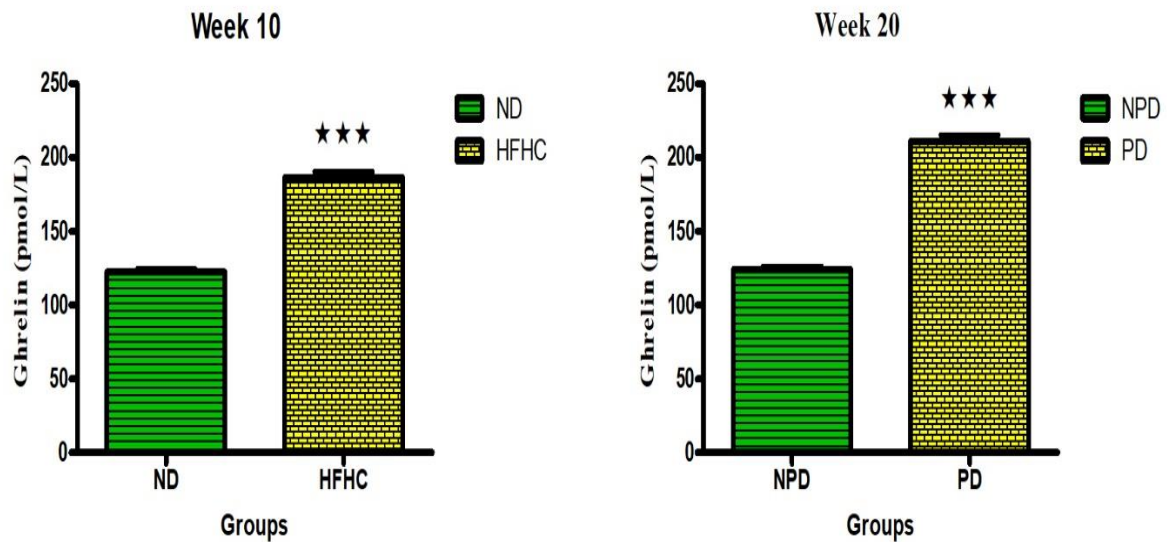


Figure 6: Plasma ghrelin concentrations in the standard and HFHC diet-fed animals for 10 and 20 weeks. Values are presented as mean $\pm$ SEM (n=6). ★ $p < 0.05$ denotes comparison with the standard diet group; ★★ $p = 0.0001$.

3.8. Plasma Glycated Hemoglobin (HbA1c)

The plasma HbA1c concentrations were measured at week 10 (n=12), halfway to prediabetes induction and at week 20 (n=12) when prediabetes is induced. The findings showed that the HFHC group slightly increased with no significant difference compared to the ND group. The PD group had a significantly higher glycated hemoglobin concentration than the NPD group.

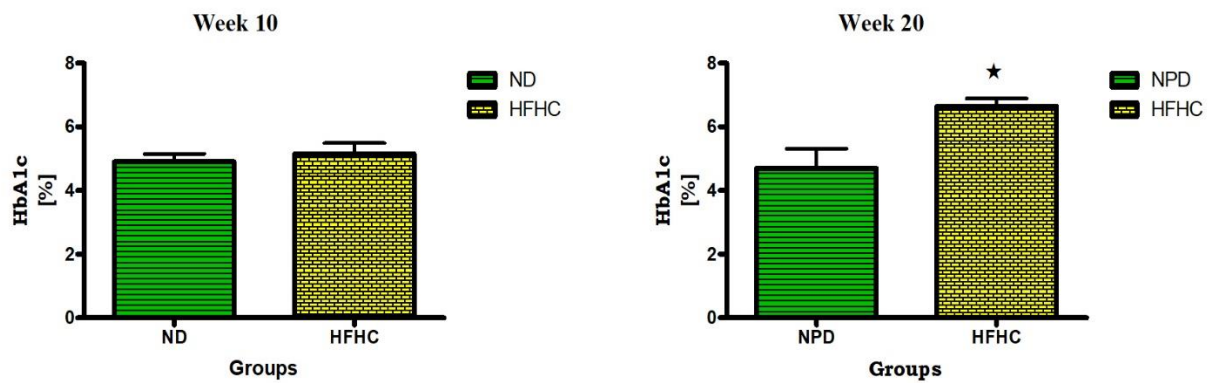


Figure 7: Plasma HbA1c concentrations in the standard and HFHC diet-fed animals for 10 and 20 weeks. Values are presented as mean ± SEM (n=6). ★ $p < 0.05$ denotes comparison with the standard diet group; $p = 0.5667$ and ★ $p = 0.0141$ respectively.

4. Discussion

Studies have shown that subjecting rats to a chronic HFHC diet and drinking water supplemented with fructose for a period of 20 weeks leads to moderate insulin resistance and the development of prediabetes (81, 82). This diet-induced animal model of prediabetes has been shown to mimic the natural progression of prediabetes as it displays moderate hyperglycaemia and moderate insulin resistance (82). Incretin peptides associated with the appetite-regulating hormones work together and aid in glucose homeostasis. The role of incretin in the pathology of type 2 diabetes mellitus has been established. However, minimal work has been done in assessing the link between incretin peptide levels and the development of prediabetes. Therefore, this study sought to characterize the association between incretin peptide concentrations and the development of prediabetes in a diet-induced animal model of prediabetes.

The homeostatic model assessment (HOMA) is a technique for evaluating β -cell function and insulin resistance (IR) using fasting blood glucose and insulin concentrations (83). Elevated blood glucose and insulin concentrations lead to an increased HOMA-IR index, blood glucose concentration is regulated by insulin (83). After food intake, the blood glucose concentration increases, and insulin secretion is stimulated in the pancreatic β -cells (4). Insulin reduces blood glucose concentration by binding to its receptors, therefore, promoting the movement of glucose from the blood into tissue cells (4). However, in conditions like prediabetes and T2DM, there is a dysregulation of insulin caused by a reduced insulin sensitivity (42, 84). Several factors, such as chronic ingestion of high-calorie diets and a sedentary lifestyle, have been implicated in the development of prediabetes. In prediabetes, there is elevated insulin and blood glucose observed in figure 1 and 2 respectively; this resulted in increased HOMA-IR index values as seen in table1.

In this study, we speculate that the elevation of blood glucose and insulin at 10 weeks may be a direct response to the prolonged ingestion of the HFHC diet which was then sustained for a further 10 weeks leading to overt insulin resistance. The American Diabetes Association (ADA) recommends an additional assessment of glycated hemoglobin (HbA_{1c}) in the diagnosis of prediabetes. The HbA_{1c} test measures the amount of blood glucose attached to hemoglobin (85). Hemoglobin is an iron-rich protein in the blood and also transports oxygen and carbon dioxide for respiration (86). Sustained high plasma glucose concentrations lead to glucose-mediated non-enzymatic glycation of hemoglobin which results in a high HbA_{1c} value (85). Glucose in the bloodstream spontaneously links to hemoglobin, and the amount combined with this protein is proportional to how much glucose is in the bloodstream at the time (87). The red blood cells in the human body only last 8-12 weeks (approximately 120 days) before they need to be replaced; this means HbA_{1c} can be used to reflect the average blood glucose concentration over that period, offering a valuable longer-term assessment of blood glucose control (87, 88). The current study, in figure 3 we observed that after 20 weeks of HFHC diet consumption there was a significant increase in HbA_{1c}, indicating developed prediabetes in our animals. Interestingly, after 10 weeks of HFHC diet consumption, there was no statistical difference in HbA_{1c} values in the

HFHC diet-fed animals compared to those kept on a standard diet. In this study, the glucose and insulin results suggested that prediabetes had been developed at week 10, but this was not the case according to ADA criteria which require the additional HbA1c test. While the diagnostic criteria of prediabetes are not the same across the various international organizations, the clinical significance of HbA1c values in monitoring glucose homeostasis is strongly supported by a multitude of studies (89, 90).

The stomach secretes ghrelin in a fasting state into the blood (31). Increased circulating peripheral ghrelin concentration moves to the brain in the hypothalamus and activates the hypothalamic arcuate nucleus, promoting an increased appetite and food intake (52). After ingesting food, elevated blood glucose concentrations result in insulin and leptin secretion (53). Insulin reduces blood glucose, and leptin reduces appetite and food intake; this reduces ghrelin secretion, thus regulating glucose homeostasis (55). However, in T2DM, this pathway is impaired due to IR and hyperlipidaemia, the cells cannot access glucose from the blood, and they constantly transmit signals to the hypothalamus to stimulate increased food intake (56). Ghrelin is secreted in the stomach, and it stimulates the appetite centres in the hypothalamus to increase appetite, hence, the development of chronic hyperphagia (57). After 20 weeks, when prediabetes was induced in the PD group, in figure 4 we found elevated ghrelin levels. This suggests that the hyperphagia observed in T2DM develops from prediabetes as increased ghrelin concentrations induce increased appetite and food intake. However, after 10 weeks of the HFHC diet, before the induction of prediabetes, we found elevated ghrelin concentration with increased blood glucose, which could mean early onset IR. This mimics the human condition as humans develop prediabetes between the ages of 25-45 years. Due to chronic consumption of the HFHC diet, it could be that the body adapts to elevated blood glucose levels over long periods. This could mean that if the blood glucose concentration is reduced, higher concentrations of ghrelin are secreted to induce food intake to increase the blood glucose concentration to the abnormal new normal level, resulting in increased secretion of insulin by the incretin peptides.

GIP is a glucose-dependent peptide; this means its secretion is stimulated by the presence of glucose (91). The presence of glucose in the stomach after a meal stimulates the secretion and the release of this peptide into the bloodstream from the endocrine cells of the small intestines (91). After secretion, GIP induces insulin secretion from the pancreas to reduce the blood glucose concentration. Once there is a low blood glucose concentration, GIP secretion is inhibited, and GIP is degraded by DPP-4 (91). In conditions such as T2DM, where there are significantly elevated blood glucose concentrations, the pathway of GIP secretion is impaired (92). GIP is constantly secreted because of the high blood glucose concentration; the high GIP concentrations are responsible for hyperinsulinemia; GIP constantly induces insulin secretion (92). The animal used in this study were fed a HFHC diet every day for 20 weeks, there were significantly higher concentrations of GIP in the HFHC group compared to the ND group as observed in figure 5. Elevated insulin levels accompany increased GIP concentrations, possibly due to chronic secretion by the pancreatic beta cells. Interestingly at 10 weeks, before the induction of prediabetes, we found significantly high GIP concentrations in the HFHC group. The elevated GIP

levels could be due to the increased blood glucose as GIP is glucose-dependent, increased blood glucose over time stimulates the continued secretion of GIP. The elevated concentration of GIP over a long period could then be responsible for hyperinsulinemia due to continued stimulation of insulin secretion.

In the case of an increase in blood glucose concentration, insulin is secreted and released into the bloodstream (93). Insulin binds to its receptors and is degraded in the liver, adipose tissues and skeletal muscle to allow for the entrance of glucose, thereby reducing its amount in the blood (93). In conditions like T2DM, where there is high glucose concentration, the insulin signalling pathway is disturbed (94). As blood glucose concentration increases, GIP secretion increases, and the pancreatic β cells are forced to overly work to release even more insulin, intending to reduce the blood glucose concentration (94). A prolonged increase in insulin secretion negatively affects the insulin receptors as they gradually lose their sensitivity to insulin (94). The reduced sensitivity by the insulin receptors disturbs the insulin signaling cascade, leading to insulin resistance (93). There were indeed elevated insulin concentrations at 20 weeks in the PD group compared to NPD; this was accompanied by the increased HOMA-IR value, which confirmed the presence of overt insulin resistance in prediabetes. The current literature supports our findings, there is increased insulin concentration in the prediabetic group, which is due to hyperglycaemia and the developing IR present in prediabetes (83). However, after 10 weeks of HFHC diet consumption, before the onset of prediabetes there is significantly high insulin concentration (hyperinsulinemia) in the HFHC group compared to the ND group as seen on figure 2; the high insulin concentration is possibly due to chronic secretion by the elevated GIP concentration. There were elevated insulin concentrations over a long period due to increased blood glucose and GIP. The chronic secretion of insulin leads to hyperinsulinemia, which could be the reason for the onset of insulin resistance shown by the elevated HOMA-IR value in table 1. GIP could be responsible for the sustained hyperinsulinemia from 10 weeks to the induction of prediabetes. GIP could, therefore, be responsible for the onset of insulin resistance observed after 10 weeks. Kanemaru *et al.* published a systematic review and confirmed that prolonged high concentrations of GIP induce insulin resistance (95). Therefore, the findings suggest that GIP might play a significant role in the onset and progression of prediabetes to type 2 diabetes mellitus.

Increased blood glucose concentration has been shown to promote the secretion of leptin into the bloodstream by the adipose tissues, and this is suggested to be due to the increase in insulin which stimulates leptin secretion (96). After secretion by white adipose tissues, leptin increases circulation and crosses the BBB as a saturable transport to the hypothalamic arcuate nucleus, where it binds to LEPR-b (97). Leptin binding promotes the activation of appetite suppressor neurons, thus inducing satiety and boosting energy expenditure by activating the satiety centre (Ventromedial hypothalamus) (97). Thus, leptin acts as an afferent signal in a negative feedback loop that keeps the body fat reserves steady. Leptin secretion is stimulated by insulin in the adipose tissues (98). In conditions like T2DM, the leptin signaling pathway is disturbed (20). Due to the developing IR and hyperinsulinemia, stimulation of the adipose tissues to secrete leptin is diminished. Our findings on figure 6 show

significantly decreased leptin concentrations in the PD group compared to the NPD group after 20 weeks; this could be due to the overt IR due to elevated GIP and ghrelin concentrations. However, after 10 weeks of HFHC diet consumption, the PD group had a significantly reduced leptin concentration; this could be due to the onset of IR and severely increased ghrelin concentrations. Wang *et al.* conducted a study to establish the relationship between IR and leptin in prediabetes; they found that IR is associated positively with diminished leptin concentrations (99). The reduced leptin concentration also means diminished secretion of GLP-1 and decreased satiety which is observed in this study.

GLP-1 is secreted simultaneously with GIP immediately after food consumption and is secreted by the small intestines' endocrine cells, which induce insulin secretion from the pancreas (10). GLP-1 has receptors in the hypothalamus, and once secreted, it induces satiety by binding to the receptors mentioned above (25). Unlike GIP, GLP-1 is degraded faster by dipeptidyl-peptidase 4 (DPP-4) shortly after its secretion due to the presence of the induced insulin secreted into the bloodstream (73). In conditions like T2DM, GLP-1 secretion is diminished due to leptin resistance and GLP-1 degradation is increased to hyperinsulinemia and DPP-4 (73). Increased plasma DPP-4 is associated with increased blood glucose; it degrades the reduced concentration of GLP-1 shortly after secretion (73). Satiety is, therefore, not induced via the GLP-1 pathway. In the current study, on figure 7 there were severely low levels of GLP-1 in the PD group, possibly due to the reduced Leptin concentration and increased DPP4. However, at 10 weeks, we also observed a significant decrease in GLP-1 in the prediabetic group compared to the non-prediabetic control, this could be due to the onset of IR and the reduced concentration of leptin present. Hira *et al.* conducted a study to find out what GLP-1 does. They found that it was reduced in prediabetic subjects; therefore, the current literature supports the results obtained in this study (100). Both the leptin and GLP-1 satiety-inducing pathways are disturbed. Therefore, there is diminished satiety in prediabetes.

Conclusion

In this study, we observed that the prolonged consumption of a HFHC diet results in abnormal incretin concentrations. This is evidenced by elevated ghrelin and decreased plasma leptin levels. This increases food intake and blood glucose levels, resulting in increased GIP and reduced plasma levels of GLP-1. The increased GIP levels may then cause chronic insulin stimulation and can result in hyperinsulinemia, which can result in insulin resistance. Hence, chronic consumption of the HFHC diet increases blood glucose and insulin. The dysregulated plasma incretin concentrations then maintain these abnormal concentrations leading to the onset of insulin resistance and later prediabetes. As a result, the incretins are linked to the onset and progression of prediabetes. This could further suggest that incretins play a role in the development of prediabetes to T2DM.

Acknowledgments

The authors would like to acknowledge the endocrine group for their support, the Biomedical Resource

Units of KwaZulu-Natal for their technical assistance. Additionally, the authors are grateful for the generous support from the National Research Foundation and UKZN College of Health Science who provided financial assistance.

Author Contributions

NM and NRD contributed to the study design, conducted the experiments, collected, analyzed, and interpreted data, as well as being involved in writing the manuscript. AMS and PSN were involved in the study design, interpretation of data and editing of the manuscript. AK was involved in the conceptualization of the study, study design, analysis, and interpretation of data, writing and editing of the manuscript as well as provide funding. All authors have read and approved submission of the final manuscript.

Conflict of Interest

The authors declare no conflict of interest.

Availability of Data and Materials

The datasets presented in this study are included in the article and available on reasonable request from the corresponding author.

Ethics Approval and Consent to Participate

The animal study protocol was reviewed and approved by the Animal Research Ethics Committee (AREC) of the University of KwaZulu-Natal, South Africa (AREC/00003627/2021).

References

1. Oguntibeju OO. Type 2 diabetes mellitus, oxidative stress and inflammation: examining the links. *Int J Physiol Pathophysiol Pharmacol*. 2019;11(3):45-63.
2. Zamora-Kapoor A, Fyfe-Johnson A, Omidpanah A, Buchwald D, Sinclair Ki. Risk factors for pre-diabetes and diabetes in adolescence and their variability by race and ethnicity. *Prev Med*. 2018;115:47-52.
3. Hostalek U. Global epidemiology of prediabetes - present and future perspectives. *Clin Diabetes Endocrinol*. 2019;5:5.
4. Suyama S, Yada T. New insight into GABAergic neurons in the hypothalamic feeding regulation. *The Journal of Physiological Sciences*. 2018;68(6):717-22.
5. Joo E, Harada N, Yamane S, Fukushima T, Taura D, Iwasaki K, et al. Inhibition of Gastric Inhibitory Polypeptide Receptor Signaling in Adipose Tissue Reduces Insulin Resistance and Hepatic Steatosis in High-Fat Diet–Fed Mice. *Diabetes*. 2017;66(4):868.
6. Lim GE, Brubaker PL. Glucagon-Like Peptide 1 Secretion by the L-Cell. *Diabetes*. 2006;55(Supplement 2):S70.
7. Müller TD, Finan B, Bloom SR, D'Alessio D, Drucker DJ, Flatt PR, et al. Glucagonlike peptide 1 (GLP-1). *Molecular Metabolism*. 2019;30:72-130.
8. Van Doorn C, Macht VA, Grillo CA, Reagan LP. Leptin resistance and hippocampal behavioral deficits. *Physiology & behavior*. 2017;176:207-13.
9. Ronveaux CC, Tomé D, Raybould HE. Glucagon-like peptide 1 interacts with ghrelin and leptin to regulate glucose metabolism and food intake through vagal afferent neuron signaling. *J Nutr*. 2015;145(4):672-80.
10. Nauck MA, Quast DR, Wefers J, Meier JJ. GLP-1 receptor agonists in the treatment of type 2 diabetes – state-of-the-art. *Molecular Metabolism*. 2021;46:101102.
11. Fu Y, Kaneko K, Lin H-Y, Mo Q, Xu Y, Suganami T, et al. Gut Hormone GIP Induces Inflammation and Insulin Resistance in the Hypothalamus. *Endocrinology*. 2020;161(9).
12. Shankar K, Takemi S, Gupta D, Varshney S, Mani BK, Osborne-Lawrence S, et al. Ghrelin cell-expressed insulin receptors mediate meal- and obesity-induced declines in plasma ghrelin. *JCI Insight*. 2021;6(18).

13. Omran DM, Alaraji SM, Albayati AH, Essam W. Relationship between Ghrelin and Leptin with Insulin Resistance in Obese Patients and Non-Obese Individuals. *Research Journal of Pharmacy and Technology*. 2018;11(1):281-3.
14. Zhou Y, Rui L. Leptin signaling and leptin resistance. *Frontiers of medicine*. 2013;7(2):207-22.
15. Papatheodorou K, Banach M, Bekiari E, Rizzo M, Edmonds M. Complications of Diabetes 2017. *Journal of Diabetes Research*. 2018;2018:3086167.
16. Abdul-Ghani MA, DeFronzo RA. Pathophysiology of prediabetes. *Current Diabetes Reports*. 2009;9(3):193-9.
17. Fisman EZ, Tenenbaum A. The dual glucose-dependent insulinotropic polypeptide (GIP) and glucagon-like peptide-1 (GLP-1) receptor agonist tirzepatide: a novel cardiometabolic therapeutic prospect. *Cardiovasc Diabetol*. 2021;20(1):225.
18. Muzurović EM, Volčanšek Š, Tomšić KZ, Janež A, Mikhailidis DP, Rizzo M, et al. Glucagon-Like Peptide-1 Receptor Agonists and Dual Glucose-Dependent Insulinotropic Polypeptide/Glucagon-Like Peptide-1 Receptor Agonists in the Treatment of Obesity/Metabolic Syndrome, Prediabetes/Diabetes and Non-Alcoholic Fatty Liver Disease—Current Evidence. *Journal of Cardiovascular Pharmacology and Therapeutics*. 2022;27:10742484221146371.
19. Ahren B. DPP-4 inhibitors. *Best Practice & Research Clinical Endocrinology & Metabolism*. 2007;21(4):517-33.
20. Tabák AG, Herder C, Rathmann W, Brunner EJ, Kivimäki M. Prediabetes: a high-risk state for diabetes development. *Lancet*. 2012;379(9833):2279-90.
21. Mzimela NC, Ngubane PS, Khathi A. The changes in immune cell concentration during the progression of pre-diabetes to type 2 diabetes in a high-fat high-carbohydrate diet-induced pre-diabetic rat model. *Autoimmunity*. 2019;52(1):27-36.
22. Ha K, Joung H, Song Y. Inadequate fat or carbohydrate intake was associated with an increased incidence of type 2 diabetes mellitus in Korean adults: a 12-year community-based prospective cohort study. *Diabetes research and clinical practice*. 2019;148:254-61.

23. Gamede M, Mabuza L, Ngubane P, Khathi A. The effects of plant-derived oleanolic acid on selected parameters of glucose homeostasis in a diet-induced pre-diabetic rat model. *Molecules*. 2018;23(4):794.
24. Luvuno M, Khathi A, Mabandla MV. Diet-induced prediabetes: effects of exercise treatment on risk factors for cardiovascular complications. *Nutrition & Metabolism*. 2021;18(1):1-9.
25. Sosibo AM. Investigating changes to the insulin signalling pathway in a diet-induced pre-diabetic rat model: effects on selected markers 2019.
26. Wang T, Wang J, Hu X, Huang X-J, Chen G-X. Current understanding of glucose transporter 4 expression and functional mechanisms. *World journal of biological chemistry*. 2020;11(3):76.
27. Association AD. 2. Classification and Diagnosis of Diabetes: Standards of Medical Care in Diabetes—2021. *Diabetes Care*. 2020;44(Supplement_1):S15-S33.
28. Freeman AM, Pennings N. Insulin resistance. *StatPearls [Internet]*. 2021.
29. Kostov K, Blazhev A. Use of glycated hemoglobin (A1c) as a biomarker for vascular risk in type 2 diabetes: Its relationship with matrix metalloproteinases-2,-9 and the metabolism of collagen IV and elastin. *Medicina*. 2020;56(5):231.
30. Ahmed MH, Ghatge MS, Safo MK. Hemoglobin: structure, function and allostery. *Subcellular biochemistry*. 2020;94:345.
31. Sacks DB, Bebu I, Lachin JM. Refining measurement of hemoglobin A1c. *Clinical chemistry*. 2017;63(9):1433-5.
32. Wang C, Ye Y, Sun W, Yu J, Wang J, Lawrence DS, et al. Red blood cells for glucoseresponsive insulin delivery. *Advanced Materials*. 2017;29(18):1606617.
33. Sherwani SI, Khan HA, Ekhzaimy A, Masood A, Sakharkar MK. Significance of HbA1c test in diagnosis and prognosis of diabetic patients. *Biomarker insights*. 2016;11:BMI. S38440.
34. Renz PB, Cavagnolli G, Weinert LS, Silveiro SP, Camargo JL. HbA1c test as a tool in the diagnosis of gestational diabetes mellitus. *PLoS One*. 2015;10(8):e0135989.
35. Sun Y, Wang P, Zheng H, Smith RG. Ghrelin stimulation of growth hormone release and appetite is mediated through the growth hormone secretagogue receptor. *Proceedings of the National Academy of Sciences*. 2004;101(13):4679-84.

36. Korbonits M, Goldstone AP, Gueorguiev M, Grossman AB. Ghrelin—a hormone with multiple functions. *Frontiers in Neuroendocrinology*. 2004;25(1):27-68.
37. Yaney G, Corkey B. Fatty acid metabolism and insulin secretion in pancreatic beta cells. *Diabetologia*. 2003;46(10):1297-312.
38. Satoh N, Ogawa Y, Katsuura G, Tsuji T, Masuzaki H, Hiraoka J, et al. Pathophysiological Significance of the Obese Gene Product, Leptin, in Ventromedial Hypothalamus (VMH)-Lesioned Rats: Evidence for Loss of Its Satiety Effect in VMH-Lesioned Rats★. *Endocrinology*. 1997;138(3):947-54.
39. Adamska E, Ostrowska L, Górski M, Krętownski A. The role of gastrointestinal hormones in the pathogenesis of obesity and type 2 diabetes. *Prz Gastroenterol*. 2014;9(2):6976.
40. Ibrahim Abdalla MM. Ghrelin - Physiological Functions and Regulation. *Eur Endocrinol*. 2015;11(2):90-5.
41. El K, Campbell JE. The role of GIP in α -cells and glucagon secretion. *Peptides*. 2020;125:170213.
42. Samms RJ, Coghlan MP, Sloop KW. How may GIP enhance the therapeutic efficacy of GLP-1? *Trends in Endocrinology & Metabolism*. 2020;31(6):410-21.
43. Petersen MC, Shulman GI. Mechanisms of insulin action and insulin resistance. *Physiological reviews*. 2018;98(4):2133-223.
44. Fu Y, Kaneko K, Lin H-Y, Mo Q, Xu Y, Suganami T, et al. Gut hormone GIP induces inflammation and insulin resistance in the hypothalamus. *Endocrinology*. 2020;161(9):bqaa102.
45. Kanemaru Y, Harada N, Shimazu-Kuwahara S, Yamane S, Ikeguchi E, Murata Y, et al. Absence of GIP secretion alleviates age-related obesity and insulin resistance. *Journal of Endocrinology*. 2020;245(1):13-20.
46. D'souza AM, Neumann UH, Glavas MM, Kieffer TJ. The glucoregulatory actions of leptin. *Molecular metabolism*. 2017;6(9):1052-65.
47. Obradovic M, Sudar-Milovanovic E, Soskic S, Essack M, Arya S, Stewart AJ, et al. Leptin and Obesity: Role and Clinical Implication. *Frontiers in Endocrinology*. 2021;12.

48. Paz-Filho G, Mastronardi C, Wong ML, Licinio J. Leptin therapy, insulin sensitivity, and glucose homeostasis. *Indian J Endocrinol Metab.* 2012;16(Suppl 3):S549-55.
49. Wang G, Liu X, Christoffel KK, Zhang S, Wang B, Liu R, et al. Prediabetes is not all about obesity: association between plasma leptin and prediabetes in lean rural Chinese adults. *European Journal of Endocrinology.* 2010;163(2):243-9.
50. Hira T, Pinyo J, Hara H. What is GLP-1 really doing in obesity? *Trends in Endocrinology & Metabolism.* 2020;31(2):71-80.
51. Laakso M, Zilinskaite J, Hansen T, Boesgaard TW, Vanttinen M, Stancáková A, Jansson PA, Pellmé F, Holst JJ, Kuulasmaa T, Hribal ML, Sesti G, Stefan N, Fritsche A, Häring H, Pedersen O, Smith U; EUGENE2 Consortium. Insulin sensitivity, insulin release and glucagon-like peptide-1 levels in persons with impaired fasting glucose and/or impaired glucose tolerance in the EUGENE2 study. *Diabetologia.* 2008 Mar;51(3):502-11. doi: 10.1007/s00125-007-0899-2. Epub 2007 Dec 14. PMID: 18080106.
52. Vollmer K, Holst JJ, Baller B, Ellrichmann M, Nauck MA, Schmidt WE, Meier JJ. Predictors of incretin concentrations in subjects with normal, impaired, and diabetic glucose tolerance. *Diabetes.* 2008 Mar;57(3):678-87. doi: 10.2337/db07-1124. Epub 2007 Dec 5. PMID: 18057091.

Chapter 3: Manuscript 2

Investigating the effect of a low carbohydrate, high unsaturated fats diet on diet-induced prediabetes.

Prologue 2

In the first manuscript we found that the abnormal incretin concentrations not only precede, but they also play a role in the development of prediabetes. In this study, we sought to further explore the involvement of incretins in prediabetes by observing how subjecting our prediabetic animal model with a low carbohydrate, high unsaturated fat diet would affect them.

The manuscript in chapter 2 titled "**Investigating the effect of a low carbohydrate, high fat diet in diet-induced prediabetes**" is authored by N. Mzimela, NR Dimba, A.M Sosibo, and A Khathi.

This manuscript has been submitted and is currently under review in the Endocrinologia, Diabetes y Nutricion journal.

Investigating the effect of a low carbohydrate, high fat diet in diet-induced prediabetes.

Nhlakanipho Mzimela¹★, Nosipho R. Dimba¹, Aubrey M. Sosibo¹, and Andile Khathi ¹

¹ School of Laboratory Medicine and Medical Sciences, Discipline of Human Physiology, University
of KwaZulu-Natal, South Africa

★Correspondence: 218006756@stu.ukzn.ac.za (Nhlakanipho Mzimela)

Abstract

Type 2 diabetes mellitus is a chronic metabolic disease that results from a loss of sensitivity to insulin by-insulin receptors in insulin dependent cells. In this condition, there are chronically increased insulin and blood glucose concentrations. An intermediate hyperglycemic condition known as prediabetes was found to precede the onset of T2DM. Prediabetes is a chronic metabolic condition that has been shown to be reversible using dietary intervention and has become a therapeutic target to prevent T2DM. However, the effect of a low carbohydrate, high unsaturated fat (LCHF) diet on incretin levels during prediabetes has not been established. Hence, this study aims to characterise the effect of a LCHF on incretin levels in a diet-induced prediabetic animal model. This study was conducted using 18 male Sprague-Dawley rats, divided into two groups, group A was given a standard rat diet (SD) (n=6), while group B was given a high-fat high carbohydrate (HFHC) (n=12) diet. The animals were given their respective diet for a total of 20 weeks. After 20 weeks, pre-diabetes was induced in group B. Group A continued with the standard diet and was used as a non-prediabetic control (NPDC) (n=6). The pre-diabetic group B (n=12) was split into two experimental groups. One of the groups continued a HFHC diet and served as the pre-diabetic control group (PD)(n=6), while the other group had a diet intervention where the diet was changed to a low carbohydrate-high unsaturated fats diet (PD+DI) (n=6). All groups were then maintained on their respective diets for a further 12 weeks. Chronic consumption of the HFHC diet by the PD group led to elevated blood glucose (p=0.0010) and insulin concentration (p=0.0001), resulting in abnormal concentrations of GIP (p=0.0010), GLP-1 (0.0199), Leptin (p=0.0001) and Ghrelin (p=0.8647). The abnormal incretins then maintained this state of hyperglycemia and hyperinsulinemia, resulting in pre-diabetes. Chronic consumption of the LCHF by the PD+DI group led to reduced concentrations of GIP (p=0.0010), GLP-1 (0.0199), Leptin (p=0.0212) and Ghrelin (p=0.8112), which could have led to a reduced HbA1c and eventually to the reversal of pre-diabetes.

Keywords: incretins, prediabetes, low carbohydrates, high unsaturated fat diet.

1. Introduction

The global prevalence of type two diabetes mellitus (T2DM) has increased rapidly in recent years and is one of the leading causes of morbidity and mortality worldwide (101). T2DM is often preceded by a condition known as pre-diabetes (102). According to the American Diabetes Association (ADA), prediabetes is 4 by impaired fasting glucose (IFG), impaired glucose tolerance (IGT) and elevated glycated hemoglobin (HbA1c) concentration (81). Pre-diabetes is a long-lasting condition resulting from prolonged consumption of high-calorie diets and a sedentary lifestyle (103). Literature evidence shows that a diet rich in saturated fat and refined carbohydrates is strongly linked to the development of pre-diabetes. (64, 80, 129). This was demonstrated in a recent study in our laboratory, where we produced an animal model of diet-induced pre-diabetes by exposing rats to chronic consumption of a high-fat, high-carbohydrate (HFHC) diet (129). The findings of that study revealed that chronic ingestion of a HFHC diet is directly related to a change in incretin levels, which may play a role in the emergence of mild insulin resistance and pre-diabetes (129).

It has been demonstrated that incorporating moderate exercise into the management of pre-diabetes may benefit pre-diabetic patients in improving glucose tolerance, but these effects are short-lived as they are often associated with poor patient compliance (104). Literature evidence shows that incorporating a low-carbohydrate and high-unsaturated fat diet may alleviate the moderate insulin resistance observed in pre-diabetes (104). Hence, dietary intervention has become a target to reduce the incidence of T2DM. Several randomized controlled trials (RCT) studies indicated that lifestyle interventions that seek to reduce caloric intake or increase physical activity could delay or prevent the onset of T2DM (105, 106). Under normal physiological conditions, gut-derived incretin hormones such as glucose-dependent insulinotropic polypeptide (GIP) and glucagon-like peptide-1 (GLP-1) are secreted shortly after the detection of glucose in the stomach (107). They induce insulin secretion before glucose is absorbed into the blood to prepare for increased blood glucose concentration (108). The incretin effect explains how oral glucose consumption evokes more robust insulin secretory responses than intravenous glucose despite yielding the same glycemia levels (109). GIP and GLP-1 mediate this action, which is consistently deficient in type 2 diabetes patients due to impaired incretin release (109). However, a previous study in our laboratory shows how chronic ingestion of a HFHC diet is associated with increased/decreased incretin hormones, leading to the development of insulin resistance observed in pre-diabetes (64). The incretin hormones are influenced by diet because they are glucose-dependent polypeptides produced by the enteroendocrine cells (107). The effect of incretins on glucose homeostasis is well understood, and incretin-based medicines are among the most promising novel treatments for type 2 diabetes (107, 108, 109). Incretins work closely with the appetite-controlling hormones ghrelin and leptin to regulate blood glucose concentration (110). Ghrelin and leptin regulate appetite and work in opposition to each other; ghrelin is secreted just before a meal and increases appetite to induce food intake, while leptin is secreted when there is increased blood glucose and reduces

appetite (111). A higher glucose-content diet stimulates a higher secretion of the incretin hormones and, therefore, a constant secretion of insulin, resulting in diabetes mellitus (112).

Previous papers have examined the effects of low carbohydrates and high unsaturated fats on glucose homeostasis in T2DM and found them to be beneficial in its management (109, 113). However, the effects of this type of diet on incretin levels in the pre-diabetic state remain unknown. In this study, we sought to investigate the effects of a low carbohydrate and high unsaturated fats diet on incretin hormones and the association with glucose homeostasis.

2. Methods and Materials

2.1. Chemicals and drugs

All chemicals and reagents used were purchased from standard pharmaceutical suppliers and were of analytical grade.

2.2. Animals and housing

This study used 18 male Sprague-Dawley rats (150-180 g). The animals were housed in a room with a 12-hour light/12hour dark cycle, at room temperature (25°C), for the duration of the study. The animals in each group had access to food and water. The Animal Research Ethics Committee approved all animal procedures and housing conditions of the University of KwaZulu-Natal (Ethics number: AREC/00003627/2021). The animals were monitored for pain, discomfort and distress using the criteria listed in the university's Animal Research Ethics Committee's humane endpoint document.

2.3.1. Induction of pre-diabetes

The animals were randomly assigned to the following diet groups; Group A (n=6): rats that consumed a standard diet with drinking water (NPDC). Group B (n=12); rats that consumed a high fat, high carbohydrate diet (HFHC) with drinking water supplemented with 15% fructose (AVI Products (Pty) Ltd, Waterfall, South Africa) (83). Pre-diabetes was induced by allowing the animals to feed on the HFHC diet for 20 weeks, as previously described (64, 80). Glucose tolerance was evaluated 5 days after the 20 weeks of induction with a well-known, established laboratory protocol, the oral glucose tolerance test (OGTT), to determine pre-diabetes according to the American Diabetes Association criteria (81). Group A was found to be non-diabetic while group B was found to be pre-diabetic. The animals in group B were then grouped further for pharmacological studies. The animals fed the normal diet were also tested and found to be normoglycemic and without pre-diabetes.

2.3.2. Experimental design

After 20 weeks of pre-diabetes induction, group A continued with the standard diet and was used as a non-prediabetic control (NPDC) (n=6). The pre-diabetic group B (n=12) was split into two experimental groups. One of the groups continued a HFHC diet and served as the pre-diabetic control group (PD)(n=6), while the other group had a diet intervention where the diet was changed to a low

carbohydrate-high unsaturated fats diet (PD+DI) (n=6). All groups were then maintained on their respective diets for a further 12 weeks.

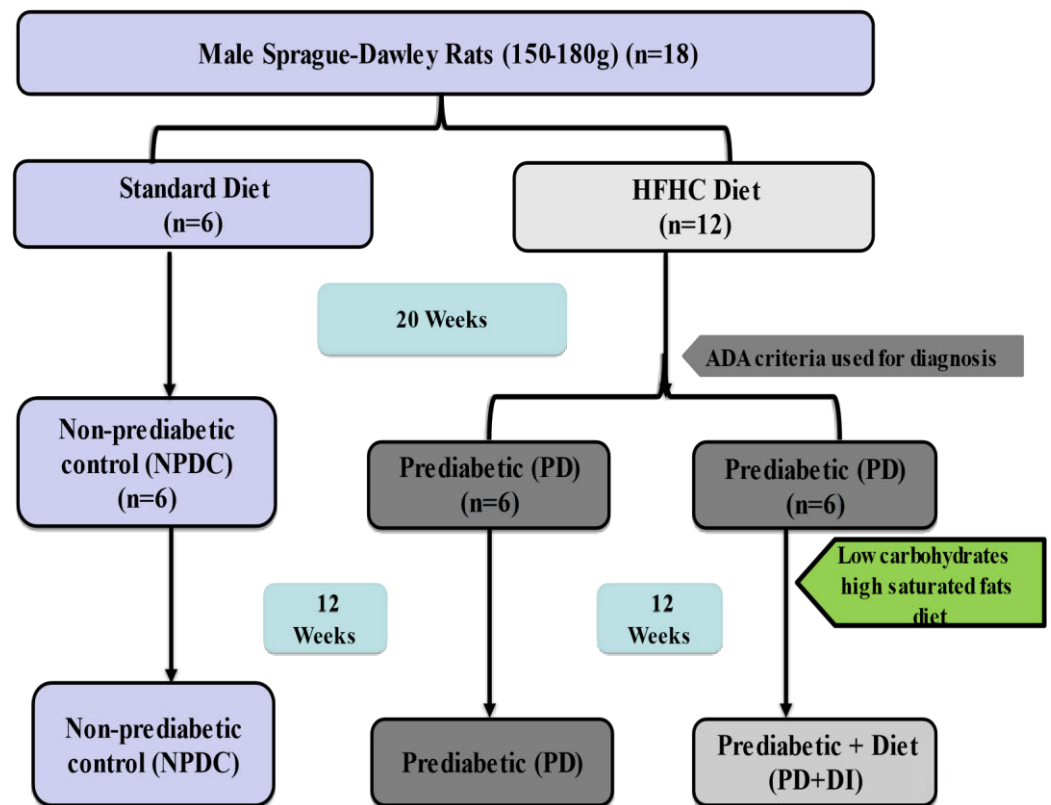


Figure 1. Schematic diagram showing the experimental design.

2.3.3. Blood collection and tissue harvesting

All animals were anaesthetized with Isofor (100 mg/kg) (Safeline Pharmaceuticals (Pty) Ltd, Roodeport, South Africa) using a gas anesthetic chamber (Biomedical Resource Unit, University of KwaZulu-Natal, Durban, South Africa) and allowed to inhale for 3 minutes. Blood was collected by cardiac puncture and injected into pre-cooled heparinized containers. The collected blood was injected into individual heparinized and EDTA containers and then centrifuged (Eppendorf centrifuge 5403, Germany) at 4°C, 503 g for 15 minutes. Plasma was collected and stored at -80 °C in a Bio Ultra freezer (Snijders Scientific, Holland) until ready for biochemical analysis.

2.4 Biochemical analysis

Plasma GIP, GLP-1, insulin, leptin and ghrelin concentrations were determined using specific ELISA kits in accordance with the manufacturer's instructions (Elabscience and Biotechnology, Wuhan, China). The kits included a micro-ELISA plate that was coated with antibodies specific to each of the parameters measured. The protocols for GLP-1, insulin and leptin were the same as they all required

sandwich ELISA. Ghrelin required competitive ELISA, and the procedure was still the same except for the incubation periods, as ghrelin required less time. Standards and samples were pipetted into the appropriate wells of the micro-ELISA plate and incubated for 90 min at 37 °C. This was followed by adding the relevant biotinylated detection antibody (100 µl). After 60 min incubation at 37 °C, avidin–horseradish peroxidase conjugate (100 µl) was added to each microplate well. After a further 30 min incubation at 37 °C, the unbound components were washed away using the wash buffer provided. Substrate solution (100 µl) was added to each microplate well, and after 15 min incubation at 37 °C, the stop solution (50 µl) was added. Optical density was measured using a nano spectrophotometer 47 (BMG Labtech, Ortenburg, Germany) at 450 nm. The concentrations of each parameter in the samples were extrapolated from their respective standard curves.

2.5. HOMA2-IR index

The homeostatic model assessment (HOMA) is a recognized method for measuring insulin resistance (IR) using fasting glucose and insulin. Two variants, known as HOMA1-IR or HOMA2-IR, make up the HOMA. The updated HOMA2-IR index was the choice model since it produced a more accurate index than the original HOMA1-IR that Matthews and Cols released in 1985. An average HOMA-IR value is within the range of 0.5 to 1.4. Insulin sensitivity peaks when the index value is less than 1.0. While index values over 1.9 indicate the onset of insulin resistance, and index values over 2.9, indicate substantial insulin resistance, respectively. HOMA-IR was calculated using fasting plasma insulin (µU/ml) x fasting plasma glucose (mmol/L)/22.5.

2.6 Statistical analysis

Data were reported as mean ± SEM. GraphPad Prism Software (version 5) was used to perform statistical analysis. The differences between non-diabetic, pre-diabetic and dietary intervention pre-diabetic groups were analyzed using One-way analysis of variance (ANOVA) followed by Tukey-Kramer. Values of $P < 0.05$ show statistical significance between the compared groups. Correlation analysis was also performed using a scatter chart from Pearson's comparative analysis. A value of $r > 0.7$ or $r < -0.7$ was considered a strong association.

3. Results

3.1. Plasma GIP

The plasma GIP concentrations were measured at the end of the experimental period at 32 weeks. The findings showed that the PD group had significantly higher plasma GIP levels than the NPDC group ($p=0.0010$) and the PD+DI group. The plasma GIP concentration of the PD+DI group was slightly lower with no significant difference than that of the NPDC group ($p=0.7914$) and significantly lower than the PD group ($p=0.0010$).

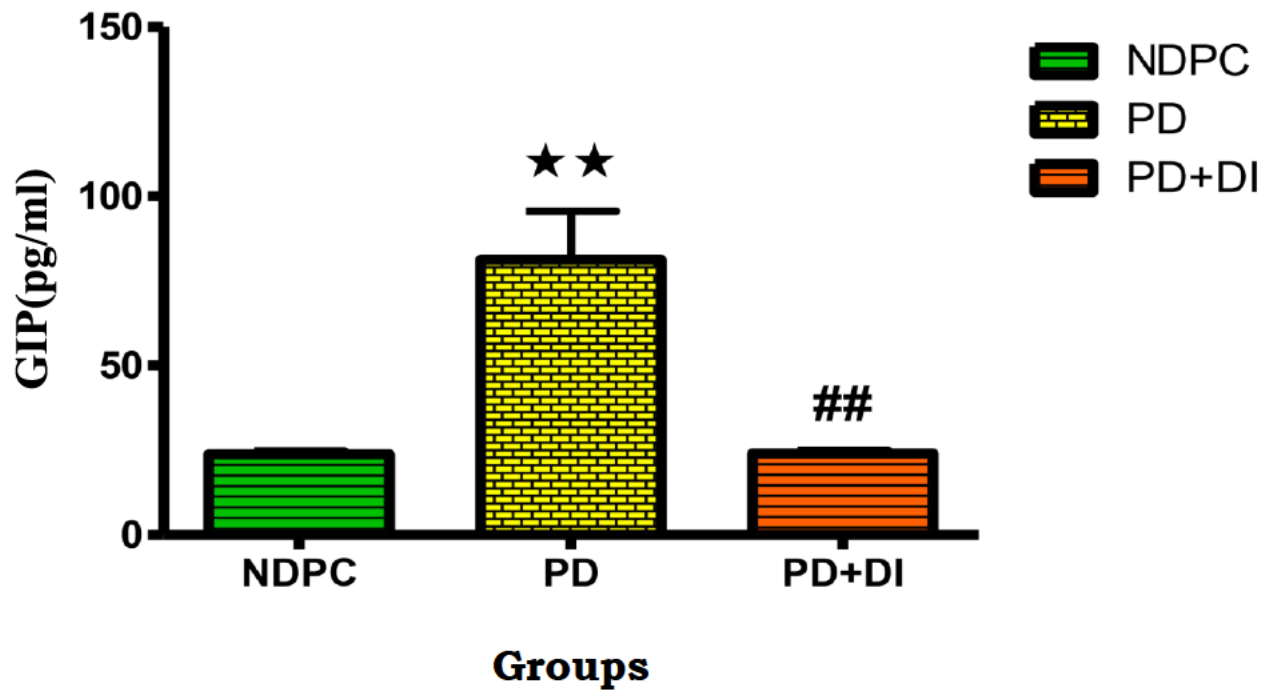


Figure 2: Plasma GIP concentrations in non-prediabetic control animals (NPDC), untreated pre-diabetic control animals (PD), as well as pre-diabetic animals that had a dietary intervention (PD +DI) for 12 weeks. Data expressed as means $\pm$ SEM ($n=6$). ★★ $p < 0.05$ denotes comparison with NPDC group; ## $p < 0.05$ denotes comparison with PD group.

3.2. Plasma GLP-1

The plasma GLP-1 concentrations were measured at the end of the experimental period at 32 weeks. The findings showed that compared to the NPDC group ($p=0.0019$) and the PD+DI group, the PD group had significantly lower plasma GLP-1 levels. The plasma GLP-1 concentration of the PD+DI group was lower than that of the NPDC group ($p=0.6111$) and higher than that of the PD group ($p=0.0199$).

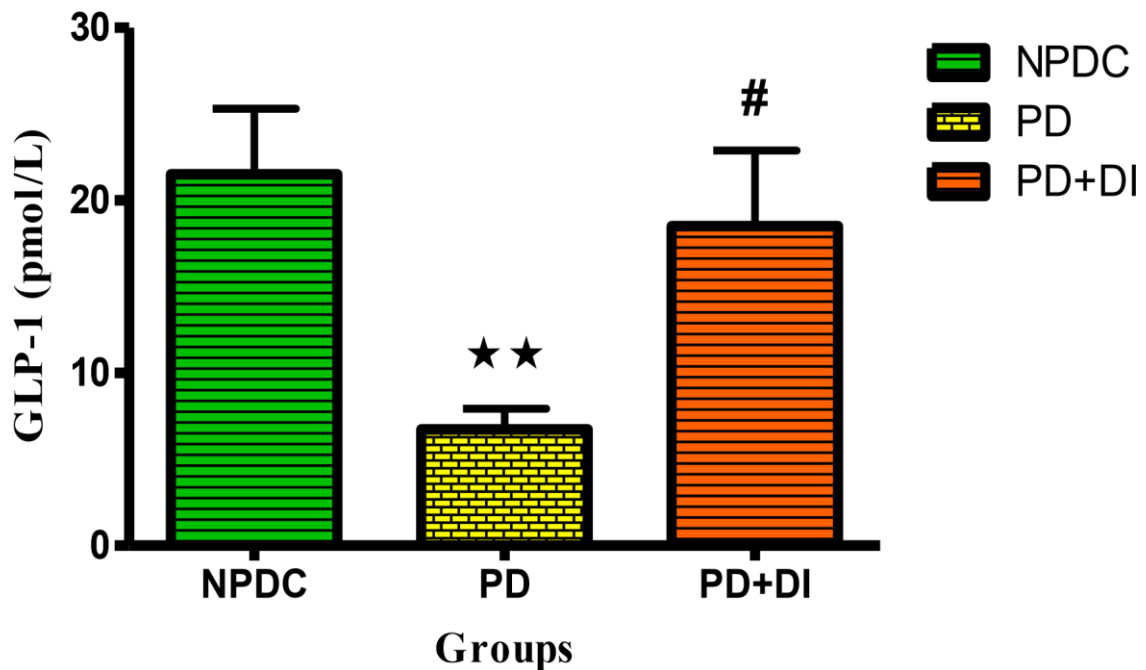


Figure 3: Plasma GLP-1 concentrations in non-prediabetic control animals (NPDC), untreated pre-diabetic control animals (PD), as well as pre-diabetic animals that had a dietary intervention (PD +DI) for 12 weeks. Data expressed as means $\pm$ SEM (n=6). ★★ $p < 0.05$ denotes comparison with NPDC group; # $p < 0.05$ denotes comparison with PD group.

3.3. Plasma Insulin

The plasma insulin concentrations were measured at the end of the experimental period at 32 weeks.

The results demonstrated that the PD group had significantly higher plasma insulin levels than the NPDC group ($p=0.0001$) and the PD+DI group. The PD+DI group's plasma insulin concentration was higher than the NPDC group ($p=0.2661$) and significantly lower than the PD group ($p=0.0066$).

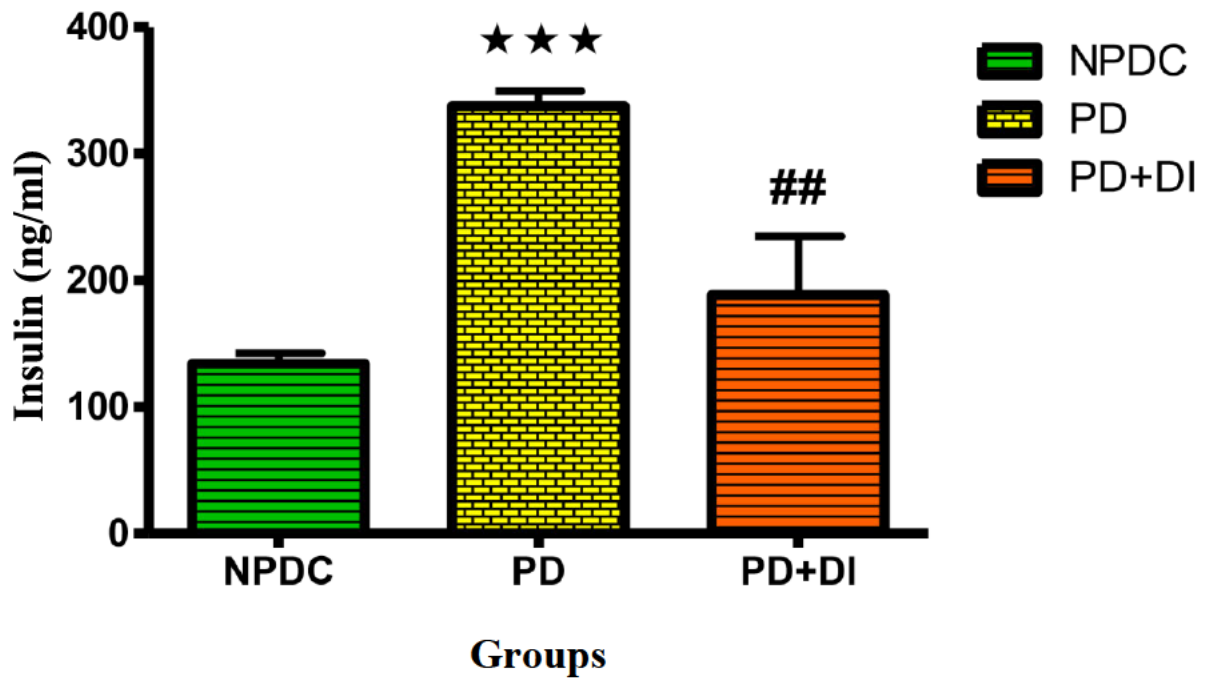


Figure 4: Plasma insulin concentrations in non-prediabetic control animals (NPDC), untreated prediabetic control animals (PD), as well as pre-diabetic animals that had a dietary intervention (PD +DI) for 12 weeks. Data expressed as means $\pm$ SEM ($n=6$). *** $p < 0.05$ denotes comparison with NPDC group; ## $p < 0.05$ denotes comparison with PD group.

3.4. Plasma Leptin

The plasma leptin concentrations were measured at the end of the experimental period at 32 weeks. The results demonstrated that the PD group had significantly lower plasma leptin levels than the NPDC group ($p=0.0001$) and the PD+DI group. The PD+DI group's plasma leptin concentration was higher than the PD group ($p=0.0212$) and significantly lower than the NPDC group ($p=0.2295$).

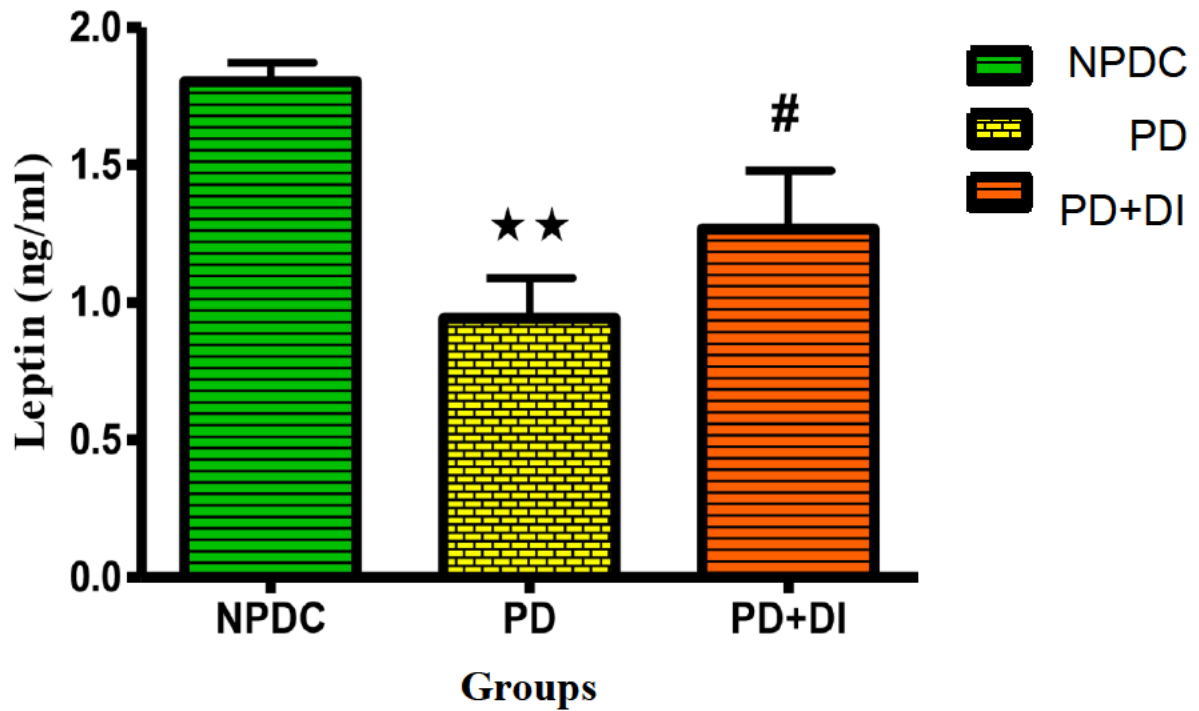


Figure 5: Plasma leptin concentrations in non-prediabetic control animals (ND), untreated pre-diabetic control animals (PD), as well as pre-diabetic animals that had a dietary intervention (PD +DI) for 12 weeks. Data expressed as means $\pm$ SEM ($n=6$). $\star\star p < 0.05$ denotes comparison with NPDC group; $\#p < 0.05$ denotes comparison with PD group.

3.5. Plasma Ghrelin

The plasma Ghrelin concentrations were measured at the end of the experimental period at 32 weeks.

The results demonstrated that the PD group had significantly higher plasma ghrelin levels than the NPDC group ($p=0.8647$) and the PD+DI group. The PD+DI group's plasma ghrelin concentration was slightly higher than the NPDC group ($p=0.8112$) and significantly lower than the PD group ($p=0.4644$).

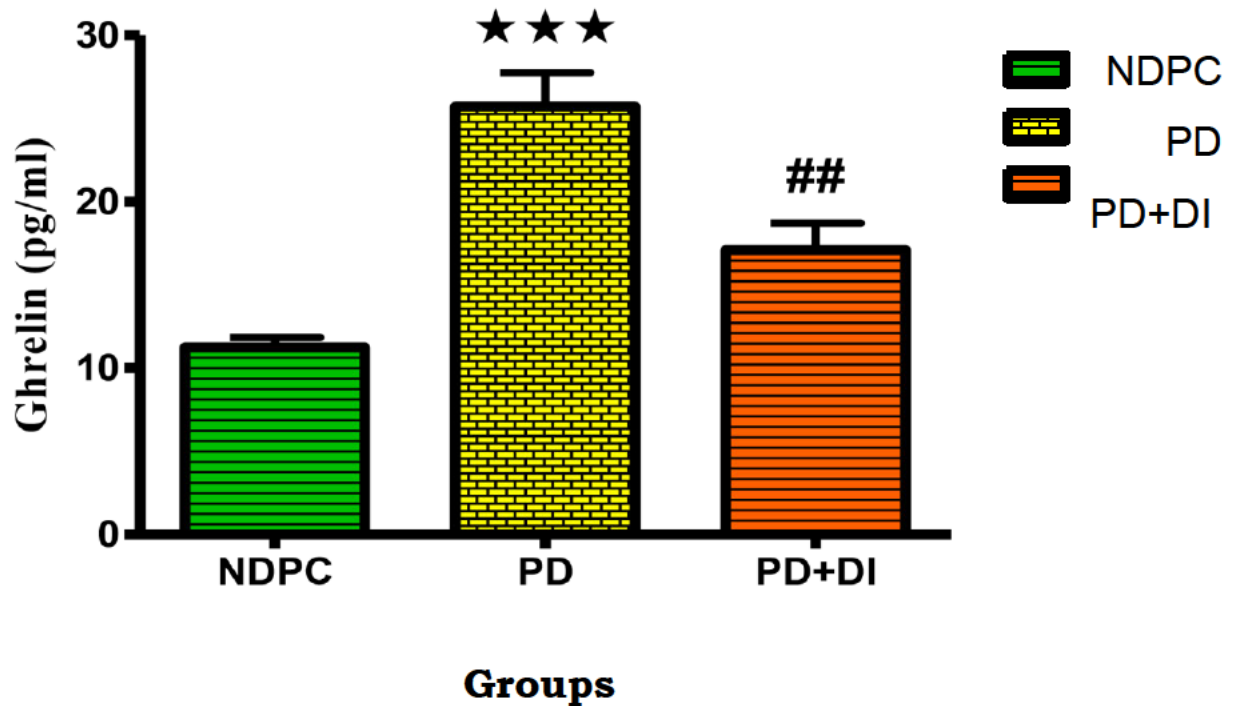


Figure 6: Plasma GLP-1 concentrations in non-prediabetic control animals (NPDC), untreated prediabetic control animals (PD), as well as pre-diabetic animals that had a dietary intervention (PD +DI) for 12 weeks. Data expressed as means $\pm$ SEM (n=6). *** $p < 0.05$ denotes comparison with NPDC group; ## $p < 0.05$ denotes comparison with PD group.

3.6. Plasma Glucose

The plasma glucose concentrations were measured at the end of the experimental period at 32 weeks.

The results demonstrated that the PD group had significantly higher plasma glucose levels than the NPDC group ($p=0.0010$) and the PD+DI group. The PD+DI group's plasma glucose concentration was significantly lower than both the NPDC group ($p=0.0010$) and the PD group ($p<0.0001$).

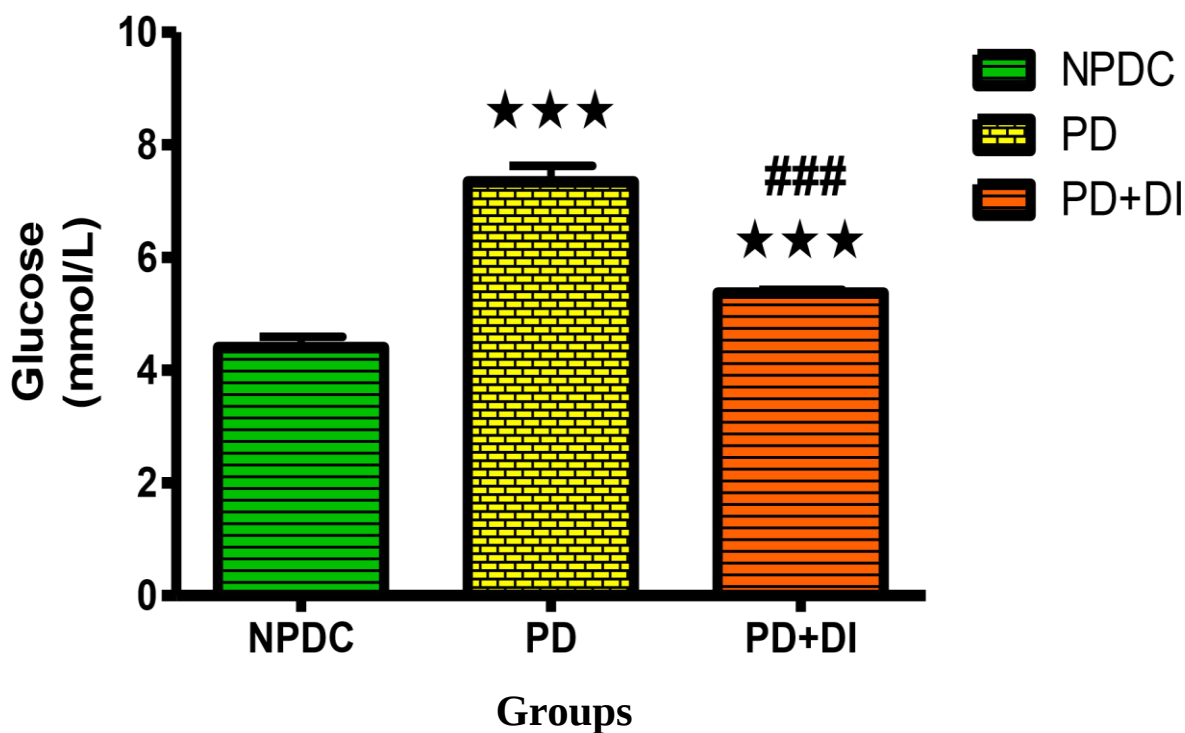


Figure 7: Plasma glucose concentrations in non-prediabetic control animals (NPDC), untreated prediabetic control animals (PD), as well as pre-diabetic animals that had a dietary intervention (PD +DI) for 12 weeks. Data expressed as means $\pm$ SEM ($n=6$). $***p < 0.05$ denotes comparison with NPD group; $###p < 0.05$ denotes comparison with PD group.

3.7. Plasma Glycated Hemoglobin (HbA1c).

The plasma HbA1c concentrations were measured after pre-diabetes induction at week 20 and the end of the experimental period at 32 weeks. The results demonstrated that the PD group had significantly higher HbA1c levels than the NPDC group ($p=0.0010$) and the PD+DI group. The PD+DI group's HbA1c levels were slightly lower with no significance compared to the NPDC Group ($p=0.0010$) and significantly lower than the PD group ($p<0.0001$).

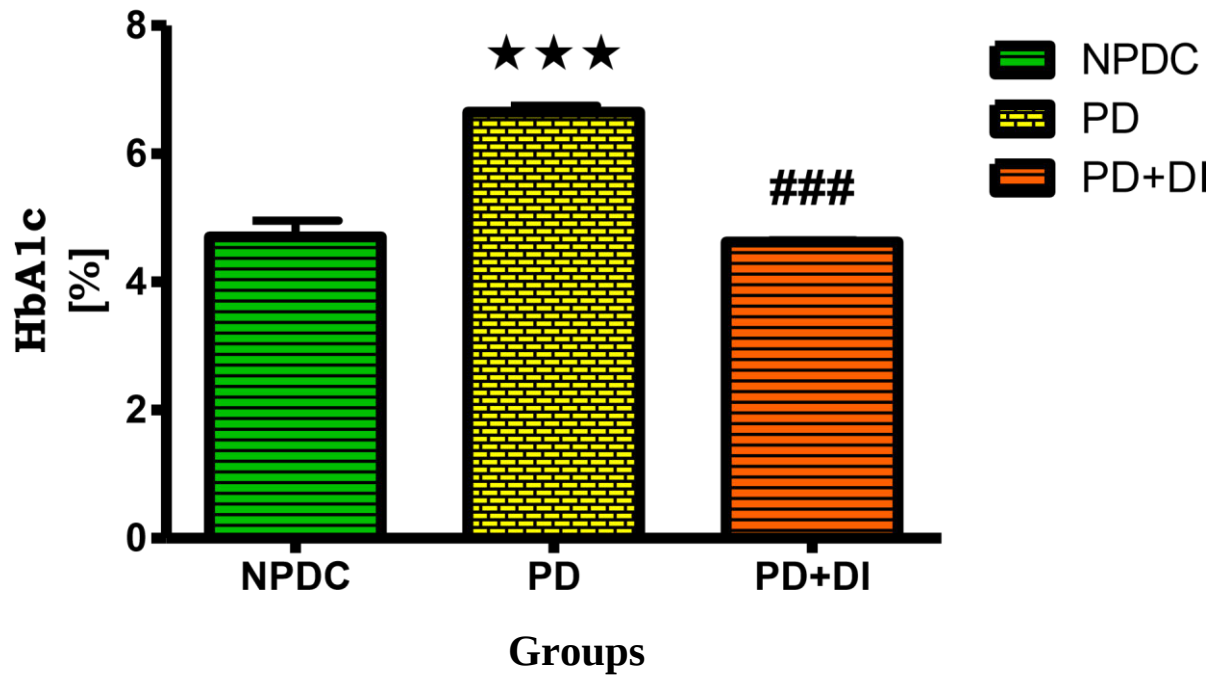


Figure 8: Plasma HbA1c concentrations in non-prediabetic control animals (NPDC), untreated prediabetic control animals (PD), as well as pre-diabetic animals that had a dietary intervention (PD +DI) for 12 weeks. Data expressed as means $\pm$ SEM (n=6). *** $p < 0.05$ denotes comparison with NPD group; ### $p < 0.05$ denotes comparison with PD group.

3.8. HOMA2-IR Index

The HOMA2-IR index was calculated in the NPD, PD and PD + DI groups at 32 weeks. The HOMA2IR value of the NPDC group was above the insulin-sensitivity range (<1.0) and below the early IR range (>1.9). By comparison with the NPDC group, the PD group showed a higher HOMA2-IR value, which was above the early IR range (>1.9) and the overt IR range (>2.9). The HOMA2-IR value of the PD + DI group was above the insulin-sensitivity range (<1.0) and below the early IR range (>1.9). (★ = IR range > 1.9).

GROUPS	SEM GLUCOSE	SEM INSULIN	p value Glucose	p value insulin	HOMA2-IR
NPDC	0.20000	8.465	0.0010	0.0001	1.08
PD	0.28000	11.74	0.0001	0.0066	2.99★
PD + DI	0.06455	46.37	0.0010	0.2661	1.58

Table 1: Displays the HOMA2-IR index calculated at week 32 using the HOMA2 calculator in the NPDC, PD and PD + DI groups (n=6 per group).

4. Discussion

Pre-diabetes is a long-lasting condition characterised by moderate hyperglycemia that often precedes the onset of type 2 diabetes mellitus. Literature evidence shows that a diet rich in saturated fat and refined carbohydrates is strongly linked to pre-diabetes development. About 5–10% of people with prediabetes go on to develop T2DM annually (58). Recently, several studies have shown that pre-diabetes is reversible, while the exact mechanism involved in the reversal of pre-diabetes has not yet been established (114, 115, 116). The pre-diabetic state has been identified as a target to reduce the onset of T2DM.

Several studies have shown that continued ingestion of a low-carbohydrate, high-fat diet is able to reverse the pre-diabetic state. The studies postulated that the reduced carbohydrate content in the LCHF diet yields a low glucose concentration that is absorbed into the bloodstream during digestion (117). The body is promoted to enhance fat oxidation to meet energy needs with reduced carbohydrate intake (117, 118, 119). Therefore, this results in a reduction in glycated hemoglobin levels and an improvement in glucose tolerance (118, 120). Our laboratory produced a rat model of diet-induced pre-diabetes by allowing chronic consumption of a high-fat, high-carbohydrate (HFHC) diet (81). According to a recent study from our laboratory, prolonged consumption of the HFHC diet leads to abnormal incretin concentrations, which may lead to the development of insulin resistance and the onset of pre-diabetes (129). To advance from the previous study, we sought to investigate the possibility of reversing pre-diabetes primarily using a LCHF diet to influence the incretin peptides.

Previous studies have established that chronic consumption of a high-fat and calorie diet causes elevated blood glucose concentrations for prolonged periods, resulting in the development of pre-diabetes (83). In pre-diabetes, there are high blood glucose concentrations for extended periods, and these elevated blood glucose concentrations are not high enough to be diagnosed as T2DM (83). A recent study in our laboratory established that abnormal concentrations of incretins precede pre-diabetes development, and incretins play a significant role in its development (129). Chronic HFHC diet consumption for a prolonged period leads to increased glucose absorption and chronic secretion of GIP and GLP-1 (121). GLP-1 degradation is increased as the dipeptidyl peptidase-4 enzyme concentration is also increased under high glucose conditions; this results in a severely reduced GLP-1 concentration (121). The elevated GIP constantly induces insulin secretion, resulting in elevated insulin over long periods (107). Over time, the insulin receptors lose their sensitivity to insulin and become resistant to insulin due to being overworked in the process of reducing the blood glucose concentration (83). The onset of insulin resistance results in fewer cells that receive glucose, and fewer cells receive glucose and send signals to the hypothalamus to suppress appetite (122). Therefore, there are reduced leptin concentrations and increased ghrelin concentrations; cells which do not receive glucose send signals to the hypothalamus to induce food intake by increasing appetite, which increases the secretion of ghrelin (75, 76). The increased secretion of ghrelin inhibits the leptin hormone's secretion, thus reducing satiety (75, 76). Leptin plays a role in the secretion of GLP-1; therefore, the secretion of GLP-1 is also impaired, which

further reduces its concentration (129). The incretins maintain these abnormalities, which leads to pre-diabetes, which can be confirmed by increased glycated hemoglobin concentration (129).

In the current study, we intervened by subjecting our pre-diabetic animal model to 12 weeks of LCHF diet consumption to reverse the effects of pre-diabetes. After 12 weeks, the significantly reduced blood glucose concentration confirmed the reversal of pre-diabetes. We speculate that the chronic consumption of a LCHF diet allowed for the absorption of a low amount of new glucose entering the bloodstream, which could have resulted in reduced blood glucose which could reduce the HbA1c concentration. The HbA1c concentration was indeed reduced to within the non-prediabetic range, indicating a restoration of glucose homeostasis.

Several hormones play a role in glucose homeostasis. GIP is a glucose-dependent peptide secreted shortly after meal consumption due to glucose in the stomach and its absorption into the bloodstream (107). It induces insulin secretion to reduce the upcoming blood glucose (107, 108). There are increased GIP concentrations in pre-diabetes due to prolonged elevated blood glucose concentration (74). Increased GIP concentration results in chronic insulin secretion, which causes hyperinsulinemia and later insulin resistance (74, 123). After chronic consumption of the LCHF diet, there are significantly reduced concentrations of GIP. We postulated that the secretion of GIP was reduced due to reduced blood glucose, which could also mean reduced plasma insulin concentrations. There were indeed significantly reduced plasma insulin concentrations after prolonged consumption of a LCHF diet. Taken together, the reduced blood glucose, insulin and HbA1c indicate improved insulin sensitivity. The homeostatic model assessment (HOMA) is a recognized method for measuring insulin resistance (IR) using fasting glucose and insulin (83, 124). HOMA-IR is calculated according to the formula: fasting plasma insulin ($\mu\text{U/ml}$) $\times$ fasting plasma glucose (mmol/L)/22.5 (83, 124). Using this method, we determined overt insulin resistance in the PD group, which fed on the HFHC diet for 32 weeks. After prolonged consumption of the LCHF, there was indeed improved insulin sensitivity in the PD + DI group.

In pre-diabetes, there are reduced leptin concentrations due to the developing insulin resistance (111, 125); The cells in tissues such as skeletal muscle does not receive glucose and send signals into the hypothalamus to increase appetite and food intake, which results in increased ghrelin concentrations (72). Increased ghrelin concentrations inhibit leptin secretion; therefore, there are reduced leptin concentrations (72). After chronic consumption of a LCHF diet, there was a significantly reduced ghrelin concentration and a significantly increased leptin concentration. We speculated that after the movement of glucose into the cells, the cells sent signals into the hypothalamus to inhibit appetite and induce satiety which could have resulted in a reduced ghrelin secretion and an increased leptin concentration. Leptin plays a role in the secretion of GLP-1; increased leptin could result in improved GLP-1 concentration (11). GLP-1 is secreted simultaneously with GIP after the entrance of glucose into the stomach and its absorption in the small intestines; there are reduced GLP-1 concentrations in prediabetes due to the increase of circulating DPP4 enzyme, which speeds up its degradation (73). The

prolonged blood glucose increases the circulating DPP4 enzyme (73). There are increased GLP-1 concentrations after chronic consumption of the LCHF diet. We speculate that due to reduced blood glucose, there could be reduced concentrations of the DPP4 enzyme, which could have allowed for a reduced degradation of GLP-1. Using the information gathered from the previous study, we speculated that the incretins abnormalities amelioration preceded the reversal of pre-diabetes as it was established that they precede pre-diabetes development (129).

Conclusion.

In this study, we observed the effect of diet on incretins as they play a significant role in developing and reversing pre-diabetes. Chronic consumption of a HFHC diet led to elevated blood glucose and insulin concentration, resulting in abnormal concentrations of incretins. The abnormal incretins then maintained this state of hyperglycemia and hyperinsulinemia, resulting in pre-diabetes. Chronic consumption of a LCHF by the pre-diabetic rats led to reduced concentrations of incretins, which could have led to a reduced HbA1c and eventually to the reversal of pre-diabetes.

Acknowledgments

The authors would like to acknowledge the endocrine group for their support, the Biomedical Resource Units of KwaZulu-Natal for their technical assistance, as well as the College of Health Sciences and the National Research Foundation (NRF) South Africa for their funding.

Author Contributions

NM and NRD contributed to the study design, conducted the experiments, collected, analyzed, and interpreted data, as well as being involved in writing the manuscript. AMS and PSN were involved in the study design, interpretation of data and editing of the manuscript. AK was involved in the conceptualization of the study, study design, analysis, and interpretation of data, writing and editing of the manuscript as well as provide funding. All authors have read and approved submission of the final manuscript.

Conflict of Interest

The authors declare no conflict of interest.

Availability of Data and Materials

The datasets presented in this study are included in the article and available on reasonable request from the corresponding author.

Ethics Approval and Consent to Participate

The animal study protocol was reviewed and approved by the Animal Research Ethics Committee (AREC) of the University of KwaZulu-Natal, South Africa (AREC/00003627/2021).

References

1. Toi PL, Anothaisintawee T, Chaikledkaew U, Briones JR, Reutrakul S, Thakkestian A. Preventive role of diet interventions and dietary factors in type 2 diabetes mellitus: an umbrella review. *Nutrients*. 2020;12(9):2722.
2. Cai X, Zhang Y, Li M, Wu JH, Mai L, Li J, et al. Association between prediabetes and risk of all cause mortality and cardiovascular disease: updated meta-analysis. *BMJ*. 2020;370:m2297.
3. Gamede M, Mabuza L, Ngubane P, Khathi A. The effects of plant-derived oleanolic acid on selected parameters of glucose homeostasis in a diet-induced pre-diabetic rat model. *Molecules*. 2018;23(4):794.
4. Stentz FB, Mikhael A, Kineish O, Christman J, Sands C. High protein diet leads to prediabetes remission and positive changes in incretins and cardiovascular risk factors. *Nutrition, Metabolism and Cardiovascular Diseases*. 2021;31(4):1227-37.
5. Mzimela NC, Ngubane PS, Khathi A. The changes in immune cell concentration during the progression of pre-diabetes to type 2 diabetes in a high-fat high-carbohydrate diet-induced pre-diabetic rat model. *Autoimmunity*. 2019;52(1):27-36.
6. Ha K, Joung H, Song Y. Inadequate fat or carbohydrate intake was associated with an increased incidence of type 2 diabetes mellitus in Korean adults: a 12-year community-based prospective cohort study. *Diabetes research and clinical practice*. 2019;148:254-61.
7. Röhling M, Kempf K, Banzer W, Berg A, Braumann K-M, Tan S, et al. Prediabetes conversion to normoglycemia is superior adding a low-carbohydrate and energy deficit formula diet to lifestyle intervention—A 12-month subanalysis of the ACOORH trial. *Nutrients*. 2020;12(7):2022.
8. Gastaldello A, Giampieri F, De Giuseppe R, Grosso G, Baroni L, Battino M. The rise of processed meat alternatives: A narrative review of the manufacturing, composition, nutritional profile and health effects of newer sources of protein, and their place in healthier diets. *Trends in Food Science & Technology*. 2022;127:263-71.
9. Gastaldello A, Giampieri F, Quiles JL, Navarro-Hortal MD, Aparicio S, García Villena E, et al. Adherence to the Mediterranean-Style Eating Pattern and Macular Degeneration: A Systematic Review of Observational Studies. *Nutrients*. 2022;14(10):2028.
10. Nauck MA, Meier JJ. Incretin hormones: Their role in health and disease. *Diabetes, Obesity and Metabolism*. 2018;20:5-21.
11. Campbell Jonathan E, Drucker Daniel J. Pharmacology, Physiology, and Mechanisms of Incretin Hormone Action. *Cell Metabolism*. 2013;17(6):819-37.

12. Nauck MA, Meier JJ. The incretin effect in healthy individuals and those with type 2 diabetes: physiology, pathophysiology, and response to therapeutic interventions. *The Lancet Diabetes & Endocrinology*. 2016;4(6):525-36.
13. Yaribeygi H, Maleki M, Atkin SL, Jamialahmadi T, Sahebkar A. Impact of IncretinBased Therapies on Adipokines and Adiponectin. *Journal of Diabetes Research*. 2021;2021:3331865.
14. Stoica L, Gadea R, Navolan D-B, Lazar F, Duta C, Stoian D, et al. Plasma ghrelin, adiponectin and leptin levels in obese rats with type 2 diabetes mellitus after sleeve gastrectomy and gastric plication. *Experimental and Therapeutic Medicine*. 2021;21(3):1-.
15. Boer GA, Holst JJ. Incretin hormones and type 2 diabetes—mechanistic insights and therapeutic approaches. *Biology*. 2020;9(12):473.
16. Forouhi NG, Misra A, Mohan V, Taylor R, Yancy W. Dietary and nutritional approaches for prevention and management of type 2 diabetes. *Bmj*. 2018;361.
17. Sosibo AM. Investigating changes to the insulin signalling pathway in a diet-induced pre-diabetic rat model: effects on selected markers 2019.
18. Tabák AG, Herder C, Rathmann W, Brunner EJ, Kivimäki M. Prediabetes: a high-risk state for diabetes development. *Lancet*. 2012;379(9833):2279-90.
19. Amer OE, Sabico S, Alfawaz HA, Aljohani N, Hussain SD, Alnaami AM, et al. Reversal of Prediabetes in Saudi Adults: Results from an 18 Month Lifestyle Intervention. *Nutrients*. 2020;12(3):804.
20. Liu X, Wu S, Song Q, Xizhu W. Reversion From Pre-Diabetes Mellitus to Normoglycemia and Risk of Cardiovascular Disease and All-Cause Mortality in a Chinese Population: A Prospective Cohort Study. *Journal of the American Heart Association*. 2021;10.
21. Vistisen D, Kivimäki M, Perreault L, Hulman A, Witte DR, Brunner EJ, et al. Reversion from prediabetes to normoglycaemia and risk of cardiovascular disease and mortality: the Whitehall II cohort study. *Diabetologia*. 2019;62(8):1385-90.
22. Joshi S, Ostfeld RJ, McMacken M. The ketogenic diet for obesity and diabetes—enthusiasm outpaces evidence. *JAMA internal medicine*. 2019;179(9):1163-4.
23. Astrup A, Larsen TM, Harper A. Atkins and other low-carbohydrate diets: hoax or an effective tool for weight loss? *The Lancet*. 2004;364(9437):897-9.
24. Crowe T. Safety of low-carbohydrate diets. *Obesity reviews*. 2005;6(3):235-45.
25. Noakes TD. Low-carbohydrate and high-fat intake can manage obesity and associated conditions: Occasional survey. *South African Medical Journal*. 2013;103(11):826-30.
26. Seino Y, Fukushima M, Yabe D. GIP and GLP-1, the two incretin hormones: similarities and differences. *Journal of diabetes investigation*. 2010;1(1-2):8-23.

27. Ghadge AA, Khaire AA. Leptin as a predictive marker for metabolic syndrome. *Cytokine*. 2019;121:154735.
28. Shankar K, Takemi S, Gupta D, Varshney S, Mani BK, Osborne-Lawrence S, et al. Ghrelin cell-expressed insulin receptors mediate meal- and obesity-induced declines in plasma ghrelin. *JCI Insight*. 2021;6(18).
29. Omran DM, Alaraji SM, Albayati AH, Essam W. Relationship between Ghrelin and Leptin with Insulin Resistance in Obese Patients and Non-Obese Individuals. *Research Journal of Pharmacy and Technology*. 2018;11(1):281-3.
30. Fu Y, Kaneko K, Lin H-Y, Mo Q, Xu Y, Suganami T, et al. Gut Hormone GIP Induces Inflammation and Insulin Resistance in the Hypothalamus. *Endocrinology*. 2020;161(9).
31. Camilleri M. Gastrointestinal hormones and regulation of gastric emptying. *Curr Opin Endocrinol Diabetes Obes*. 2019;26(1):3-10.
32. Demir AK, Şahin Ş, Kaya SU, Bütün İ, Çıtlı R, Önder Y, et al. Prevalence of insulin resistance and identifying HOMA1-IR and HOMA2-IR indexes in the Middle Black Sea region of Turkey. *African Health Sciences*. 2020;20(1):277-86.
33. Bungau S, Behl T, Tit DM, Banica F, Bratu OG, Diaconu CC, et al. Interactions between leptin and insulin resistance in patients with prediabetes, with and without NAFLD. *Experimental and Therapeutic Medicine*. 2020;20(6):1-.
34. Van Doorn C, Macht VA, Grillo CA, Reagan LP. Leptin resistance and hippocampal behavioral deficits. *Physiology & behavior*. 2017;176:207-13.
35. Ronveaux CC, Tomé D, Raybould HE. Glucagon-like peptide 1 interacts with ghrelin and leptin to regulate glucose metabolism and food intake through vagal afferent neuron signaling. *J Nutr*. 2015;145(4):672-80.
36. Nauck MA, Quast DR, Wefers J, Meier JJ. GLP-1 receptor agonists in the treatment of type 2 diabetes – state-of-the-art. *Molecular Metabolism*. 2021;46:101102.

Chapter 4: Synthesis and Conclusion

Synthesis

Pre-diabetes is a condition of intermediate hyperglycemia that often precedes the onset of type 2 diabetes mellitus (T2DM) (70). In this condition, we observe chronically elevated blood glucose concentrations that are not high enough for a diagnosis of T2DM (70, 81). Several studies report that often life-threatening complications associated with T2DM begin in the pre-diabetic state (69, 78). While T2DM can be managed, it has no known cure, while pre-diabetes has been shown to be reversible (126, 127). Therefore, pre-diabetes has become a therapeutic target to prevent the onset of T2DM (70). Incretins are hormones secreted in the gut that aid in maintaining glucose homeostasis by enhancing insulin secretion as they contribute significantly to overall postprandial insulin release (8). Incretin peptides include glucose-dependent insulinotropic polypeptide (GIP) and glucagon-like peptide 1 (GLP-1), associated with ghrelin and leptin. Hyperglycemia in T2DM has been observed to impair the normal functioning of incretins, resulting in abnormal incretin concentrations (20, 73). Furthermore, abnormal incretins have been shown to play a role in the pathogenesis of T2DM (20). However, the changes in incretins in the pre-diabetic state have not been established. Hence, the first manuscript of this study aimed to investigate the effect of incretin concentrations in the development of diet-induced pre-diabetes. The Second manuscript of this study sought to investigate the effect of a low carbohydrate, high unsaturated fat (LCHF) diet on incretin concentrations in a diet-induced pre-diabetic animal model as this diet has been shown to be beneficial in the management of prediabetes.

Incretin peptides are associated with the appetite-regulating hormones, together they aid in glucose homeostasis. Several studies have shown that abnormal incretin concentrations play a role in the progression of the pathology of T2DM (20, 121). However, minimal work has been done in assessing the link between incretin peptide levels and pre-diabetes development. In our laboratory, a pre-diabetic animal model mimicking the human pre-diabetic state has been developed (64, 82). Previous studies on this model have established that pre-diabetes is induced in male Sprague Dawley rats after 20 weeks of a chronic high-fat, high-carbohydrate diet consumption (82, 83). This model has been used to observe changes that occur in the pre-diabetic state. However, no work had been done to explore the changes in incretin concentration and their possible involvement in the development of pre-diabetes as well as the effect of a low carbohydrate, high unsaturated fat diet in incretin levels of a prediabetic animal model. This led to manuscript 1, where we induced pre-diabetes using a HFHC diet to observe the involvement of incretins in the development of pre-diabetes, and manuscript 2.

The first manuscript separated twenty-four (24) animals separated into two randomized groups: group 1 (n=12) and group 2 (n=12). Group 1 was kept on a standard rat diet (SD) while group 2 was kept on the experimental high-fat, high carbohydrates diet (HFHC) (n=6). The animals were maintained on their respective diets for 20 weeks. After 10 weeks being on their respective diets, 6 animals were sacrificed from each group and biochemical analysis was done. The rest of the animals continued in the study for

an additional 10 weeks while continuing their respective diets. At week 20, all the remaining animals in each group were sacrificed and biochemical analysis was carried out.

In line with previous studies, the results of this study showed that ingestion of a HFHC diet for 20 weeks resulted in elevated plasma glucose and insulin levels and increased HbA1c and HOMA-IR compared to the SD group. These results suggested that the rats fed the HFHC diet had indeed developed pre-diabetes (PD) while those on the standard (SD) diet remained non-pre-diabetic. Additionally, at week 20, the results depicted significantly higher concentrations of ghrelin and GIP with significantly reduced leptin and GLP-1 concentrations, suggesting a derangement in incretin homeostasis during the prediabetic state.

Of interest to this study, at 10 weeks, the results showed that ingestion of a HFHC diet resulted in elevated plasma glucose but other markers such as, plasma insulin, HbA1c and HOMA-IR showed that the animals had not developed prediabetes. Furthermore, at 10 weeks, the results showed significantly high plasma ghrelin, and GIP concentrations as well as significantly reduced leptin and GLP-1 thus suggesting a derangement in incretin homeostasis before the onset of pre-diabetes. The chronic consumption of the HFHC diet could have led to hyperglycemia which may have led to constant secretion of GIP and GLP-1 due to the presence of glucose in the stomach and its absorption. We speculate that due to hyperglycemia, there could have been an increase in circulating DPP-4 concentration, leading to increased degradation and reduced concentrations of GLP-1. Chronic GIP secretion could be responsible for hyperinsulinaemia due to the constant induction of insulin secretion; This may have led to the development of insulin resistance. Increased circulating ghrelin could be attributed to the presence of insulin resistance. Tissue cells with insulin-resistant receptors could not have been able to receive glucose. They could have constantly sent signals to the hypothalamus to induce food intake, which could have resulted in the constant secretion of ghrelin. Increased levels of ghrelin have been shown to lead to reduced leptin concentrations (76); Which could be the reason for the significantly reduced levels of leptin. Increased ghrelin concentration could have led to increased food intake, further increasing blood glucose. Additionally, the reduced leptin concentrations could have also been due to the reduced number of tissue cells that receive glucose and send signals to the hypothalamus to induce satiety, leading to leptin secretion. Leptin also plays a role in the secretion of GLP-1 (128); This could mean a reduction in GLP-1 secretion. This study, for the first time suggests that chronic ingestion of a HFHC could possibly dysregulate plasma incretins and that if this is maintained for extended periods, could have led to the development of pre-diabetes. The findings of this manuscript suggests that incretins play a role in the development of pre-diabetes.

To advance on this knowledge, the second manuscript sought to investigate the possibility of reversing pre-diabetes primarily using a LCHF diet to influence the incretin peptides as their concentrations are directly affected by diet. In a previous study, it has been postulated that the reduced carbohydrate content in the LCHF diet yields a low glucose concentration that is absorbed into the bloodstream during digestion (117). For this study, we had six (6) non-prediabetic animals that were kept on a standard rat

diet. These animals served as the non-prediabetic control (NPC) group. We also induced prediabetes in twelve (12) male Sprague Dawley rats using the HFHC diet. After induction, the animals were split into two experimental groups (n=6 in each group). One of the groups continued a HFHC diet and served as the pre-diabetic control group (PD). The other group was changed to a low carbohydrate-high unsaturated fats diet (PD+DI). Both groups were then maintained on their respective diets for an additional 12 weeks. Following the additional 12 weeks, all three groups were sacrificed, and biochemical analysis was performed.

By comparison to the non-prediabetic control, the elevated plasma glucose, insulin, HOMA-IR index, and HbA1c in the PD group suggested that there was overt insulin resistance thus confirming that the prediabetes had progressed. However, by comparison to the prediabetic control group, the PD+DI group had improved incretin concentration, reduced plasma insulin, reduced blood glucose, and reduced HbA1c concentrations suggesting a reversal of prediabetes. These findings were in agreement with several studies that have shown that continued ingestion of a low-carbohydrate, high-fat diet is able to reverse the pre-diabetic state. These studies postulated that the reduced carbohydrate content in the LCHF diet yields a low glucose concentration that is absorbed into the bloodstream during digestion (117). The body is promoted to enhance fat oxidation to meet the energy needs with reduced carbohydrate intake (117, 118, 119). Therefore, this results in a reduction in glycated hemoglobin levels and an improvement in glucose tolerance (118, 120).

Based on the findings of manuscript 1, which showed that changes in incretin levels precede changes in the onset of prediabetes, we suggest that chronic consumption of the LCHF diet could have resulted in the reversal of prediabetes, in part, by modulating incretin levels. In pre-diabetes, GIP is responsible for the constant secretion of insulin, a reduced GIP secretion could therefore result in reduced stimulation of insulin secretion which could allow for the circulating insulin to be degraded over time without a lot of new insulin being secreted into the bloodstream. The gradual degradation of insulin could be responsible for reduced blood glucose as it facilitates its absorption into tissue cells. The reduced blood glucose could have led to a reduced plasma DPP-4, resulting in the increased plasma GLP-1 observed after 12 weeks. The absorption of blood glucose into tissue cells could have also resulted in the secretion of leptin and inhibition of ghrelin induced by tissue cells sending signals into the hypothalamus to inhibit food intake and induce satiety. The reduced blood glucose, plasma insulin, and HOMA-IR index suggest improved insulin sensitivity. After 12 weeks, the significantly reduced blood glucose concentration and reduced HbA1c confirmed the reversal of pre-diabetes. Therefore, for the first time, this study suggests that incretins play a significant role in the mechanism to reverse pre-diabetes.

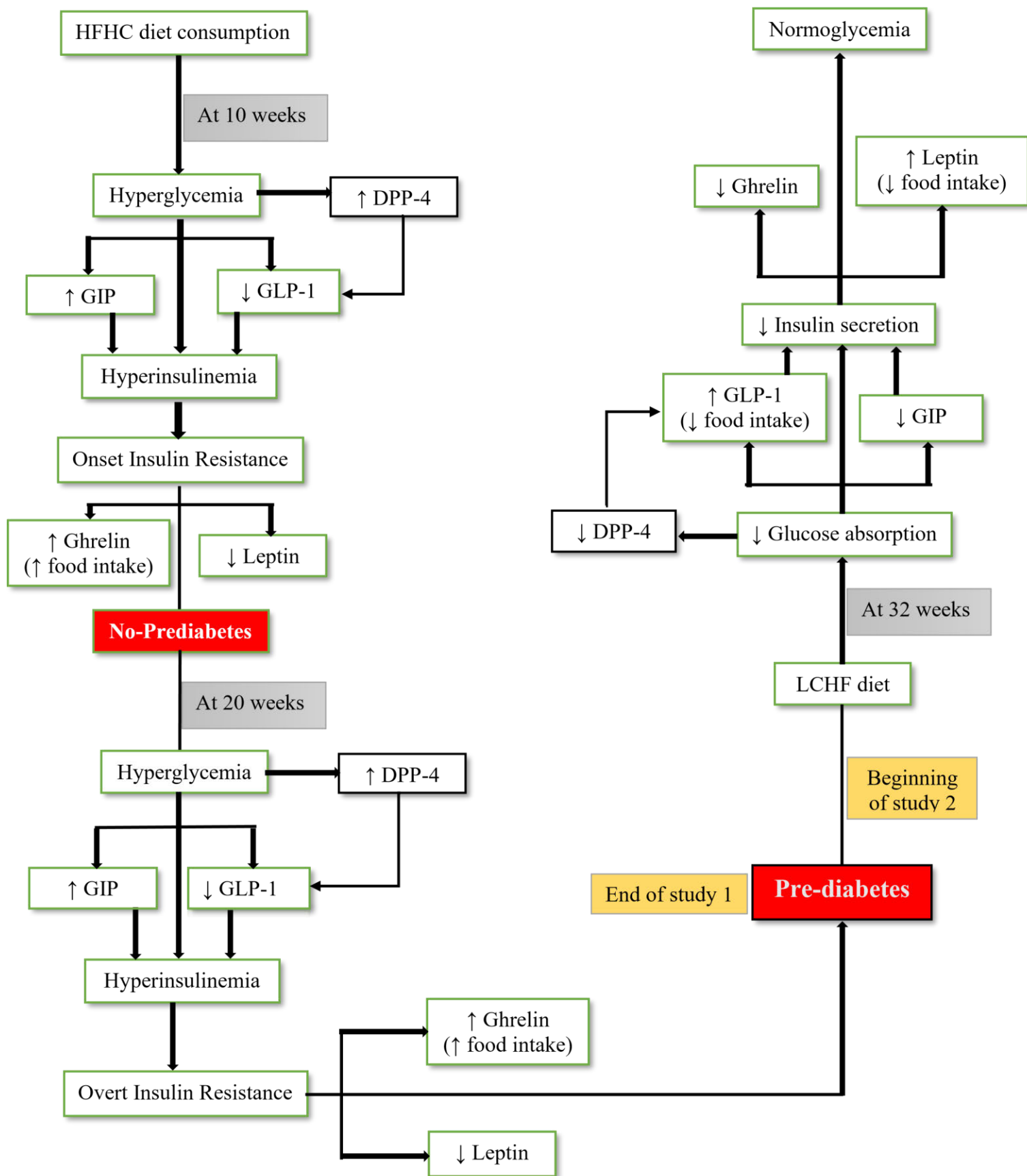


Figure 1: Diagram illustrating the role of incretins in developing and reversing pre-diabetes.

Conclusion

Taken together, the findings of this study revealed that incretin concentrations are indeed influenced by diet. Hence, the dysregulation of incretin levels may contribute to the onset of pre-diabetes. This study suggested, for the first time, that incretin abnormalities precede and significantly contribute to pre-diabetes development. This research also suggested that incretins significantly impact the reversal of pre-diabetes. The research may help identify specific incretin hormone concentration patterns or changes that are associated with the development of diet-induced prediabetes. Identifying biomarkers can aid in targeted interventions and preventive measures for at-risk individuals. This may open a new avenue of drugs in the management of prediabetes that seek to modulate the functioning of incretin levels.

Shortfalls and Future Studies

In manuscript 1, we looked at the involvement of incretins in the development of diet-induced prediabetes. This study could have further looked at the DPP-4 enzyme pathway, as it plays a major role in the regulation of incretin levels; however, due to budget constraints, this was not possible. Therefore, future studies should investigate the involvement of DPP-4 in the development of prediabetes. Some studies show conflicting results to our study showing normal, increased, and decreased GLP-I and GIP levels in individuals with prediabetes which highlights, we therefore recommend more studies on incretins, specifically in animal models without any confounding factors.

In manuscript 2, we looked at the effect a LCHF diet has on incretin levels, hoping for a possibility of reversing pre-diabetes primarily using the LCHF diet to influence the incretin peptides. This study could have looked at the incretin levels at the halfway point into the reversal of prediabetes. This could have shown conclusively whether changes to incretin levels precede the reversal of prediabetes. Therefore, future studies should investigate the effect of a LCHF diet on the incretin concentrations at 6 weeks, halfway to the reversal of prediabetes to conclusively show their involvement in the reversal of prediabetes.

References

1. Güemes M, Rahman SA, Hussain K. What is a normal blood glucose? Archives of disease in childhood. 2016;101(6):569-74.
2. Holst JJ, Holland W, Gromada J, Lee Y, Unger RH, Yan H, et al. Insulin and glucagon: partners for life. Endocrinology. 2017;158(4):696-701.
3. Qaid MM, Abdelrahman MM. Role of insulin and other related hormones in energy metabolism—A review. Cogent Food & Agriculture. 2016;2(1):1267691.
4. Wang T, Wang J, Hu X, Huang X-J, Chen G-X. Current understanding of glucose transporter 4 expression and functional mechanisms. World journal of biological chemistry. 2020;11(3):76.
5. Klip A, McGraw TE, James DE. Thirty sweet years of GLUT4. Journal of Biological Chemistry. 2019;294(30):11369-81.
6. Tengholm A, Gylfe E. cAMP signalling in insulin and glucagon secretion. Diabetes, obesity and metabolism. 2017;19:42-53.
7. Abdulkader F. Metabolic Pathways and Cell Signaling. Essential Aspects of Immunometabolism in Health and Disease: Springer; 2022. p. 13-30.
8. Wu T, Rayner CK, Horowitz M. Incretins. Metabolic Control. 2015:137-71.
9. Frost F, Jones GH, Dyce P, Jackson V, Nazareth D, Walshaw MJ. Loss of incretin effect contributes to postprandial hyperglycaemia in cystic fibrosis-related diabetes. Diabetic Medicine. 2019;36(11):1367-74.
10. Lim GE, Brubaker PL. Glucagon-Like Peptide 1 Secretion by the L-Cell. Diabetes. 2006;55(Supplement 2):S70.
11. Ronveaux CC, Tomé D, Raybould HE. Glucagon-like peptide 1 interacts with ghrelin and leptin to regulate glucose metabolism and food intake through vagal afferent neuron signaling. J Nutr. 2015;145(4):672-80.
12. Skuratovskaia D, Vulf M, Chasovskikh N, Komar A, Kirienkova E, Shunkin E, et al. The links of ghrelin to incretins, insulin, glucagon, and leptin after bariatric surgery. Frontiers in genetics. 2021;12.
13. Rocha S, Corvo ML, Fernandes E, Freitas M. The Emerging Target Protein Tyrosine Phosphatase 1B (PTP1B) for Type 2 Diabetes Mellitus Management. 2021.
14. Mkhize B, Mosili P, Ngubane P, Sibiya N, Khathi A. Diet-induced prediabetes: Effects on the systemic and renal renin-angiotensin-aldosterone system. 2020.
15. Lynggaard MB, Gasbjerg LS, Christensen MB, Knop FK. GIP (3-30) NH₂—a tool for the study of GIP physiology. Current Opinion in Pharmacology. 2020;55:31-40.

16. Baggio LL, Drucker DJ. Biology of incretins: GLP-1 and GIP. *Gastroenterology*. 2007;132(6):2131-57.
17. Joo E, Harada N, Yamane S, Fukushima T, Taura D, Iwasaki K, et al. Inhibition of gastric inhibitory polypeptide receptor signaling in adipose tissue reduces insulin resistance and hepatic steatosis in high-fat diet-fed mice. *Diabetes*. 2017;66(4):868-79.
18. Sun Y, Asnicar M, Saha PK, Chan L, Smith RG. Ablation of ghrelin improves the diabetic but not obese phenotype of ob/ob mice. *Cell metabolism*. 2006;3(5):379-86.
19. Gupta K, Raja A. Physiology, Gastric Inhibitory Peptide. StatPearls [Internet]: StatPearls Publishing; 2020.
20. Zhou Y, Rui L. Leptin signaling and leptin resistance. *Frontiers of medicine*. 2013;7(2):207-22.
21. Izadi V, Saraf-Bank S, Azadbakht L. Dietary intakes and leptin concentrations. *ARYA Atheroscler*. 2014;10(5):266-72.
22. Kolodziejski PA, Sassek M, Chalupka D, Leciejewska N, Nogowski L, Mackowiak P, et al. GLP1 and GIP are involved in the action of synbiotics in broiler chickens. *Journal of animal science and biotechnology*. 2018;9(1):1-9.
23. Lim GE, Brubaker PL. Glucagon-Like Peptide 1 Secretion by the L-Cell. The View From Within. 2006;55(Supplement 2):S70-S7.
24. Zhang Y, Liu Y, Xu J, Sun Q, Yu F, Cheng J, et al. Inhibition of DPP4 enhances inhibitory synaptic transmission through activating the GLP-1/GLP-1R signaling pathway in a rat model of febrile seizures. *Biochemical Pharmacology*. 2018;156:78-85.
25. Müller TD, Finan B, Bloom SR, D'Alessio D, Drucker DJ, Flatt PR, et al. Glucagonlike peptide 1 (GLP-1). *Molecular Metabolism*. 2019;30:72-130.
26. Ronveaux CC, Tomé D, Raybould HE. Glucagon-like peptide 1 interacts with ghrelin and leptin to regulate glucose metabolism and food intake through vagal afferent neuron signaling. *The Journal of nutrition*. 2015;145(4):672-80.
27. Andersen A, Lund A, Knop FK, Vilsbøll T. Glucagon-like peptide 1 in health and disease. *Nature Reviews Endocrinology*. 2018;14(7):390-403.
28. Hosoda H, Kojima M, Matsuo H, Kangawa K. Ghrelin and des-acyl ghrelin: two major forms of rat ghrelin peptide in gastrointestinal tissue. *Biochemical and biophysical research communications*. 2000;279(3):909-13.
29. Lv Y, Liang T, Wang G, Li Z. Ghrelin, a gastrointestinal hormone, regulates energy balance and lipid metabolism. *Bioscience reports*. 2018;38(5).

30. Poher A-L, Tschöp MH, Müller TD. Ghrelin regulation of glucose metabolism. *Peptides*. 2018;100:236-42.
31. Sun Y, Wang P, Zheng H, Smith RG. Ghrelin stimulation of growth hormone release and appetite is mediated through the growth hormone secretagogue receptor. *Proceedings of the National Academy of Sciences*. 2004;101(13):4679-84.
32. Suyama S, Yada T. New insight into GABAergic neurons in the hypothalamic feeding regulation. *The Journal of Physiological Sciences*. 2018;68(6):717-22.
33. Akalu Y, Molla MD, Dessie G, Ayelign B. Physiological effect of ghrelin on body systems. *International journal of endocrinology*. 2020;2020.
34. Nakazato M, Murakami N, Date Y, Kojima M, Matsuo H, Kangawa K, et al. A role for ghrelin in the central regulation of feeding. *Nature*. 2001;409(6817):194-8.
35. Blázquez E, Velázquez E, Hurtado-Carneiro V, Ruiz-Albusac JM. Insulin in the brain: its pathophysiological implications for States related with central insulin resistance, type 2 diabetes and Alzheimer's disease. *Frontiers in endocrinology*. 2014;5:161.
36. Satoh N, Ogawa Y, Katsuura G, Tsuji T, Masuzaki H, Hiraoka J, et al. Pathophysiological significance of the obese gene product, leptin, in ventromedial hypothalamus (VMH)-lesioned rats: evidence for loss of its satiety effect in VMH-lesioned rats. *Endocrinology*. 1997;138(3):947-54.
37. Park H-K, Ahima RS. Leptin signaling. *F1000Prime Rep*. 2014;6:73-.
38. Irwin N, Gault VA, O'Harte FP, Flatt PR. Blockade of gastric inhibitory polypeptide (GIP) action as a novel means of countering insulin resistance in the treatment of obesitydiabetes. *Peptides*. 2020;125:170203.
39. Sarkar J, Nargis T, Tantia O, Ghosh S, Chakrabarti P. Increased Plasma Dipeptidyl Peptidase-4 (DPP4) Activity Is an Obesity-Independent Parameter for Glycemic Deregulation in Type 2 Diabetes Patients. *Front Endocrinol (Lausanne)*. 2019;10:505-.
40. David L. 2019 ESC Guidelines on diabetes, pre-diabetes, and cardiovascular diseases developed in collaboration with the EASD. *European heart journal*. 2020;41(2):255-323.
41. DeFronzo RA, Ferrannini E, Groop L, Henry RR, Herman WH, Holst JJ, et al. Type 2 diabetes mellitus. *Nature reviews Disease primers*. 2015;1(1):1-22.
42. Freeman AM, Pennings N. Insulin resistance. *StatPearls [Internet]*. 2021.
43. Sultan E, Taha I, Saber LM. Altered bone metabolic markers in type 2 diabetes mellitus: impact of glycemic control. *Journal of Taibah University Medical Sciences*. 2008;3(2):104-16.
44. Rydén L, Grant PJ, Anker SD, Berne C, Cosentino F, Danchin N, et al. Erratum: ESC Guidelines on diabetes, pre-diabetes, and cardiovascular diseases

- developed in collaboration with the EASD (European Heart Journal (2013) 34 (3035-3087. European Heart Journal. 2014;35(27):1824.
45. Khanduker S, Ahmed R, Khondker F, Aharama A, Afrose N, Chowdhury MAA. Electrolyte Disturbances in Patients with Diabetes Mellitus. Bangladesh Journal of Medical Biochemistry. 2017;10(1):27-35.
 46. Cosentino F, Grant PJ, Aboyans V, Bailey CJ, Ceriello A, Delgado V, et al. 2019 ESC Guidelines on diabetes, pre-diabetes, and cardiovascular diseases developed in collaboration with the EASD: The Task Force for diabetes, pre-diabetes, and cardiovascular diseases of the European Society of Cardiology (ESC) and the European Association for the Study of Diabetes (EASD). European heart journal. 2020;41(2):255-323.
 47. Pederson RA, McIntosh CH. Discovery of gastric inhibitory polypeptide and its subsequent fate: Personal reflections. J Diabetes Investig. 2016;7 Suppl 1(Suppl 1):4-7.
 48. Deloose E, Verbeure W, Depoortere I, Tack J. Motilin: from gastric motility stimulation to hunger signalling. Nat Rev Endocrinol. 2019;15(4):238-50.
 49. Mani BK, Zigman JM. Ghrelin as a Survival Hormone. Trends Endocrinol Metab. 2017;28(12):843-54.
 50. Fasano A. Zonulin, regulation of tight junctions, and autoimmune diseases. Ann N Y Acad Sci. 2012;1258(1):25-33.
 51. Bistola V, Lambadiari V, Dimitriadis G, Ioannidis I, Makrilakis K, Tentolouris N, et al. Possible mechanisms of direct cardiovascular impact of GLP-1 agonists and DPP4 inhibitors. Heart Failure Reviews. 2018;23(3):377-88.
 52. Korbonits M, Goldstone AP, Gueorguiev M, Grossman AB. Ghrelin—a hormone with multiple functions. Frontiers in Neuroendocrinology. 2004;25(1):27-68.
 53. Yaney G, Corkey B. Fatty acid metabolism and insulin secretion in pancreatic beta cells. Diabetologia. 2003;46(10):1297-312.
 54. Meier U, Gressner AM. Endocrine Regulation of Energy Metabolism: Review of Pathobiochemical and Clinical Chemical Aspects of Leptin, Ghrelin, Adiponectin, and Resistin. Clinical Chemistry. 2004;50(9):1511-25.
 55. Satoh N, Ogawa Y, Katsuura G, Tsuji T, Masuzaki H, Hiraoka J, et al. Pathophysiological Significance of the Obese Gene Product, Leptin, in Ventromedial Hypothalamus (VMH)-Lesioned Rats: Evidence for Loss of Its Satiety Effect in VMH-Lesioned Rats★. Endocrinology. 1997;138(3):947-54.
 56. Adamska E, Ostrowska L, Górska M, Krętowski A. The role of gastrointestinal hormones in the pathogenesis of obesity and type 2 diabetes. Prz Gastroenterol. 2014;9(2):6976.

57. Ibrahim Abdalla MM. Ghrelin - Physiological Functions and Regulation. *Eur Endocrinol.* 2015;11(2):90-5.
58. Tabák AG, Herder C, Rathmann W, Brunner EJ, Kivimäki M. Prediabetes: a high-risk state for diabetes development. *Lancet.* 2012;379(9833):2279-90.
59. Andes LJ, Cheng YJ, Rolka DB, Gregg EW, Imperatore G. Prevalence of prediabetes among adolescents and young adults in the United States, 2005-2016. *JAMA pediatrics.* 2020;174(2):e194498-e.
60. Punthakee Z, Goldenberg R, Katz P. Definition, classification and diagnosis of diabetes, prediabetes and metabolic syndrome. *Canadian journal of diabetes.* 2018;42:S10-S5.
61. Tabák AG, Herder C, Rathmann W, Brunner EJ, Kivimäki M. Prediabetes: a high-risk state for diabetes development. *The Lancet.* 2012;379(9833):2279-90.
62. Stefanaki C, Bacopoulou F, Peppas M. Prediabetes and adolescence—trends, causes, effects, and screening. *US Endocrinol.* 2016;12(2):94-8.
63. Sharma P, Panchal A, Yadav N, Narang J. Analytical techniques for the detection of glycated haemoglobin underlining the sensors. *International Journal of Biological Macromolecules.* 2020;155:685-96.
64. Mzimela NC, Ngubane PS, Khathi A. The changes in immune cell concentration during the progression of pre-diabetes to type 2 diabetes in a high-fat high-carbohydrate diet-induced pre-diabetic rat model. *Autoimmunity.* 2019;52(1):27-36.
65. Khan RMM, Chua ZJY, Tan JC, Yang Y, Liao Z, Zhao Y. From Pre-Diabetes to Diabetes: Diagnosis, Treatments and Translational Research. *Medicina (Kaunas).* 2019;55(9).
66. Marín-Peñalver JJ, Martín-Timón I, Sevillano-Collantes C, Del Cañizo-Gómez FJ. Update on the treatment of type 2 diabetes mellitus. *World J Diabetes.* 2016;7(17):354-95.
67. Khathi A, Luvuno M, Mabandla M. VOLUNTARY INGESTION OF A HIGH-FAT HIGH-CARBOHYDRATE DIET: A MODEL FOR PREDIABETES. *PONTE International Scientific Researchs Journal.* 2018;74.
68. Oguntibeju OO. Type 2 diabetes mellitus, oxidative stress and inflammation: examining the links. *Int J Physiol Pathophysiol Pharmacol.* 2019;11(3):45-63.
69. Zamora-Kapoor A, Fyfe-Johnson A, Omidpanah A, Buchwald D, Sinclair Ki. Risk factors for pre-diabetes and diabetes in adolescence and their variability by race and ethnicity. *Prev Med.* 2018;115:47-52.
70. Hostalek U. Global epidemiology of prediabetes - present and future perspectives. *Clin Diabetes Endocrinol.* 2019;5:5.

71. Joo E, Harada N, Yamane S, Fukushima T, Taura D, Iwasaki K, et al. Inhibition of Gastric Inhibitory Polypeptide Receptor Signaling in Adipose Tissue Reduces Insulin Resistance and Hepatic Steatosis in High-Fat Diet–Fed Mice. *Diabetes*. 2017;66(4):868.
72. Van Doorn C, Macht VA, Grillo CA, Reagan LP. Leptin resistance and hippocampal behavioral deficits. *Physiology & behavior*. 2017;176:207-13.
73. Nauck MA, Quast DR, Wefers J, Meier JJ. GLP-1 receptor agonists in the treatment of type 2 diabetes – state-of-the-art. *Molecular Metabolism*. 2021;46:101102.
74. Fu Y, Kaneko K, Lin H-Y, Mo Q, Xu Y, Suganami T, et al. Gut Hormone GIP Induces Inflammation and Insulin Resistance in the Hypothalamus. *Endocrinology*. 2020;161(9).
75. Shankar K, Takemi S, Gupta D, Varshney S, Mani BK, Osborne-Lawrence S, et al. Ghrelin cell-expressed insulin receptors mediate meal- and obesity-induced declines in plasma ghrelin. *JCI Insight*. 2021;6(18).
76. Omran DM, Alaraji SM, Albayati AH, Essam W. Relationship between Ghrelin and Leptin with Insulin Resistance in Obese Patients and Non-Obese Individuals. *Research Journal of Pharmacy and Technology*. 2018;11(1):281-3.
77. Papatheodorou K, Banach M, Bekiari E, Rizzo M, Edmonds M. Complications of Diabetes 2017. *Journal of Diabetes Research*. 2018;2018:3086167.
78. Abdul-Ghani MA, DeFronzo RA. Pathophysiology of prediabetes. *Current Diabetes Reports*. 2009;9(3):193-9.
79. Ahren B. DPP-4 inhibitors. *Best Practice & Research Clinical Endocrinology & Metabolism*. 2007;21(4):517-33.
80. Ha K, Joung H, Song Y. Inadequate fat or carbohydrate intake was associated with an increased incidence of type 2 diabetes mellitus in Korean adults: a 12-year community-based prospective cohort study. *Diabetes research and clinical practice*. 2019;148:254-61.
81. Gamede M, Mabuza L, Ngubane P, Khathi A. The effects of plant-derived oleanolic acid on selected parameters of glucose homeostasis in a diet-induced pre-diabetic rat model. *Molecules*. 2018;23(4):794.
82. Luvuno M, Khathi A, Mabandla MV. Diet-induced prediabetes: effects of exercise treatment on risk factors for cardiovascular complications. *Nutrition & Metabolism*. 2021;18(1):1-9.
83. Sosibo AM. Investigating changes to the insulin signalling pathway in a diet-induced pre-diabetic rat model: effects on selected markers 2019.

84. Association AD. 2. Classification and Diagnosis of Diabetes: Standards of Medical Care in Diabetes—2021. *Diabetes Care*. 2020;44(Supplement_1):S15-S33.
85. Kostov K, Blazhev A. Use of glycated hemoglobin (A1c) as a biomarker for vascular risk in type 2 diabetes: Its relationship with matrix metalloproteinases-2,-9 and the metabolism of collagen IV and elastin. *Medicina*. 2020;56(5):231.
86. Ahmed MH, Ghatge MS, Safo MK. Hemoglobin: structure, function and allostery. *Subcellular biochemistry*. 2020;94:345.
87. Sacks DB, Bebu I, Lachin JM. Refining measurement of hemoglobin A1c. *Clinical chemistry*. 2017;63(9):1433-5.
88. Wang C, Ye Y, Sun W, Yu J, Wang J, Lawrence DS, et al. Red blood cells for glucoseresponsive insulin delivery. *Advanced Materials*. 2017;29(18):1606617.
89. Sherwani SI, Khan HA, Ekhzaimy A, Masood A, Sakharkar MK. Significance of HbA1c test in diagnosis and prognosis of diabetic patients. *Biomarker insights*. 2016;11:BMI. S38440.
90. Renz PB, Cavagnolli G, Weinert LS, Silveiro SP, Camargo JL. HbA1c test as a tool in the diagnosis of gestational diabetes mellitus. *PLoS One*. 2015;10(8):e0135989.
91. El K, Campbell JE. The role of GIP in α -cells and glucagon secretion. *Peptides*. 2020;125:170213.
92. Samms RJ, Coghlan MP, Sloop KW. How may GIP enhance the therapeutic efficacy of GLP-1? *Trends in Endocrinology & Metabolism*. 2020;31(6):410-21.
93. Petersen MC, Shulman GI. Mechanisms of insulin action and insulin resistance. *Physiological reviews*. 2018;98(4):2133-223.
94. Fu Y, Kaneko K, Lin H-Y, Mo Q, Xu Y, Suganami T, et al. Gut hormone GIP induces inflammation and insulin resistance in the hypothalamus. *Endocrinology*. 2020;161(9):bqaa102.
95. Kanemaru Y, Harada N, Shimazu-Kuwahara S, Yamane S, Ikeguchi E, Murata Y, et al. Absence of GIP secretion alleviates age-related obesity and insulin resistance. *Journal of Endocrinology*. 2020;245(1):13-20.
96. D'souza AM, Neumann UH, Glavas MM, Kieffer TJ. The glucoregulatory actions of leptin. *Molecular metabolism*. 2017;6(9):1052-65.
97. Obradovic M, Sudar-Milovanovic E, Soskic S, Essack M, Arya S, Stewart AJ, et al. Leptin and Obesity: Role and Clinical Implication. *Frontiers in Endocrinology*. 2021;12.
98. Paz-Filho G, Mastronardi C, Wong ML, Licinio J. Leptin therapy, insulin sensitivity, and glucose homeostasis. *Indian J Endocrinol Metab*. 2012;16(Suppl 3):S549-55.

99. Wang G, Liu X, Christoffel KK, Zhang S, Wang B, Liu R, et al. Prediabetes is not all about obesity: association between plasma leptin and prediabetes in lean rural Chinese adults. *European Journal of Endocrinology*. 2010;163(2):243-9.
100. Hira T, Pinyo J, Hara H. What is GLP-1 really doing in obesity? *Trends in Endocrinology & Metabolism*. 2020;31(2):71-80.
101. Toi PL, Anothaisintawee T, Chaikledkaew U, Briones JR, Reutrakul S, Thakkinstian A. Preventive role of diet interventions and dietary factors in type 2 diabetes mellitus: an umbrella review. *Nutrients*. 2020;12(9):2722.
102. Cai X, Zhang Y, Li M, Wu JH, Mai L, Li J, et al. Association between prediabetes and risk of all cause mortality and cardiovascular disease: updated meta-analysis. *BMJ*. 2020;370:m2297.
103. Stentz FB, Mikhael A, Kineish O, Christman J, Sands C. High protein diet leads to prediabetes remission and positive changes in incretins and cardiovascular risk factors. *Nutrition, Metabolism and Cardiovascular Diseases*. 2021;31(4):1227-37.
104. Röhling M, Kempf K, Banzer W, Berg A, Braumann K-M, Tan S, et al. Prediabetes conversion to normoglycemia is superior adding a low-carbohydrate and energy deficit formula diet to lifestyle intervention—A 12-month subanalysis of the ACOORH trial. *Nutrients*. 2020;12(7):2022.
105. Gastaldello A, Giampieri F, De Giuseppe R, Grosso G, Baroni L, Battino M. The rise of processed meat alternatives: A narrative review of the manufacturing, composition, nutritional profile and health effects of newer sources of protein, and their place in healthier diets. *Trends in Food Science & Technology*. 2022;127:263-71.
106. Gastaldello A, Giampieri F, Quiles JL, Navarro-Hortal MD, Aparicio S, García Villena E, et al. Adherence to the Mediterranean-Style Eating Pattern and Macular Degeneration: A Systematic Review of Observational Studies. *Nutrients*. 2022;14(10):2028.
107. Nauck MA, Meier JJ. Incretin hormones: Their role in health and disease. *Diabetes, Obesity and Metabolism*. 2018;20:5-21.
108. Campbell Jonathan E, Drucker Daniel J. Pharmacology, Physiology, and Mechanisms of Incretin Hormone Action. *Cell Metabolism*. 2013;17(6):819-37.
109. Nauck MA, Meier JJ. The incretin effect in healthy individuals and those with type 2 diabetes: physiology, pathophysiology, and response to therapeutic interventions. *The Lancet Diabetes & Endocrinology*. 2016;4(6):525-36.

110. Yaribeygi H, Maleki M, Atkin SL, Jamialahmadi T, Sahebkar A. Impact of IncretinBased Therapies on Adipokines and Adiponectin. *Journal of Diabetes Research*. 2021;2021:3331865.
111. Stoica L, Gadea R, Navolan D-B, Lazar F, Duta C, Stoian D, et al. Plasma ghrelin, adiponectin and leptin levels in obese rats with type 2 diabetes mellitus after sleeve gastrectomy and gastric plication. *Experimental and Therapeutic Medicine*. 2021;21(3):1-.
112. Boer GA, Holst JJ. Incretin hormones and type 2 diabetes—mechanistic insights and therapeutic approaches. *Biology*. 2020;9(12):473.
113. Forouhi NG, Misra A, Mohan V, Taylor R, Yancy W. Dietary and nutritional approaches for prevention and management of type 2 diabetes. *Bmj*. 2018;361.
114. Amer OE, Sabico S, Alfawaz HA, Aljohani N, Hussain SD, Alnaami AM, et al. Reversal of Prediabetes in Saudi Adults: Results from an 18 Month Lifestyle Intervention. *Nutrients*. 2020;12(3):804.
115. Liu X, Wu S, Song Q, Xizhu W. Reversion From Pre-Diabetes Mellitus to Normoglycemia and Risk of Cardiovascular Disease and All-Cause Mortality in a Chinese Population: A Prospective Cohort Study. *Journal of the American Heart Association*. 2021;10.
116. Vistisen D, Kivimäki M, Perreault L, Hulman A, Witte DR, Brunner EJ, et al. Reversion from prediabetes to normoglycaemia and risk of cardiovascular disease and mortality: the Whitehall II cohort study. *Diabetologia*. 2019;62(8):1385-90.
117. Joshi S, Ostfeld RJ, McMacken M. The ketogenic diet for obesity and diabetes—enthusiasm outpaces evidence. *JAMA internal medicine*. 2019;179(9):1163-4.
118. Astrup A, Larsen TM, Harper A. Atkins and other low-carbohydrate diets: hoax or an effective tool for weight loss? *The Lancet*. 2004;364(9437):897-9.
119. Crowe T. Safety of low-carbohydrate diets. *Obesity reviews*. 2005;6(3):235-45.
120. Noakes TD. Low-carbohydrate and high-fat intake can manage obesity and associated conditions: Occasional survey. *South African Medical Journal*. 2013;103(11):826-30.
121. Seino Y, Fukushima M, Yabe D. GIP and GLP-1, the two incretin hormones: similarities and differences. *Journal of diabetes investigation*. 2010;1(1-2):8-23.
122. Ghadge AA, Khair AA. Leptin as a predictive marker for metabolic syndrome. *Cytokine*. 2019;121:154735.
123. Camilleri M. Gastrointestinal hormones and regulation of gastric emptying. *Curr Opin Endocrinol Diabetes Obes*. 2019;26(1):3-10.

124. Demir AK, Şahin Ş, Kaya SU, Bütün İ, Çıtlı R, Önder Y, et al. Prevalence of insulin resistance and identifying HOMA1-IR and HOMA2-IR indexes in the Middle Black Sea region of Turkey. *African Health Sciences*. 2020;20(1):277-86.
125. Bungau S, Behl T, Tit DM, Banica F, Bratu OG, Diaconu CC, et al. Interactions between leptin and insulin resistance in patients with prediabetes, with and without NAFLD. *Experimental and Therapeutic Medicine*. 2020;20(6):1-.
126. Mobbs CV, Mastaitis J, Yen K, Schwartz J, Mohan V, Poplawski M, et al. Lowcarbohydrate diets cause obesity, low-carbohydrate diets reverse obesity: a metabolic mechanism resolving the paradox. *Appetite*. 2007;48(2):135-8.
127. Nah E-H, Chu J, Kim S, Cho S, Kwon E. Efficacy of lifestyle interventions in the reversion to normoglycemia in Korean prediabetics: One-year results from a randomised controlled trial. *Primary Care Diabetes*. 2019;13(3):212-20.
128. Iepsen E, Lundgren J, Dirksen C, Jensen J-E, Pedersen O, Hansen T, et al. Treatment with a GLP-1 receptor agonist diminishes the decrease in free plasma leptin during maintenance of weight loss. *International Journal of Obesity*. 2015;39(5):834-41.
129. Mzimela N, Dimba NR, Sosibo AM, Khathi A. Investigating the effect of incretin peptide-levels in the development of diet-induced prediabetes. 2023.
130. Laakso M, Zilinskaite J, Hansen T, Boesgaard TW, Vanttinen M, Stancáková A, Jansson PA, Pellmé F, Holst JJ, Kuulasmaa T, Hribal ML, Sesti G, Stefan N, Fritsche A, Häring H, Pedersen O, Smith U; EUGENE2 Consortium. Insulin sensitivity, insulin release and glucagon-like peptide-1 levels in persons with impaired fasting glucose and/or impaired glucose tolerance in the EUGENE2 study. *Diabetologia*. 2008 Mar;51(3):502-11. doi: 10.1007/s00125-007-0899-2. Epub 2007 Dec 14. PMID: 18080106.
131. Vollmer K, Holst JJ, Baller B, Ellrichmann M, Nauck MA, Schmidt WE, Meier JJ. Predictors of incretin concentrations in subjects with normal, impaired, and diabetic glucose tolerance. *Diabetes*. 2008 Mar;57(3):678-87. doi: 10.2337/db07-1124. Epub 2007 Dec 5. PMID: 18057091.

APPENDICES

Appendix 1: AREC Ethics Approval Letter



25 January 2022

Dr Andile Khathi (13246)
School of Laboratory Medicine & Medical Sciences
Westville Campus

Dear Dr Kathi,

Protocol reference number: AREC/00003627/2021

Project title: Use of stored samples collected under AREC/024/018D for supervised student research.

Full Approval – Research Application

With regard to your revised application received on 18 November 2021, the Animal Research Ethics Committee has accepted the documents submitted and **FULL APPROVAL** for the protocol has been granted.

Please note: There must be adherence to national and institutional COVID-19 regulations and guidelines at all times. Researchers will be personally responsible and liable for non-adherence to national regulations. If in doubt, please contact the Research Ethics Chair and/or the University Dean of Research for advice.

Please note: Any Veterinary and Para-Veterinary procedures must be conducted by a SAVC registered VET or SAVC authorized person.

Any alteration/s to the approved research protocol, i.e Title of Project, Location of the Study, Research Approach and Methods must be reviewed and approved through the amendment/modification prior to its implementation. In case you have further queries, please quote the above reference number.

Please note: Research data should be securely stored in the discipline/department for a period of 5 years.

The ethical clearance certificate is only valid for a period of one year from the date of issue. Renewal for the study must be applied for before 24 January 2023.

Attached to the Approval letter is a template of the Progress Report that is required at the end of the study, or when applying for Renewal (whichever comes first). An Adverse Event Reporting form has also been attached in the event of any unanticipated event involving the animals' health / wellbeing.

I take this opportunity of wishing you everything of the best with your study.

Yours faithfully

Dr Sanil D Singh, BVSc, MS, PhD
Chair: Animal Research Ethics Committee
/kr

cc BRU Manager: Dr Jaca

Animal Research Ethics Committee (AREC)

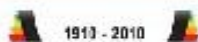
Ms Karen Reinertsen (Administrator)

Westville Campus, Govan Mbeki Building

Postal Address: Private Bag X54001, Durban 4000

Telephone: +27 (0) 31 260 8850 Facsimile: +27 (0) 31 260 4609 Email: animalethics@ukzn.ac.za

Website: <http://research.ukzn.ac.za/Research-Ethics/Animal-Ethics.aspx>



1913 - 2010

Founding Campuses: Edgewood Howard College Medical School Pietermaritzburg Westville

Appendix 2: Manuscript 1 journal guide: Canadian Journal of Physiology and Pharmacology

Prepare your manuscript.

Format and style Manuscript text must: be in English or French be double-spaced be single-column include page numbers include continuous line numbers (before acceptance only) be 8.5 x 11 inches in page size (or ISO A4)

follow this order: title page, abstract, keywords, body text (Introduction, Materials and methods, Results, Discussion), acknowledgements, references, tables, figure captions, figures, appendices

Abbreviations and acronyms

Define abbreviations and acronyms when they are first mentioned in the text.

Footnotes

In body text, try to avoid footnotes. If unavoidable, cite footnotes using superscript Arabic numbers (^{1,2,3}), in order of appearance (starting with the title page), and include the footnote at the bottom of the page on which it is cited. Do not include footnotes in the reference list.

In tables, cite footnotes using symbols (in the order ★, †, ‡, §, ||, ¶, #) or superscript lowercase italic letters (^{a,b,c}).

Mathematical expressions

Identify equations by calling out with numbers in parentheses placed flush with the left margin (for the *Canadian Journal of Physics*, place on the right).

A letter or symbol should represent only one entity and be used consistently throughout the paper.

Each variable (including those representing vectors, matrices, and tensors) must be clearly identified and defined in the text.

Supply complex equations in an editable format by using LaTeX or a math editor (MathType).

Supply simple, inline equations in Word, without using MathType. Insert symbols from Word's

“Symbol” palette, using “normal text” or “Symbol” fonts only. Insert symbols using MathType ONLY if they cannot be found in the “Symbol” palette under one of those two fonts.

Reporting guidelines

Study reporting guidelines can help authors report their work transparently and accurately. We encourage their use. Up-to-date guidelines can be found at the [EQUATOR Network](#), where authors can [search](#) or [consult the GoodReports wizard](#) to identify which guideline(s) to use. A completed copy of the guideline checklist may be submitted with the manuscript as a Supplementary file.

Spelling

Spelling should follow that of *Webster's Third New International Dictionary* or the *Oxford English Dictionary*. Authors are responsible for consistency in spelling.

Statistical analyses

The assumptions and (or) the model underlying any statistical analysis should be clearly stated. Do not use symbols such as ★ and ★★ to denote levels of significance unless accompanied by actual *p* values.

Units of measure

Use SI units of measure ([Système international d'unités](#)). If non-SI units are used, at first mention, supply the equivalent in SI units in parentheses.

Parts of the manuscript

Title page

Title

Should be accurate, informative, and brief. Include keywords in the title to optimize search engine discovery.

Author list

List all author names on the title page: check author order, spelling, capitalization, initials, and hyphens.

Format names as: first name (or initial) middle name (or initial) last name (surname/family name).

List affiliation(s) for each author that include: institution (department and university or organization), city, state or province, country. An author's affiliations should reflect where the research was conducted. If an author changes institution, the new affiliation can be listed in a footnote.

Do not include academic degrees and professional titles.

Authorship criteria: We subscribe to the [ICMJE definition of authorship](#) in most cases. Any person listed as an author must meet each of these authorship criteria, and anyone who meets these authorship criteria must be listed as an author. Contributors who do not meet authorship criteria should be listed in the Acknowledgements section. Exceptions to our definition of authorship may exist for communityengaged research projects; please see our [Publishing Policy](#) for authorship and attribution options.

Corresponding author

Clearly identify the corresponding author and their email address on the title page. The corresponding author is designated to receive post-publication queries from readers.

Note that authors may supply their ORCID iD on submission; these identifiers are not required on the title page itself.

Abstract

Provide the abstract in the manuscript text file on page 2, after the title page. Authors must supply their abstract in English and/or French. (Note that CSP plans to accommodate author-provided abstract translations in other languages in the future.)

The abstract should be a single paragraph that summarizes the article. It helps readers decide whether to keep reading. Briefly describe the study rationale, objectives, methods, findings, and impact. Use keywords (and their plain-language synonyms) in the abstract to optimize search engine discovery. Do not include headings, reference citations, tables, figures, or acronyms.

Keywords

List keywords after the abstract. Good keywords are common to your field and accurately describe your topic. Use keywords in the title, abstract, and manuscript text to optimize search engine discovery. (Consider what words you would enter in a search box to find your work online.)

Graphical abstract

(Optional) Authors are encouraged to submit an illustration, diagram, equation, or other informative visual that explains the central message of the article and entices readers. The maximum allowable size is 40 mm (150 pixels) high by 85 mm (320 pixels) wide. Graphical abstracts appear online only.

Plain language summaries

(Optional) At acceptance, authors are encouraged to submit a plain language summary of their article to increase the reach of their research. For guidelines and submission instructions, see [Writing a Plain](#)

Main body of the article

Introduction

In 1–3 paragraphs, explain the study rationale and objective(s).

What is the problem, and why is it important?

What is known on the topic? Establish context: provide background, briefly review the key literature, and mention existing gaps or controversies in the field.

What is the overall aim of the study? State your research question, hypotheses, and predictions.

Materials and methods

Describe what you did (and how you did it) clearly and comprehensively enough for the study to be replicated.

Include as appropriate the study design; primary and secondary outcome measures; computational, experimental, and statistical methods; materials; locations.

Avoid long descriptions of known procedures; provide relevant references instead.

Specify materials used (e.g., laboratory or field equipment, chemicals, biologic materials) and their sources (provide company name, city, and country in parentheses).

Ethics

approval

Information about Ethics Approvals should be reported in the Materials and methods section (usually the first paragraph).

For studies involving human participants: name the institutional ethics review committee that approved the study; and confirm that participants gave informed consent before participating in the study.

For studies involving animals:

name the animal care review committee that approved the study;

name the animal care guidelines that were followed (must be in line with the [Canadian Council on Animal Care guidelines](#) or the [Guide for the Care and Use of Laboratory Animals](#));

confirm that research involving endangered species was conducted in accordance with all applicable laws; and if permits were required for fieldwork, supply the research permit and (or) license numbers.

For studies involving collection of specimens:

confirm that research involving collection of specimens was conducted in accordance with all applicable laws, guidelines, regulations; and if permits were required for fieldwork, supply the research permit and (or) license numbers.

Results

Report results that are directly relevant to your research question. Raw data and other observations may be submitted with the manuscript as Supplementary files or uploaded to a recognized data repository.

Use subheadings, tables, and figures to organize and communicate your findings.

Begin sections and (or) sentences with high-level observations, followed by statistical data.

State the statistical tests used (if applicable), and when reporting numbers:

define the values provided (e.g., mean and standard deviation or standard error, median, and interquartile range), and include the absolute value of N when describing frequencies (i.e., percentages, proportions, ratios).

Discussion or Conclusion

Compare your findings with previously published work; include points of agreement and difference. Describe the limitations and main contributions of your work. Propose avenues of future study. Speculation should be clearly identified and based on observations related to the manuscript.

Author statements

Acknowledgements

(Optional) The names and contributions of institutions and people who contributed to the work but do not meet authorship criteria may be listed in the Acknowledgements section. This section should be written in the third person. Authors are responsible for ensuring that people named in the Acknowledgements agree to be named.

Competing interests statement

Authors are responsible for disclosing all financial and non-financial relationships that might bias or be seen to bias their work. Supply a statement of competing interests during submission and on the article title page. Authors who are unsure what to list may wish to consult the [ICMJE form for disclosure of competing interests](#).

If there is nothing to declare, the statement should read: “Competing interests: The authors declare there are no competing interests.”

If there are competing interests to declare, specify authors by full name. Statements should take the form of “Competing interests: AUTHOR is an employee and shareholder of COMPANY. AUTHOR has received speaker fees and travel honoraria from COMPANY.”

Author contribution statement

Supply an author contribution statement, identifying authors by their initials and specifying contributions using the [Contributor Roles Taxonomy \(CRediT\)](#) roles as selected for each author in the peer-review system during submission.

Community involvement statement

(Optional) For studies involving Indigenous communities or community-engaged research, authors may supply a community involvement statement that describes how the community was involved throughout the research process and how the study benefits the community. For guidelines on what to include in the statement and how to transparently report other features of the project in the article, refer to our [community-engaged research page](#).

Funding statement

Supply a funding statement that lists what support the authors received to carry out the research.

If the study was unfunded, state “Funding: The authors declare no specific funding for this work.”

If the study received funding, include each funding agency name written out in full, followed by its grant or award number in parentheses. E.g., “Funding: This research was supported by FUNDING AGENCY NAME (grant No. ###).”

Data availability statement

Supply a data availability statement that says whether any, all, or portions of the data underpinning the work are available to others.

If data are available, specify how data can be accessed and under what conditions data can be reused. Supply repository name, persistent unique identifier (PID: DOI/compact identifier/accession number), and web link.

If data are not available, explain why (e.g., describe the ethical, legal, or commercial restrictions).

For complete instructions, see [How to write a data availability statement](#).

References

See journal-specific guidelines for information on preferred citation style. Note the following:

Unpublished reports, private communications, and submitted manuscripts do not belong in the reference list. Include them as footnotes or parenthetically in the body text, e.g., (J. Jones, personal communication, 2010).

Books and articles that are in press (accepted but not yet published) should be cited in text and included in the reference list.

Publicly available datasets should be cited in text and included in the reference list (see [How to cite datasets](#)).

Tables and Figures

In general:

Cite each figure and table, and name them in order of first appearance in the text (Figure 1, Figure 2, Table 1, Figure 3, Table 2, etc.).

Figures and tables should add information to the article, not duplicate results that are (or could be) explained briefly in the text.

Figures and tables should be understandable without having to read the article. Any abbreviations or symbols used should be defined in the caption (figures) or in the Note (tables).

Table specifications:

Supply tables in an editable format, i.e., Word (table function in DOC or DOCX) or Excel (spreadsheet in XLS or XLSX). Each data point should be in a separate cell.

Provide a table caption (e.g., Table 1. Xxx).

Footnotes should be designated by symbols (order: ★, †, ‡, §, ||, ¶, #) or superscript lowercase italic letters (^{a,b,c}).

Use a Note under the table for additional information.

Figure specifications:

See [How to prepare figures](#) for figure specifications and guidance.

A caption explains how to read a figure, independent of the article text. Follow the guidelines below:

Terms, abbreviations, and symbols used must be defined and correspond with those in the text.

List tools and settings used for image capture.

Maps: captions must credit all sources used to produce the map: base map if used (e.g., [Google Map attribution guidelines](#)), and data plotted on the map (e.g., geological units and contacts, shape file data, boundary file data, data from other studies). List map projection (e.g., NAD83) and coordinate system (e.g., UTM) used, if applicable.

Include a permission statement for figures adapted, modified, or republished from elsewhere. Upload a copy of your permission from the original copyright holder with your files or send by email to the journal editorial office.

Appendices

An appendix should be able to stand alone, as a separate, self-contained document. Figures, equations, and tables used in an appendix should be numbered sequentially but separately from those used in the main body of the paper, e.g., Fig. A1, eq. (A1), Table A1.

Supplementary files

Supplementary files consist of extra tables, figures (maps), detailed calculations, and data sets produced by the authors as part of their research, but not essential for understanding or evaluating the manuscript.

Number figures, equations, and tables in a supplementary file sequentially but separately from the numbering used in the manuscript (e.g., Fig. S1, eq. S1, Table S1).

Cite supplementary files in the manuscript in a footnote.

During submission, use the “Supplementary Material” file designation when uploading supplementary files.

Supplementary files are not edited or converted and, therefore, will appear exactly as submitted.

Supplementary files should be named by manuscript number followed by “suppla”, “supplb” etc. e.g., APNM-2019-0401suppla.

Multimedia files

Authors may incorporate audio and video clips into their paper; these are published online, adding dimension to the paper. For submission guidelines and accepted formats, see technical specifications under [How to prepare a video abstract](#).

[SUBMIT ARTICLE](#)

Style guide

For general matters of style, this journal uses for biological terms, [Scientific Style and Format: The CSE Manual for Authors, Editors, and Publishers](#) (8th ed., 2014)), published by the Council of Science Editors.

Nomenclature

Nomenclature should follow the rules established by international authoritative bodies.

Abstract

Abstracts should not exceed 200 words for all article types.

Length per manuscript type

Article type	Description	Specifications (word, table/fig, reference limits)
A maximum of 10,000 words including		
Article that reports new and original article type. Editors or reviewers may request research. changes to the length and/or structure of the article at their discretion.	A completed, definitive study	references, tables and figures is suggested for this
Brief report	Reports a completed project of a smaller scope, such as a short case study or pilot research that is novel and can direct follow-up.	6000 words including references, tables and figures.
Rapid communication	Reports time-sensitive research. On submission, explain why the research figures. merits rapid publication.	3000 words including

Review	By invitation only: a review of a topic of current interest.	10 000 words including references, tables and figures.
	Prospective authors can contact the Editor to establish the suitability and timeliness of the topic for publication.	

Data registration

Nucleic acid sequences

Authors must submit the relevant data to an appropriate database. The accession number assigned to the deposited sequence must be provided in the manuscript. Learn more about submitting to these databases:

[GenBank](#)

[ENA \(European Nucleotide Archive, EMBL-EBI\)](#)

[DDBJ](#) (DNA Data Bank of Japan)

[RDP \(Ribosomal Database Project\)](#)

National Center for Biotechnology Information: <https://www.ncbi.nlm.nih.gov/>

Clinical trials

Clinical trials must have been registered in a publicly accessible clinical trial registry before patient enrolment. Supply the name of the clinical trial registry and the study's registration number online at submission, and list them at the end of the abstract in the manuscript text file. A data sharing statement is also required. Based on [ICMJE guidelines on trial registration](#), the statement should indicate:

whether individual deidentified participant data (including data dictionaries) will be shared

("undecided" is not an acceptable answer); what data will be shared; what additional, related documents will be available (e.g., study protocol, statistical analysis plan); when the data will become available and for how long; and

by what access criteria data will be shared (including with whom, for what types of analyses, and by what mechanism).

References

This journal uses the Harvard citation style. In-text citations take the form (Author Year). References must be listed in alphabetical order according to the name of the first author and not numbered.

References with the same first author are listed in the following order.

Papers with one author only are listed first in chronological order, beginning with the earliest paper.

Papers with dual authorship follow and are listed in alphabetical order by the last name of the second author.

Papers with three or more authors appear after the dual-authored papers and are arranged chronologically.

Include DOIs and hyperlinks whenever possible, and do not number the references. If your article is accepted for publication, we will format the list for you.

Appendix 3: Manuscript 2 Journal guide: Endocrinología, Diabetes y Nutrición

Guide for Authors

Types of articles

The Journal contains the following sections:

Original articles

Original clinical, translational, or experimental research papers covering any aspect related to endocrinology, nutrition, diabetes, or metabolism. The recommended designs are analytical in the form of observational studies (cross-sectional, case-control studies, cohort studies) following the [STROBE](#) methodology and randomized controlled trials following the [CONSORT](#) methodology and registered in a public database. The article should have a maximum length of 4000 words, not including references, abstract, and figure legends. It will be structured in the following sections: Introduction, Material and methods, Results, and Discussion. Subsections are only allowed in the Material and Methods and Results sections. The abstract (in Spanish and English) can have a maximum of 250 words and must be structured. The bibliography must be limited to a maximum of 30 references. A maximum of 6 tables and/or figures in total will be accepted. It will be included the main findings section, together with the abstract, no more than 80 words, divided into 4 or 5 sentences. Authors are also encouraged to provide a graphical abstract of the article.

Systematic reviews and meta-analyses

Systematic reviews using [PRISMA](#) or [MOOSE](#) criteria are recommended. It is recommended that the review protocol is registered in a public database and that the reference number is included in the publication. The article should have a maximum length of 5000 words, excluding literature references, abstract and figure legends. It should be structured in the following sections: Introduction, Material and Methods, Results and Discussion. There is no limit to the number of literature references (provided they are justified). A maximum of 6 tables and/or figures in total will be accepted. If necessary, they can be complemented with supplementary tables/figures.

Case reports

A case report is another type of review. It is an update on a topic related to a clinical case. This type of manuscript can be commissioned or submitted on the authors' own initiative. They will be written in English, will have a maximum length of 2500 words, and an unstructured abstract of 150 words, the corresponding keywords, and up to 50 literature references. Up to 3 figures and/or tables will be accepted. It will be structured as follows: description of the case, management (and supporting evidence), areas of uncertainty, guidelines and conclusion and recommendations.

Scientific letters

Descriptions may be included of one or more clinical cases of exceptional observation that represent a significant contribution to understanding the pathophysiology or other aspects of endocrine diseases. They may also report research experiences or results which, because of their characteristics, are not sufficiently important to be published as an original article. Maximum length of the text will be 1000 words, and it should not be structured in sections. It should not be accompanied by an abstract either. One figure or one table will be accepted. No more than 10 literature references should be included. There should be no more than five authors.

Letters to the Editor

Letters to the Editor may refer to studies published in the Journal or express opinions on a current issue. They may not exceed 1000 words and 1 Figure or 1 Table and a maximum of 10 literature references will be accepted. There should be no more than 4 authors.

Other sections

The journal includes Editorials commissioned by the Editorial Board. Authors who wish to contribute to any of these sections should contact the journal's Editors beforehand. *Endocrinología, Diabetes y Nutrición* will also publish clinical guidelines and consensus documents on the diagnosis or treatment of diseases, provided they are promoted directly by the SED and SEEN or by working groups. These documents must be made according to the indications recommended by the SED and SEEN (www.seen.es) and follow the AGREE methodology for clinical guidelines. The guidelines prepared jointly with other scientific societies may be published in Endocrinology, Diabetes and Nutrition, and in parallel in the publications of the other scientific societies, provided that the regulations established by the SED-SEEN and the AGREE methodology have been followed. In this case, the correspondence author will attach a cover letter stating the Societies that endorse the Document (attaching the endorsement of each Society), as well as the journals to which the consensus will be sent. The journal will publish the guideline or the consensus document accepted by the Editorial Committee as an Executive Summary, whose maximum length will be 3000 words, and an unstructured abstract that will have a maximum of 150 words. It will be accompanied by tables and figures that are considered convenient to help interpret the information in a clear and concise manner and it will be sent in English. The title will begin with the following sentence: "Executive summary of the guideline/consensus document...". The full consensus document will be published online as additional material to the Executive Summary and/or as a supplement to the journal. The objective of the publication of the Executive Summary is to offer readers a synthesis of the consensus document, but with enough detail and clarity to understand the full document's scope and most relevant points. A maximum of 6 executive summaries per year will be published in the first issue of each year, with priority being given to those produced jointly by the two societies. Papers must be submitted to the board(s) of the corresponding

society(ies) by 30 June for evaluation and selection. The definition of authorship will be guided by the [Recommendations of the ICMJE](#) (International Committee of Medical Journal Editors).

Contact details for submission

You can submit your manuscript at <https://www.editorialmanager.com/endinu>. The Guide for Authors in Spanish can be found at <https://www.elsevier.es/es-revista-endocrinologia-diabetes-nutricion-13normas-publicacion>.

Page charges

This journal has no page charges.

Language

This journal is published in Spanish and in English language.

Submission checklist

You can use this list to carry out a final check of your submission before you send it to the journal for review. Please check the relevant section in this Guide for Authors for more details.

Ensure that the following items are present:

One author has been designated as the corresponding author with contact details:

- E-mail address
- Full postal address

All necessary files have been uploaded:

Manuscript:

- Include keywords
- All figures (include relevant captions)
- All tables (including titles, description, footnotes)
- Ensure all figure and table citations in the text match the files provided
- Indicate clearly if color should be used for any figures in print

Graphical Abstracts / Highlights files (where applicable)

Supplemental files (where applicable)

Further considerations

- Manuscript has been 'spell checked' and 'grammar checked'
- All references mentioned in the Reference List are cited in the text, and vice versa
- Permission has been obtained for use of copyrighted material from other sources (including the Internet)
- A competing interests statement is provided, even if the authors have no competing interests to declare
- Journal policies detailed in this guide have been reviewed
- Referee suggestions and contact details provided, based on journal requirements

For further information, visit our [Support Center](#).

Ethics in publishing

Please see our information on [Ethics in publishing](#).

Studies in humans and animals

If the work involves the use of human subjects, the author should ensure that the work described has been carried out in accordance with [The Code of Ethics of the World Medical Association](#) (Declaration of Helsinki) for experiments involving humans. The manuscript should be in line with the [Recommendations for the Conduct, Reporting, Editing and Publication of Scholarly Work in Medical Journals](#) and aim for the inclusion of representative human populations (sex, age and ethnicity) as per those recommendations. The terms [sex and gender](#) should be used correctly.

Authors should include a statement in the manuscript that informed consent was obtained for experimentation with human subjects. The privacy rights of human subjects must always be observed.

All animal experiments should comply with the [ARRIVE guidelines](#) and should be carried out in accordance with the U.K. Animals (Scientific Procedures) Act, 1986 and associated guidelines, [EU Directive 2010/63/EU for animal experiments](#), or the National Research Council's [Guide for the Care and Use of Laboratory Animals](#) and the authors should clearly indicate in the manuscript that such guidelines have been followed. The sex of animals must be indicated, and where appropriate, the influence (or association) of sex on the results of the study.

Informed consent and patient details

Studies on patients or volunteers require ethics committee approval and informed consent, which should be documented in the paper. Appropriate consents, permissions and releases must be obtained where an author wishes to include case details or other personal information or images of patients and any other individuals in an Elsevier publication. Written consents must be retained by the author but copies should not be provided to the journal. Only if specifically requested by the journal in exceptional circumstances (for example if a legal issue arises) the author must provide copies of the consents or evidence that such consents have been obtained. For more information, please review the [Elsevier Policy on the Use of Images or Personal Information of Patients or other Individuals](#). Unless you have written permission from the patient (or, where applicable, the next of kin), the personal details of any patient included in any part of the article and in any supplementary materials (including all illustrations and videos) must be removed before submission.

Declaration of interest

All authors must disclose any financial and personal relationships with other people or organizations that could inappropriately influence (bias) their work. Examples of potential competing interests include employment, consultancies, stock ownership, honoraria, paid expert testimony, patent applications/registrations, and grants or other funding. Authors must disclose any interests in two places: 1. A summary declaration of interest statement in the title page file (if double anonymized) or the manuscript file (if single anonymized). If there are no interests to declare then please state this: 'Declarations of interest: none'. 2. Detailed disclosures as part of a separate Declaration of Interest form, which forms part of the journal's official records. It is important for potential interests to be declared in both places and that the information matches. [More information](#).

Submission declaration and verification

Submission of an article implies that the work described has not been published previously (except in the form of an abstract, a published lecture or academic thesis, see '[Multiple, redundant or concurrent publication](#)' for more information), that it is not under consideration for publication elsewhere, that its publication is approved by all authors and tacitly or explicitly by the responsible authorities where the work was carried out, and that, if accepted, it will not be published elsewhere in the same form, in English or in any other language, including electronically without the written consent of the copyright-holder. To verify compliance, your article may be checked by [Crossref Similarity Check](#) and other originality or duplicate checking software.

Preprints

Please note that [preprints](#) can be shared anywhere at any time, in line with Elsevier's [sharing policy](#). Sharing your preprints e.g. on a preprint server will not count as prior publication (see '[Multiple, redundant or concurrent publication](#)' for more information).

Use of inclusive language

Inclusive language acknowledges diversity, conveys respect to all people, is sensitive to differences, and promotes equal opportunities. Content should make no assumptions about the beliefs or commitments of any reader; contain nothing which might imply that one individual is superior to another on the grounds of age, gender, race, ethnicity, culture, sexual orientation, disability or health condition; and use inclusive language throughout. Authors should ensure that writing is free from bias, stereotypes, slang, reference to dominant culture and/or cultural assumptions. We advise to seek gender neutrality by using plural nouns ("clinicians, patients/clients") as default/wherever possible to avoid using "he, she," or "he/she." We recommend avoiding the use of descriptors that refer to personal attributes such as age, gender, race, ethnicity, culture, sexual orientation, disability or health condition unless they are relevant and valid. When coding terminology is used, we recommend to avoid offensive or exclusionary terms such as "master", "slave", "blacklist" and "whitelist". We suggest using alternatives that are more appropriate and (self-) explanatory such as "primary", "secondary", "blocklist" and "allowlist". These guidelines are meant as a point of reference to help identify appropriate language but are by no means exhaustive or definitive.

Authorship

All authors should have made substantial contributions to all of the following: (1) the conception and design of the study, or acquisition of data, or analysis and interpretation of data, (2) drafting the article or revising it critically for important intellectual content, (3) final approval of the version to be submitted.

Changes to authorship

Authors are expected to consider carefully the list and order of authors before submitting their manuscript and provide the definitive list of authors at the time of the original submission. Any addition, deletion or rearrangement of author names in the authorship list should be made only before the manuscript has been accepted and only if approved by the journal Editor. To request such a change, the Editor must receive the following from the corresponding author: (a) the reason for the change in author list and (b) written confirmation (e-mail, letter) from all authors that they agree with the addition, removal or rearrangement. In the case of addition or removal of authors, this includes confirmation from the author being added or removed.

Only in exceptional circumstances will the Editor consider the addition, deletion or rearrangement of authors after the manuscript has been accepted. While the Editor considers the request, publication of the manuscript will be suspended. If the manuscript has already been published in an online issue, any requests approved by the Editor will result in a corrigendum.

Clinical trial results

In line with the position of the International Committee of Medical Journal Editors, the journal will not consider results posted in the same clinical trials registry in which primary registration resides to be prior publication if the results posted are presented in the form of a brief structured (less than 500 words) abstract or table. However, divulging results in other circumstances (e.g., investors' meetings) is discouraged and may jeopardise consideration of the manuscript. Authors should fully disclose all posting in registries of results of the same or closely related work.

Reporting clinical trials

Randomized controlled trials should be presented according to the CONSORT guidelines. At manuscript submission, authors must provide the CONSORT checklist accompanied by a flow diagram that illustrates the progress of patients through the trial, including recruitment, enrollment, randomization, withdrawal and completion, and a detailed description of the randomization procedure. The [CONSORT checklist and template flow diagram](#) are available online.

Registration of clinical trials

Registration in a public trials registry is a condition for publication of clinical trials in this journal in accordance with [International Committee of Medical Journal Editors](#) recommendations. Trials must register at or before the onset of patient enrolment. The clinical trial registration number should be included at the end of the abstract of the article. A clinical trial is defined as any research study that prospectively assigns human participants or groups of humans to one or more health-related interventions to evaluate the effects of health outcomes. Health-related interventions include any intervention used to modify a biomedical or health-related outcome (for example drugs, surgical procedures, devices, behavioural treatments, dietary interventions, and process-of-care changes). Health outcomes include any biomedical or health-related measures obtained in patients or participants, including pharmacokinetic measures and adverse events. Purely observational studies (those in which the assignment of the medical intervention is not at the discretion of the investigator) will not require registration.

Elsevier supports responsible sharing

Find out how you can [share your research](#) published in Elsevier journals.

Role of the funding source

You are requested to identify who provided financial support for the conduct of the research and/or preparation of the article and to briefly describe the role of the sponsor(s), if any, in study design; in the collection, analysis and interpretation of data; in the writing of the report; and in the decision to submit the article for publication. If the funding source(s) had no such involvement, it is recommended to state this.

Open access

Please visit our [Open Access page](#) for more information.

Elsevier Researcher Academy

[Researcher Academy](#) is a free e-learning platform designed to support early and mid-career researchers throughout their research journey. The "Learn" environment at Researcher Academy offers several interactive modules, webinars, downloadable guides and resources to guide you through the process of writing for research and going through peer review. Feel free to use these free resources to improve your submission and navigate the publication process with ease.

Language (usage and editing services)

Please write your text in good English (American or British usage is accepted, but not a mixture of these). Authors who feel their English language manuscript may require editing to eliminate possible grammatical or spelling errors and to conform to correct scientific English may wish to use the [English Language Editing service](#) available from Elsevier's Author Services.

Submission

Our online submission system guides you stepwise through the process of entering your article details and uploading your files. The system converts your article files to a single PDF file used in the peerreview process. Editable files (e.g., Word, LaTeX) are required to typeset your article for final publication. All correspondence, including notification of the Editor's decision and requests for revision, is sent by e-mail.

Submit your article

Please submit your article via <https://www.editorialmanager.com/ending>

Suggesting reviewers

Please submit the names and institutional e-mail addresses of several potential reviewers.

You should not suggest reviewers who are colleagues, or who have co-authored or collaborated with you during the last three years. Editors do not invite reviewers who have potential competing interests

with the authors. Further, in order to provide a broad and balanced assessment of the work, and ensure scientific rigor, please suggest diverse candidate reviewers who are located in different countries/regions from the author group. Also consider other diversity attributes e.g. gender, race and ethnicity, career stage, etc. Finally, you should not include existing members of the journal's editorial team, of whom the journal are already aware.

Note: the editor decides whether or not to invite your suggested reviewers.

Peer review

This journal operates a double anonymized review process. All contributions will be initially assessed by the editor for suitability for the journal. Papers deemed suitable are then typically sent to a minimum of two independent expert reviewers to assess the scientific quality of the paper. The Editor is responsible for the final decision regarding acceptance or rejection of articles. The Editor's decision is final. Editors are not involved in decisions about papers which they have written themselves or have been written by family members or colleagues or which relate to products or services in which the editor has an interest. Any such submission is subject to all of the journal's usual procedures, with peer review handled independently of the relevant editor and their research groups. [More information on types of peer review](#).

Double anonymized review

This journal uses double anonymized review, which means the identities of the authors are concealed from the reviewers, and vice versa. [More information](#) is available on our website. To facilitate this, please include the following separately:

Title page (with author details): This should include the title, authors' names, affiliations, acknowledgements and any Declaration of Interest statement, and a complete address for the corresponding author including an e-mail address.

Anonymized manuscript (no author details): The main body of the paper (including the references, figures, tables and any acknowledgements) should not include any identifying information, such as the authors' names or affiliations.

Use of word processing software

It is important that the file be saved in the native format of the word processor used. The text should be in single-column format. Keep the layout of the text as simple as possible. Most formatting codes will be removed and replaced on processing the article. In particular, do not use the word processor's options to justify text or to hyphenate words. However, do use bold face, italics, subscripts, superscripts etc. When preparing tables, if you are using a table grid, use only one grid for each individual table and not a grid for each row. If no grid is used, use tabs, not spaces, to align columns. The electronic text should

be prepared in a way very similar to that of conventional manuscripts (see also the [Guide to Publishing with Elsevier](#)). Note that source files of figures, tables and text graphics will be required whether or not you embed your figures in the text. See also the section on Electronic artwork.

To avoid unnecessary errors you are strongly advised to use the 'spell-check' and 'grammar-check' functions of your word processor.

Article structure

Subdivision - unnumbered sections

Divide your article into clearly defined sections. Each subsection is given a brief heading (Introduction, Material and methods, Results and Discussion). Each heading should appear on its own separate line. Subsections should be used as much as possible when cross-referencing text: refer to the subsection by heading as opposed to simply 'the text'.

Introduction

State the objectives of the work and provide an adequate background, avoiding a detailed literature survey or a summary of the results.

Material and methods

Provide sufficient details to allow the work to be reproduced by an independent researcher. Methods that are already published should be summarized, and indicated by a reference. If quoting directly from a previously published method, use quotation marks and also cite the source. Any modifications to existing methods should also be described.

Results

Results should be clear and concise.

Discussion

This should explore the significance of the results of the work, not repeat them. A combined Results and Discussion section is often appropriate. Avoid extensive citations and discussion of published literature.

Conclusions

The main conclusions of the study may be presented in a short Conclusions section, which may stand alone or form a subsection of a Discussion or Results and Discussion section.

Appendices

If there is more than one appendix, they should be identified as A, B, etc. Formulae and equations in appendices should be given separate numbering: Eq. (A.1), Eq. (A.2), etc.; in a subsequent appendix, Eq. (B.1) and so on. Similarly for tables and figures: Table A.1; Fig. A.1, etc.

- *Title*. Concise and informative. Titles are often used in information-retrieval systems. Avoid abbreviations and formulae where possible.
- *Author names and affiliations*. Please clearly indicate the given name(s) and family name(s) of each author and check that all names are accurately spelled. You can add your name between parentheses in your own script behind the English transliteration. Present the authors' affiliation addresses (where the actual work was done) below the names. Indicate all affiliations with a lower-case superscript letter immediately after the author's name and in front of the appropriate address. Provide the full postal address of each affiliation, including the country name and, if available, the e-mail address of each author.
- *Corresponding author*. Clearly indicate who will handle correspondence at all stages of refereeing and publication, also post-publication. This responsibility includes answering any future queries about Methodology and Materials. Ensure that the e-mail address is given and that contact details are kept up to date by the corresponding author.
- *Present/permanent address*. If an author has moved since the work described in the article was done, or was visiting at the time, a 'Present address' (or 'Permanent address') may be indicated as a footnote to that author's name. The address at which the author actually did the work must be retained as the main, affiliation address. Superscript Arabic numerals are used for such footnotes.

Highlights

Highlights are optional yet highly encouraged for this journal, as they increase the discoverability of your article via search engines. They consist of a short collection of bullet points that capture the novel results of your research as well as new methods that were used during the study (if any). Please have a look at the examples here: [example Highlights](#).

Highlights should be submitted in a separate editable file in the online submission system. Please use 'Highlights' in the file name and include 3 to 5 bullet points (maximum 85 characters, including spaces, per bullet point).

A structured abstract, by means of appropriate headings, should provide the context or background for the research and should state its purpose, basic procedures (selection of study subjects or laboratory animals, observational and analytical methods), main findings (giving specific effect sizes and their statistical significance, if possible), and principal conclusions. It should emphasize new and important aspects of the study or observations.

The headings will consist of: "Introduction", "Materials and Methods", "Results" y "Conclusions".

Graphical abstract

Although a graphical abstract is optional, its use is encouraged as it draws more attention to the online article. The graphical abstract should summarize the contents of the article in a concise, pictorial form designed to capture the attention of a wide readership. Graphical abstracts should be submitted as a separate file in the online submission system. Image size: Please provide an image with a minimum of 531 × 1328 pixels (h × w) or proportionally more. The image should be readable at a size of 5 × 13 cm using a regular screen resolution of 96 dpi. Preferred file types: TIFF, EPS, PDF or MS Office files. You can view [Example Graphical Abstracts](#) on our information site.

Keywords

Immediately after the abstract, provide a maximum of 6 keywords, using British spelling and avoiding general and plural terms and multiple concepts (avoid, for example, 'and', 'of'). Be sparing with abbreviations: only abbreviations firmly established in the field may be eligible. These keywords will be used for indexing purposes.

Abbreviations

Define abbreviations that are not standard in this field in a footnote to be placed on the first page of the article. Such abbreviations that are unavoidable in the abstract must be defined at their first mention there, as well as in the footnote. Ensure consistency of abbreviations throughout the article.

Acknowledgements

Collate acknowledgements in a separate section at the end of the article before the references and do not, therefore, include them on the title page, as a footnote to the title or otherwise. List here those individuals who provided help during the research (e.g., providing language help, writing assistance or proof reading the article, etc.).

Formatting of funding sources

List funding sources in this standard way to facilitate compliance to funder's requirements:

Funding: This work was supported by the National Institutes of Health [grant numbers xxxx, yyyy]; the Bill & Melinda Gates Foundation, Seattle, WA [grant number zzzz]; and the United States Institutes of Peace [grant number aaaa].

It is not necessary to include detailed descriptions on the program or type of grants and awards. When funding is from a block grant or other resources available to a university, college, or other research institution, submit the name of the institute or organization that provided the funding.

If no funding has been provided for the research, it is recommended to include the following sentence:

This research did not receive any specific grant from funding agencies in the public, commercial, or not-for-profit sectors.

Units

Follow internationally accepted rules and conventions: use the international system of units (SI). If other units are mentioned, please give their equivalent in SI.

Artwork

Image manipulation

Whilst it is accepted that authors sometimes need to manipulate images for clarity, manipulation for purposes of deception or fraud will be seen as scientific ethical abuse and will be dealt with accordingly. For graphical images, this journal is applying the following policy: no specific feature within an image may be enhanced, obscured, moved, removed, or introduced. Adjustments of brightness, contrast, or color balance are acceptable if and as long as they do not obscure or eliminate any information present in the original. Nonlinear adjustments (e.g. changes to gamma settings) must be disclosed in the figure legend.

Electronic artwork

General points

- Make sure you use uniform lettering and sizing of your original artwork.
- Embed the used fonts if the application provides that option.
- Aim to use the following fonts in your illustrations: Arial, Courier, Times New Roman, Symbol, or use fonts that look similar.
- Number the illustrations according to their sequence in the text.
- Use a logical naming convention for your artwork files.
- Provide captions to illustrations separately.
- Size the illustrations close to the desired dimensions of the published version.
- Submit each illustration as a separate file.
- Ensure that color images are accessible to all, including those with impaired color vision.

A detailed [guide on electronic artwork](#) is available.

You are urged to visit this site; some excerpts from the detailed information are given here.

Formats

If your electronic artwork is created in a Microsoft Office application (Word, PowerPoint, Excel) then please supply 'as is' in the native document format.

Regardless of the application used other than Microsoft Office, when your electronic artwork is finalized, please 'Save as' or convert the images to one of the following formats (note the resolution requirements for line drawings, halftones, and line/halftone combinations given below):

EPS (or PDF): Vector drawings, embed all used fonts.

TIFF (or JPEG): Color or grayscale photographs (halftones), keep to a minimum of 300 dpi. TIFF

(or JPEG): Bitmapped (pure black & white pixels) line drawings, keep to a minimum of 1000 dpi.

TIFF (or JPEG): Combinations bitmapped line/half-tone (color or grayscale), keep to a minimum of 500 dpi.

Please do not:

- Supply files that are optimized for screen use (e.g., GIF, BMP, PICT, WPG); these typically have a low number of pixels and limited set of colors;
- Supply files that are too low in resolution;
- Submit graphics that are disproportionately large for the content.

Color artwork

Please make sure that artwork files are in an acceptable format (TIFF (or JPEG), EPS (or PDF) or MS Office files) and with the correct resolution. If, together with your accepted article, you submit usable color figures then Elsevier will ensure, at no additional charge, that these figures will appear in color online (e.g., ScienceDirect and other sites). [Further information on the preparation of electronic artwork.](#)

Figure captions

Ensure that each illustration has a caption. Supply captions separately, not attached to the figure. A caption should comprise a brief title (not on the figure itself) and a description of the illustration. Keep text in the illustrations themselves to a minimum but explain all symbols and abbreviations used.

Tables

Please submit tables as editable text and not as images. Tables can be placed either next to the relevant text in the article, or on separate page(s) at the end. Number tables consecutively in accordance with their appearance in the text and place any table notes below the table body. Be sparing in the use of tables and ensure that the data presented in them do not duplicate results described elsewhere in the article. Please avoid using vertical rules and shading in table cells. References

Citation in text

Please ensure that every reference cited in the text is also present in the reference list (and vice versa). Any references cited in the abstract must be given in full. Unpublished results and personal communications are not recommended in the reference list, but may be mentioned in the text. If these

references are included in the reference list they should follow the standard reference style of the journal and should include a substitution of the publication date with either 'Unpublished results' or 'Personal communication'. Citation of a reference as 'in press' implies that the item has been accepted for publication.

Reference links

Increased discoverability of research and high quality peer review are ensured by online links to the sources cited. In order to allow us to create links to abstracting and indexing services, such as Scopus, Crossref and PubMed, please ensure that data provided in the references are correct. Please note that incorrect surnames, journal/book titles, publication year and pagination may prevent link creation. When copying references, please be careful as they may already contain errors. Use of the DOI is highly encouraged.

A DOI is guaranteed never to change, so you can use it as a permanent link to any electronic article.

An example of a citation using DOI for an article not yet in an issue is: VanDecar J.C., Russo R.M., James D.E., Ambeh W.B., Franke M. (2003). Aseismic continuation of the Lesser Antilles slab beneath northeastern Venezuela. *Journal of Geophysical Research*, <https://doi.org/10.1029/2001JB000884>. Please note the format of such citations should be in the same style as all other references in the paper.

Web references

As a minimum, the full URL should be given and the date when the reference was last accessed. Any further information, if known (DOI, author names, dates, reference to a source publication, etc.), should also be given. Web references can be listed separately (e.g., after the reference list) under a different heading if desired, or can be included in the reference list.

Data references

This journal encourages you to cite underlying or relevant datasets in your manuscript by citing them in your text and including a data reference in your Reference List. Data references should include the following elements: author name(s), dataset title, data repository, version (where available), year, and global persistent identifier. Add [dataset] immediately before the reference so we can properly identify it as a data reference. The [dataset] identifier will not appear in your published article.

Preprint references

Where a preprint has subsequently become available as a peer-reviewed publication, the formal publication should be used as the reference. If there are preprints that are central to your work or that cover crucial developments in the topic, but are not yet formally published, these may be referenced. Preprints should be clearly marked as such, for example by including the word preprint, or the name of the preprint server, as part of the reference. The preprint DOI should also be provided.

References in a special issue

Please ensure that the words 'this issue' are added to any references in the list (and any citations in the text) to other articles in the same Special Issue.

Reference management software

Most Elsevier journals have their reference template available in many of the most popular reference management software products. These include all products that support [Citation Style Language styles](#), such as [Mendeley](#). Using citation plug-ins from these products, authors only need to select the appropriate journal template when preparing their article, after which citations and bibliographies will be automatically formatted in the journal's style. If no template is yet available for this journal, please follow the format of the sample references and citations as shown in this Guide. If you use reference management software, please ensure that you remove all field codes before submitting the electronic manuscript. [More information on how to remove field codes from different reference management software](#).

Reference style

Text: Indicate references by superscript numbers in the text. The actual authors can be referred to, but the reference number(s) must always be given.

List: Number the references in the list in the order in which they appear in the text.

Examples:

Reference to a journal publication:

1. Van der Geer J, Hanraads JAJ, Lupton RA. The art of writing a scientific article. *J Sci Commun* 2010;163:51–9. <https://doi.org/10.1016/j.Sc.2010.00372>.

Reference to a journal publication with an article number:

2. Van der Geer J, Hanraads JAJ, Lupton RA. The art of writing a scientific article. *Heliyon*. 2018;19:e00205. <https://doi.org/j.heliyon.2018.e00205>.

Reference to a book:

3. Strunk Jr W, White EB. *The elements of style*. 4th ed. New York: Longman; 2000.

Reference to a chapter in an edited book:

4. Mettam GR, Adams LB. How to prepare an electronic version of your article. In: Jones BS, Smith RZ, editors. *Introduction to the electronic age*, New York: E-Publishing Inc; 2009, p. 281–304.

Reference to a website:

5. Cancer Research UK. Cancer statistics reports for the UK, <http://www.cancerresearchuk.org/aboutcancer/statistics/cancerstatsreport/>; 2003 [accessed 13 March 2003].

Reference to a dataset:

[dataset] 6. Oguro M, Imahiro S, Saito S, Nakashizuka T. Mortality data for Japanese oak wilt disease and surrounding forest compositions, Mendeley Data, v1; 2015. <https://doi.org/10.17632/xwj98nb39r.1>.

Note shortened form for last page number. e.g., 51–9, and that for more than 6 authors the first 6 should be listed followed by 'et al.' For further details you are referred to 'Uniform Requirements for Manuscripts submitted to Biomedical Journals' (J Am Med Assoc 1997;277:927–34)(see also [Samples of Formatted References](#)).

Journal abbreviations source

Journal names should be abbreviated according to the [List of Title Word Abbreviations](#).

Video

Elsevier accepts video material and animation sequences to support and enhance your scientific research. Authors who have video or animation files that they wish to submit with their article are strongly encouraged to include links to these within the body of the article. This can be done in the same way as a figure or table by referring to the video or animation content and noting in the body text where it should be placed. All submitted files should be properly labeled so that they directly relate to the video file's content. In order to ensure that your video or animation material is directly usable, please provide the file in one of our recommended file formats with a preferred maximum size of 150 MB per file, 1 GB in total. Video and animation files supplied will be published online in the electronic version of your article in Elsevier Web products, including [ScienceDirect](#). Please supply 'stills' with your files: you can choose any frame from the video or animation or make a separate image. These will be used instead of standard icons and will personalize the link to your video data. For more detailed instructions please visit our [video instruction pages](#). Note: since video and animation cannot be embedded in the print version of the journal, please provide text for both the electronic and the print version for the portions of the article that refer to this content.

Supplementary material

Supplementary material such as applications, images and sound clips, can be published with your article to enhance it. Submitted supplementary items are published exactly as they are received (Excel or PowerPoint files will appear as such online). Please submit your material together with the article and supply a concise, descriptive caption for each supplementary file. If you wish to make changes to supplementary material during any stage of the process, please make sure to provide an updated file. Do not annotate any corrections on a previous version. Please switch off the 'Track Changes' option in Microsoft Office files as these will appear in the published version.

Research data

This journal encourages and enables you to share data that supports your research publication where appropriate, and enables you to interlink the data with your published articles. Research data refers to the results of observations or experimentation that validate research findings. To facilitate reproducibility and data reuse, this journal also encourages you to share your software, code, models, algorithms, protocols, methods and other useful materials related to the project.

Below are a number of ways in which you can associate data with your article or make a statement about the availability of your data when submitting your manuscript. If you are sharing data in one of these ways, you are encouraged to cite the data in your manuscript and reference list. Please refer to the "References" section for more information about data citation. For more information on depositing, sharing and using research data and other relevant research materials, visit the [research data](#) page.

Data linking

If you have made your research data available in a data repository, you can link your article directly to the dataset. Elsevier collaborates with a number of repositories to link articles on ScienceDirect with relevant repositories, giving readers access to underlying data that gives them a better understanding of the research described.

There are different ways to link your datasets to your article. When available, you can directly link your dataset to your article by providing the relevant information in the submission system. For more information, visit the [database linking page](#).

For [supported data repositories](#) a repository banner will automatically appear next to your published article on ScienceDirect.

In addition, you can link to relevant data or entities through identifiers within the text of your manuscript, using the following format: Database: xxxx (e.g., TAIR: AT1G01020; CCDC: 734053; PDB: 1XFN).

Data statement

To foster transparency, we encourage you to state the availability of your data in your submission. This may be a requirement of your funding body or institution. If your data is unavailable to access or unsuitable to post, you will have the opportunity to indicate why during the submission process, for example by stating that the research data is confidential. The statement will appear with your published article on ScienceDirect. For more information, visit the [Data Statement page](#).

Proofs

One set of page proofs (as PDF files) will be sent by e-mail to the corresponding author (if we do not have an e-mail address then paper proofs will be sent by post) or a link will be provided in the e-mail so that authors can download the files themselves. To ensure a fast publication process of the article, we kindly ask authors to provide us with their proof corrections within two days. Elsevier now provides authors with PDF proofs which can be annotated; for this you will need to [download the free Adobe Reader](#), version 9 (or higher). Instructions on how to annotate PDF files will accompany the proofs (also given online). The exact system requirements are given at the [Adobe site](#).

If you do not wish to use the PDF annotations function, you may list the corrections (including replies to the Query Form) and return them to Elsevier in an e-mail. Please list your corrections quoting line number. If, for any reason, this is not possible, then mark the corrections and any other comments (including replies to the Query Form) on a printout of your proof and scan the pages and return via email. Please use this proof only for checking the typesetting, editing, completeness and correctness of the text, tables and figures. Significant changes to the article as accepted for publication will only be considered at this stage with permission from the Editor. We will do everything possible to get your article published quickly and accurately. It is important to ensure that all corrections are sent back to us in one communication: please check carefully before replying, as inclusion of any subsequent corrections cannot be guaranteed. Proofreading is solely your responsibility.

Offprints

The corresponding author will, at no cost, receive a customized [Share Link](#) providing 50 days free access to the final published version of the article on [ScienceDirect](#). The Share Link can be used for sharing the article via any communication channel, including email and social media. For an extra charge, paper offprints can be ordered via the offprint order form which is sent once the article is accepted for publication. Corresponding authors who have published their article gold open access do not receive a Share Link as their final published version of the article is available open access on ScienceDirect and can be shared through the article DOI link.

Visit the [Elsevier Support Center](#) to find the answers you need. Here you will find everything from Frequently Asked Questions to ways to get in touch.

You can also [check the status of your submitted article](#) or find out [when your accepted article will be published](#).

Appendix 4: Abstract of CHS symposium 2022 oral presentation

SCHOOL OF LABORATORY MEDICINE AND MEDICAL SCIENCES RESEARCH DAY

30 NOVEMBER 2022

SENATE CHMABER, WESTVILLE CAMPUS

INSTRUCTIONS:

- Please forward your abstracts electronically to dudhrajhp@ukzn.ac.za by **11 Nov 2022**.
- Participants should indicate their preference of presentation type (oral or poster).

CRITERIA FOR THE ACCEPTANCE OF ABSTRACTS:

- Abstracts are open to all postgraduate and post-doc students, developmental lecturers and credentialing staff in the School of Laboratory Medicine and Medical Sciences.
- Abstracts must adhere to the format indicated on the template provided.
- It is the author's responsibility to submit an abstract that is free of spelling and grammatical errors.
- All submissions must have ethical clearance.
- Abstracts will be reviewed by the scientific committee based on the following:
 - (1) Clear background/statement of the problem.
 - (2) Appropriateness of the methodology/study design to achieve the objectives.
 - (3) Clear presentation/significance of the results
 - (4) Relevance of the research findings; are the conclusions supported by the results.
 - (5) Novelty of the study.

GUIDELINES:

- Use Arial 11-point font throughout.
- The abstract must have a title: **IN BOLD TYPE AND UPPER CASE** (capitals).
- All authors must be listed: Surname followed by initials; do not include titles. (**Bold and underline** the presenter's name).
- Author affiliations must be shown: Department, Centre or Unit; Use * and # symbols to match affiliation with author.

Example: Taylor, M.^{*} Suleman, F.[#]

^{*}Department of Public Health; [#] Discipline of Pharmaceutical Sciences

- Abstract Layout: Please fit into the frame. **250 words max.**
 - **Background/Aim(s):** Clearly state the purpose(s) of the study.
 - **Methods:** Clearly state how your study was conducted, sample selection, tools and instruments used
 - **Results:** Present your results in a logical sequence
 - **Discussion/Conclusion:** Emphasize new and important aspects of the study and conclusions that are drawn from them.

PLEASE FILL IN ALL PRESENTER DETAILS

Presenter Information

Surname: Mzimela	First Name: Nhlakanipho
Research Theme: Prediabetes	Tel: 0655035416
E-Mail: 218006756@stu.ukzn.ac.za	Date: 23 November 2022

ABSTRACT TEMPLATE

Only abstracts that have strictly adhered to this template will be considered.
250 words max.

TITLE,
AUTHOR AND
AFFILIATION

THE ASSOCIATION BETWEEN INCRETIN PEPTIDES LEVELS AND THE DEVELOPMENT OF DIET-INDUCED PREDIABETES.

Mzimela, N.*; Khathi, A.*; Ngubane, P.S.*; Sosibo, A.M.*.

*Discipline of Human Physiology, School of Laboratory Medicine and Medical Sciences, College of Health Sciences, University of Kwa Zulu Natal, Westville, South Africa

ABSTRACT
BODY:

Prediabetes is a chronic metabolic condition that frequently occurs before type 2 diabetes mellitus develops. This condition is characterized by a gradual reduction of insulin sensitivity by insulin receptors in insulin-dependent cells, which is frequently followed by an increase in plasma glucose levels. These glucose levels, however, are still below the diabetes threshold. Pre-diabetic patients are more prone to have issues with immune dysregulation and diabetic nephropathy. Studies already conducted have looked on how incretin peptides affect the pathology of type 2 diabetes mellitus. The link between incretin peptide levels and the onset of prediabetes, on the other hand, remains unknown. Thus, the purpose of this study is to assess the role that incretin peptide levels play in the emergence of prediabetes. This study was conducted using male Sprague-Dawley rats, divided into the normal group (control) and the HFHC group (prediabetic control). The groups were given their diet accordingly for 20 weeks to induce prediabetes. After that, the animals were sacrificed, and blood was collected for final glucose readings. The findings of this study show that long-term use of HFHC or an unhealthy diet causes abnormal incretin concentrations, which trigger ongoing insulin stimulation. According to this study, incretin peptides contribute favorably to the onset of prediabetes. Therefore, GIP could be employed as therapeutic target and biomarker for early diagnosis of prediabetes.

Keywords: Type two diabetes mellitus, incretins, insulin resistance, prediabetes.

Research Theme: Prediabetes

Ethics Number: AREC/00003627/2021

Please tick the appropriate box

☐	Poster Presentation
☒	Oral Presentation

*Please follow the guidelines as abstracts will be reproduced exactly as submitted.

*The Title, Author affiliations and abstract must fit into the boxes above.