

ESSAY

Which New Health Technologies Do We Need to Achieve an End to HIV/AIDS?

Glenda E. Gray^{1,2*}, Fatima Laher², Tanya Doherty^{1,3,4}, Salim Abdool Karim^{5,6}, Scott Hammer⁷, John Mascola⁸, Chris Beyrer^{9,10}, Larry Corey¹¹

1 South African Medical Research Council, Cape Town, South Africa, **2** Perinatal HIV Research Unit, Faculty of Health Sciences, University of the Witwatersrand, Johannesburg, South Africa, **3** School of Public Health, University of Western Cape, Bellville, South Africa, **4** School of Public Health, University of the Witwatersrand, Johannesburg, South Africa, **5** Centre for the AIDS Programme of Research in South Africa, University of KwaZulu-Natal, Durban, South Africa, **6** Mailman School of Public Health, Columbia University, New York City, New York, United States of America, **7** Columbia University Medical Center, New York City, New York, United States of America, **8** Vaccine Research Center, National Institute of Allergy and Infectious Diseases, National Institutes of Health, Bethesda, Maryland, United States of America, **9** Johns Hopkins Bloomberg School of Public Health, Baltimore, Maryland, United States of America, **10** The International AIDS Society, Geneva, Switzerland, **11** HIV Vaccine Trials Network, Vaccine and Infectious Disease Division, Fred Hutchinson Cancer Research Center, Seattle, Washington

* Glenda.gray@mrc.ac.za



 OPEN ACCESS

Citation: Gray GE, Laher F, Doherty T, Abdool Karim S, Hammer S, Mascola J, et al. (2016) Which New Health Technologies Do We Need to Achieve an End to HIV/AIDS? *PLoS Biol* 14(3): e1002372. doi:10.1371/journal.pbio.1002372

Published: March 2, 2016

Copyright: This is an open access article, free of all copyright, and may be freely reproduced, distributed, transmitted, modified, built upon, or otherwise used by anyone for any lawful purpose. The work is made available under the [Creative Commons CC0](https://creativecommons.org/licenses/by/4.0/) public domain dedication.

Funding: The Article Processing Charges for this paper were funded by The Bill & Melinda Gates Foundation, who also provided funding for a commissioning fee of \$2,000. The Bill & Melinda Gates Foundation had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Competing Interests: The authors have declared that no competing interests exist.

Abbreviations: ADCC, antibody-dependent cellular cytotoxicity; ART, antiretroviral therapy; ARV, antiretroviral; CMV, cytomegalovirus; FDC, fixed-dose combination; GMP, good manufacturing practice; IAVI, International AIDS Vaccine Initiative; MSM, men who have sex with men; MTCT, mother-to-child transmission; MVA, Modified Vaccinia Ankara; NIH, National Institutes of Health; NHP, nonhuman primate; NRTI, Nucleoside/Nucleotide Reverse

Abstract

In the last 15 years, antiretroviral therapy (ART) has been the most globally impactful life-saving development of medical research. Antiretrovirals (ARVs) are used with great success for both the treatment and prevention of HIV infection. Despite these remarkable advances, this epidemic grows relentlessly worldwide. Over 2.1 million new infections occur each year, two-thirds in women and 240,000 in children. The widespread elimination of HIV will require the development of new, more potent prevention tools. Such efforts are imperative on a global scale. However, it must also be recognised that true containment of the epidemic requires the development and widespread implementation of a scientific advancement that has eluded us to date—a highly effective vaccine. Striving for such medical advances is what is required to achieve the end of AIDS.

In the last 15 years, antiretroviral therapy (ART) has been the most globally impactful life-saving development of medical research. Antiretrovirals (ARVs) are used with great success for both the treatment and prevention of HIV infection. In the United States, the widespread implementation of combination ARVs led to the virtual eradication of mother-to-child transmission of HIV from 1,650 cases in 1991 to 110 cases in 2011, and a turnaround in AIDS deaths from an almost 100% five-year mortality rate to a five-year survival rate of 91% in HIV-infected adults [1]. Currently, the estimated average lifespan of an HIV-infected adult in the developed world is well over 40 years post-diagnosis. Survival rates in the developing world, although lower, are improving: in sub-Saharan Africa, AIDS deaths fell by 39% between 2005 and 2013, and the biggest decline, 51%, was seen in South Africa [2].

Transcriptase Inhibitor; P5, Pox Protein Public Private Partnership; PMTCT, prevention of mother-to-child transmission; PrEP, pre-exposure prophylaxis; SGA, small for gestational age; SHIV, simian-human immunodeficiency virus; SIV, simian immunodeficiency virus; TAF, tenofovir alafenamide fumarate; TDF, tenofovir disoproxil fumarate; USMHRP, US Military HIV Research Program.

Provenance: The paper was commissioned by the Collection Coordinators, Gavin Yamey and Carlos Morel, in collaboration with the *PLOS Biology* and *PLOS Medicine* editors.

Furthermore, the association between ART, viremia, and transmission has led to the concept of “test and treat,” with the hope of reducing community viral load by testing early and initiating treatment as soon as a diagnosis of HIV is made [3]. Indeed, selected regions of the world have begun to actualize the public health value of ARVs, from gains in life expectancy to impact on onward transmission, with a potential 1% decline in new infections for every 10% increase in treatment coverage [2]. In September 2015, WHO released new guidelines removing all limitations on eligibility for ART among people living with HIV and recommending pre-exposure prophylaxis (PrEP) to population groups at significant HIV risk, paving the way for a global onslaught on HIV [4].

Despite these remarkable advances, this epidemic grows relentlessly worldwide. Over 2.1 million new infections occur each year, two-thirds in women and 240,000 in children [2]. In heavily affected countries, HIV infection rates have only stabilized at best: the annualized acquisition rates in persons in their first decade of sexual activity average 3%–5% yearly in southern Africa [5–7]. These figures are hardly compatible with the international health community’s stated goal of an “AIDS-free generation” [8,9]. In highly resourced settings, microepidemics of HIV still occur, particularly among gays, bisexuals, and men who have sex with men (MSM) [10]. HIV epidemics are expanding in two geographic regions in 2015—the Middle East/North Africa and Eastern Europe/Central Asia—largely due to challenges in implementing evidence-based HIV policies and programmes [2]. Even for the past decade in the US, almost 50,000 new cases recorded annually, two-thirds among MSM, has been a stable figure for years and shows no evidence of declining [1].

While treatment scale-up, medical male circumcision [11], and the implementation of strategies to prevent mother-to-child transmission [12] have received global traction, systemic or topical ARV-based biomedical advances to prevent sexual acquisition of HIV have, as yet, made limited impressions on a population basis, despite their reported efficacy. Factors such as their adherence requirements, cost, potential for drug resistance, and long-term feasibility have restricted the appetite for implementation, even though these approaches may reduce HIV incidence in select populations.

Already, several trials have shown that daily oral administration of the ARV tenofovir disoproxil fumarate (TDF), taken singly or in combination with emtricitabine, as PrEP by HIV-uninfected individuals, reduces HIV acquisition among serodiscordant couples (where one partner is HIV-positive and the other is HIV-negative) [13], MSM [14], at-risk men and women [15], and people who inject drugs [16,17] by between 44% and 75%. Long-acting injectable antiretroviral agents such as rilpivirine and cabotegravir, administered every two and three months, respectively, are also being developed for PrEP. All of these PrEP approaches are dependent on repeated HIV testing and adherence to drug regimens, which may challenge effectiveness in some populations and contexts.

The widespread elimination of HIV will require the development of new, more potent prevention tools. Because HIV acquisition occurs subclinically, the elimination of HIV on a population basis will require a highly effective vaccine. Alternatively, if vaccine development is delayed, supplementary strategies may include long-acting pre-exposure antiretroviral cocktails and/or the administration of neutralizing antibodies through long-lasting parenteral preparations or the development of a “genetic immunization” delivery system, as well as scaling up delivery of highly effective regimens to eliminate mother-to-child HIV transmission (Fig 1).

The Quest for an HIV Vaccine

HIV vaccine development has been challenging. The failure of five vaccine efficacy trials [17–21] and the modest efficacy of one [18–22] has limited the pharmaceutical industry’s role in

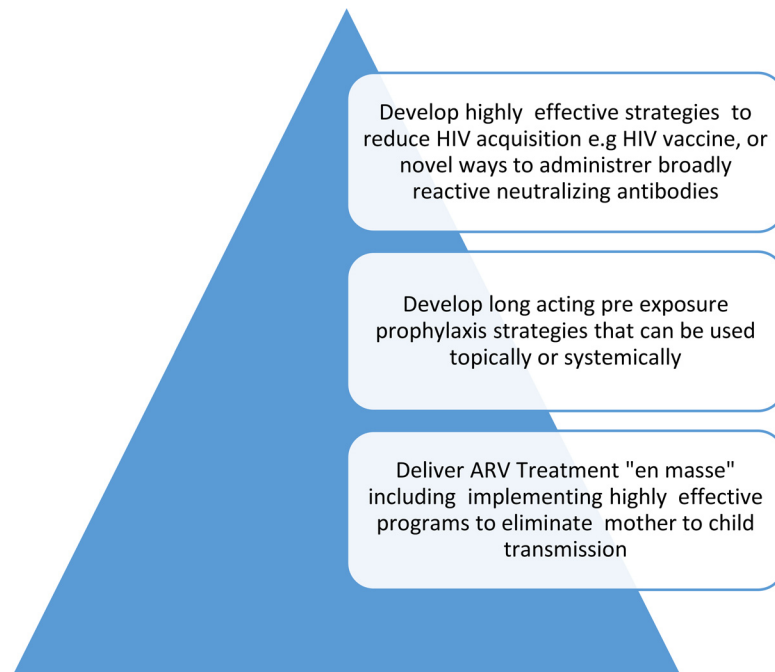


Fig 1. Medical interventions required to end the epidemic of HIV. Image credit: Glenda Gray.

doi:10.1371/journal.pbio.1002372.g001

HIV vaccine science. Current funding for HIV vaccine development is almost solely through government or philanthropic foundations [23]. The various approaches tested in late-stage clinical development, employing either viral vectors alone [17,18], in combination with DNA plasmids [21] or glycoprotein 120 [20], or glycoprotein 120 alone [19,20], have either been futile or have increased susceptibility to HIV acquisition in certain subgroups of vaccine recipients [17,18].

However, a milestone in the field was reached in late 2009, when the RV144 trial [24] of a recombinant canary pox prime and protein boost demonstrated modest efficacy with waning durability from 60% at 12 months to 31% at 36 months against sexually acquired HIV in Thailand. Although this vaccine regimen did not induce neutralizing antibodies, an immune correlates analysis indicated that it induced antibodies to the scaffolded V1V2 region of Env and that this response was associated with a reduced risk of infection [25,26]. Vaccine recipients who did not develop antibodies to the V1V2 scaffold demonstrated no vaccine efficacy, while those who developed antibodies to the V1V2 scaffold had an overall vaccine efficacy of 69% [27]. Additional analyses demonstrate that these V1V2 antibodies could mediate antibody-dependent cellular cytotoxicity (ADCC) and that the vaccine regimen induced immune pressure on viral sequences in the V1V2 region, providing increased support that this region is an important point of “attack” for nonneutralizing antibodies [28]. Other immune responses associated with vaccine efficacy in RV144 included polyfunctional CD4+ T cell responses as well as antibodies to envelope proteins of the IgG3 subclass [26,29]. These data, which suggest that nonneutralizing antibodies, especially to the V1V2 region of the HIV envelope, are associated with vaccine-induced protection, are provocative and require corroboration, as this V1V2 region had heretofore been largely ignored as a critical component of the human response to HIV.

Studies are underway to determine whether enhancing such responses by developing related vaccine prototypes with better T-cell priming as well as enhanced antibody responses will reproduce and enhance the vaccine efficacy seen in RV144. The Pox Protein Public Private

Partnership (P5) has designed an HIV vaccine program to be executed in the clade-C-predominant areas of southern Africa. The program will initially evaluate the role of alternate adjuvants coupled with a bivalent clade C protein in combination with a clade C optimised ALVAC vector as a means of developing a more potent and durable antibody response. More recently, with initial funding from the US National Institutes of Health (NIH), a group of collaborators at the Ragon Institute, the US Military HIV Research Program (USMHRP), the International AIDS Vaccine Initiative (IAVI), and, more recently, Jansen Pharmaceuticals have initiated development of a vaccine regimen that consists of a replication defective Adenovirus type 26 and Modified Vaccinia Ankara (MVA) vaccine in combination with a gp 140 protein boost that incorporates a series of synthetic Mosaic genes designed to make immune responses to all the major clades of HIV. Nonneutralizing antibodies appear to be the major correlate of immune protection in nonhuman primate (NHP) challenge models. These vaccines are now entering expanded human clinical trials.

Other approaches to develop an effective HIV vaccine are also underway and include novel vector strategies as well as structural biology-based approaches. A recent study demonstrated that a live replicating cytomegalovirus (CMV) vaccine encoding several simian immunodeficiency virus (SIV) proteins, administered prior to SIV challenge, can lead to complete viral suppression and elimination of detectable SIV infection in about half of the rhesus macaques studied, implying that some lentiviral infections may be susceptible to clearance via effector memory T cell-mediated mechanisms [30]. This study provides new insights into the role of effector memory CD8 T-cells in control and elimination of SIV infection and, more generally, how to elicit long-term tissue resident immune responses at the mucosal sites where HIV acquisition occurs. Whether similar immune responses are translatable to humans and can lead to continued clinical development of this live attenuated vaccine approach remains to be determined. In addition, further evaluation of the immune responses of the macaques that did not clear their SIV infection is imperative to further the understanding of the correlates of sustained SIV infection post CMV vaccination.

A worldwide effort to isolate broadly neutralizing antibodies to HIV and to understand the structural basis for their neutralization as well as the immunological basis for their development [31–33] has brought insights into the design of new candidate vaccines as well as antibody-targeted approaches.

A major impediment to advancing HIV vaccine development has been the painstakingly slow pace of conducting vaccine efficacy trials, largely due to the difficulty in producing good manufacturing practice (GMP) quantities of novel HIV envelope proteins or recombinant vectors with novel inserts. After RV144, it took five years to manufacture the next generation of recombinant pox/recombinant gp120 proteins. The effective manufacturing of novel, often complex, immunogens has occurred almost exclusively in an industry where much of the “lore” of consistently manufacturing reproducible amounts of biologics is collectively learned over time. Engaging industry to actively embrace such high-risk ventures has been difficult, but it is a necessary hurdle for the HIV vaccine field to solve. Developing such resolve and expertise is difficult in a field in which scientific pluralism is valued and required.

Despite the challenge of developing an HIV vaccine, mathematical modelling has verified the profound impact a vaccine with variable coverage—even low-efficacy vaccines in combination with other interventions—would have at a population level [34,35].

Neutralizing Antibodies for Prevention of HIV Acquisition

Numerous studies over the past 15 years have demonstrated the ability of HIV-1 neutralizing antibodies to prevent the acquisition of simian–human immunodeficiency virus (SHIV)

infection in NHPs [36–39]. Technologies to manufacture these antibodies in sufficient quantities are now available, enabling test-of-concept trials exploring whether the infusion of one or more antibodies could persistently reduce viremia in HIV-infected persons.

Perhaps more importantly, human clinical trials are being convened, exploring whether infusion of monoclonal antibodies could prevent HIV acquisition. If clinical trial proof was obtained that neutralizing antibodies could markedly reduce HIV acquisition, it would be a conceptual breakthrough in the HIV prevention landscape and would validate the need to develop a neutralizing antibody vaccine. Additionally, these antibodies could either be administered via long-term infusion or provided in a continuous concentration through “genetic immunization” of the antibody using a persistent vector such as adeno-associated virus vectors or via the insertion of the HIV antibody gene in plasmablasts [40]. Applying the knowledge gleaned from cancer, autoimmune, and inflammatory diseases to increase the potency of antibodies by improving effector function of antibodies, such as enhancing ADCC and increasing serum half-life of IgG through Fc engineering [41–43], may improve the utility of this intervention for HIV treatment and prevention.

Microbicides as an Innovation for HIV Prevention

Unlike the impressive breakthroughs with treatment, the quest to find potent biomedical prevention modalities for women’s routine use remains elusive. In sub-Saharan Africa, more than two-thirds of infections occur among women aged between 15 and 24 years. Young women typically acquire HIV infection 5–7 years earlier than their male counterparts [44,45]. HIV infections among women, especially young women, can be as much as 8-fold higher than in men of the same age [46] and is an important driver of the epidemic in Africa [47]. Candidate microbicides are being developed in an array of formulations, including gels, vaginal rings, rapidly dissolving vaginal tablets, films, and gel-filled compartments in diaphragms.

In 2010, the CAPRISA 004 trial, evaluating a coitally-dependent tenofovir gel regimen, first proved that ARVs can prevent sexual acquisition of HIV in women [48], followed by a study with MSM demonstrating the effectiveness of oral ARV chemoprophylaxis [14]. Adherence markedly affects the effectiveness of all PrEP regimens, and suboptimal adherence remains a large challenge for microbicide development [48,49], which was seen again in FACTS 001, a recent phase III confirmation study of coitally applied tenofovir gel [50]. To overcome these problems, long-acting, slow-release products are under development, including dapivirine or tenofovir in a vaginal ring [51,52]. In the future, multipurpose prevention technologies (e.g., for contraception and HIV prevention) and combination antiretroviral agent microbicides are in the works [53]. Rectal microbicide formulations for men and women at risk of HIV acquisition through anal intercourse are also under investigation [54].

ARV Treatment: New Drugs and Formulations: Do We Have What It Takes?

The continued development of safer, better priced, and more convenient ARVs has resulted in the ongoing improvement of health for HIV-infected persons on both an individual and population basis. In the developed world, over a million deaths were averted by ARV treatments in the last decade.

Still, there were 1.5 million AIDS-related deaths during 2013, many of them potentially preventable, as 60% of HIV-infected people and 76% of HIV-infected children still need to be placed on treatment [2]. The most formidable barrier to the ubiquitous use of treatment is access. Communities and health systems experience enormous infrastructural and human resource shortcomings to early testing and treatment, drug supply, and retention in care.

However, important innovations in drugs, treatment strategies, and monitoring are emerging that can solidify the expansion and resilience of treatment programs.

A drug development priority in a population-based scale-up of ART is a first-line single tablet that requires minimal laboratory monitoring and possesses minimal side effects with low rates of HIV resistance. Two recent advances on the classic TDF/emtricitabine/efavirenz first-line single tablet regimen promise to move us closer to this. First has been the coformulation of once-daily integrase inhibitors such as elvitegravir/cobicistat or dolutegravir. The latest US guidelines adopt the use of an integrase inhibitor with two nucleoside reverse transcriptase inhibitors as first-line therapy. In clinical trials, the single-pill TDF/emtricitabine/elvitegravir/cobicistat coformulation showed better safety profiles compared to its efavirenz-based peer [55]. Dolutegravir with a Nucleoside/Nucleotide Reverse Transcriptase Inhibitor (NRTI) backbone demonstrated superior ability to suppress viral load and better tolerability compared to efavirenz [56]. The second improvement has been the development of TDF's pro-drug, tenofovir alafenamide fumarate (TAF), which has also been coformulated in a single daily pill with emtricitabine/elvitegravir/cobicistat. The latter coformulation showed virologic responses similar to its TDF equivalent but had better renal and bone profiles. In the developing world, future scale-up could be eased by prioritizing in-country licensure and cost reduction of such new drugs.

Even with maximum improvements on a single daily pill, ultimate scale-up must consider critical issues affecting retention in care, such as the lack of money for transport [57] and the challenges of getting time off work for patients to attend a clinic and collect medication. Drug delivery approaches that would guarantee longer-term adherence, such as drug implants, are being studied. Furthermore, the landscape of health staffing may need to transform, as the necessity of increasing personnel to handle increased patient volumes may require shifting tasks even further from nurses to trained counsellors to assist with monitoring and retention in care (Table 1).

Here and Now: Eliminating Paediatric HIV

The most dramatic alteration in the HIV epidemic has been the marked reduction in HIV infection in children [2]. The Promise study, a recent randomised clinical trial conducted in Africa, shows that a globally applicable ART regimen is available that could reduce mother-to-child transmission of HIV during pregnancy or delivery to less than 1% [58]. A multicountry pre-exposure prophylaxis trial in infants found similar infection rates of 1.4% and 1.5% in the two arms at 50 weeks [59] and provides an alternative preventive option for women who are not ready for a lifetime commitment to ARV treatment as part of a multipronged prevention of mother-to-child transmission (PMTCT) programme [60].

Table 1. Innovations for scaling up care and keeping people on treatment.

Intervention	Innovation
Diagnostics	Non-blood-based diagnostics that are available "over the counter"
Antiretroviral Treatment	<ul style="list-style-type: none"> • Coformulations of once-daily pills with minimal side effects and high genetic barrier for resistance • Long-acting antiretrovirals minimising need for regular interface with health care providers
Virological and Immunological Monitoring	Point-of-care viral load allows testing, results feedback, and responsive care recommendations at the same visit
Task Shifting	Community health care workers delivering care because drugs are safe and monitoring is simple

doi:10.1371/journal.pbio.1002372.t001

Table 2. Potential demand-side and health system innovations to control paediatric HIV.

Innovation	Benefits
Scaling up community-based delivery platforms [64]	Access to universal health coverage
Training of community health workers [65]	Improve coverage along the continuum of care through early identification of pregnant women, encouragement of early antenatal booking, HIV testing, initiation of ARV treatment, and support for lifelong adherence to medication
Universal testing of infants [66]	Increasing access to the early identification of HIV infected infants and rapid initiation of ARV treatment [67]

doi:10.1371/journal.pbio.1002372.t002

These findings must be rapidly operationalised to curtail the quarter of a million new childhood infections that occur annually. Here, again, the vexing issue is access, highlighting the kind of health infrastructure investments required to identify and treat all pregnant women with HIV (Table 2). South Africa is the first country to report on population-level effectiveness of a national PMTCT programme [61] and has demonstrated a consistent trend toward lowering transmission among infants, with rates of 2.6% in 2012–2013 [61,62]. The effectiveness of PMTCT interventions in the field are influenced by access to prenatal care, retention in care, ARV adherence, adequate education, and follow-up of the mother and infant post-delivery (Table 2). To achieve adequate viral load suppression at the time of delivery, ARVs should be initiated early in pregnancy [12]. Even in South Africa, where over 85% of pregnant women attend at least four antenatal care visits, in 2014–2015 around half (54%) of pregnant women received their first antenatal care visit before 20 weeks of pregnancy [63].

Incident cases of HIV occurring during pregnancy and lactation increase the risk of mother-to-child transmission. Implementing repeat HIV testing in late pregnancy and during breastfeeding will be a further critical step required on the road to elimination [68]. Recommendations to repeat HIV testing around 32 weeks of pregnancy have been erratically implemented in South Africa, with only 22% of women in 2012–2013 reporting that they had their last HIV test at or beyond 32 weeks [62].

Hopefully, as more women are initiated on lifelong treatment, the continuum of care will improve so that prevention benefits are realised. One report from Malawi, however, acts as a cautionary tale: one in ten women were lost to follow-up six months after ART initiation (Table 3) [69]. Furthermore, solving the issues of health care stigmatization, access, status disclosure, and spousal involvement will also be needed to improve treatment initiation, adherence, and retention in care [70].

Because ART has successfully reduced MTCT and lowered morbidity and mortality in pregnant women, it has also led to growing numbers of HIV-exposed but uninfected children. Increased morbidity and mortality amongst HIV-exposed uninfected children has been described, including birth defects [71], small for gestational age (SGA) infants [72], growth [73] and neurodevelopmental delay [74], and reduced immunity to vaccine-preventable diseases [75,76]. There is an urgent need to better understand the health consequences of

Table 3. Challenges in keeping HIV-positive pregnant women in care [69].

- Women initiated on lifelong ARVs during pregnancy were five times more likely than women who started ARVs in WHO stage 3/4 or with a CD4 cell count of 350 cells/ml or less to never return after their initial clinic visit.
- Women initiating lifelong ARVs while breastfeeding were twice as likely to miss their first follow-up visit.
- Loss to follow-up was highest in pregnant women who began lifelong ARVs at large clinics on the day they were diagnosed with HIV.

doi:10.1371/journal.pbio.1002372.t003

Table 4. Current biomedical interventions that are capable of bending the HIV epidemic curve.

Intervention	R&D innovation	Source of financing
1. Health system strengthening	Smart clinic approaches; tools for procurement; health system processes development	Financial investments in health infrastructure, human resource development, health care worker training, continual monitoring and evaluation
2. Universal access to frequent HIV testing*	Over-the-counter HIV diagnostics, R&D into non-blood-based diagnostics, Point of care HIV diagnostics for infants, developing HIV counselling on mobile platforms.	Government & donor investment into biotechnology that simplifies point of care diagnostics whilst maintaining sensitivity and specificity, training of health care workers and community health workers on the mass rollout of HIV testing, values clarification for those carrying out HIV testing to reduce stigmatizing attitudes
3. Making ARV treatment available to all HIV-infected individuals, irrespective of CD4 count	R&D in fixed drug combinations, long-acting ARVs with high barriers to resistance and low side effects	Pharmaceutical investment with government subsidies, low cost of ARVs, task shifting to community health care workers
4. Medical male circumcision for neonates, adolescent boys, and adults*	Low-cost devices for male adults and neonates to allow mass medical circumcision without doctor supervision	Medical device investment, training health care workers in neonatal circumcision, task shifting.
5. Rolling out PrEP*	Continue R&D into long-acting ARV agents	Pharmaceutical investment with government subsidies
6. Universal PMTCT*	Safe fixed-dose combination (FDC) ARV, point of care virological and immunological monitoring tools	Government commitment to implementing HIV testing at family planning, improving access to safe contraception, allowing for safe termination of pregnancy, ARVs for life, task shifting.
7. Safe needle exchange*	Cheap disposable needles	Government support of clean needle exchange programmes

*Condom provision at every opportunity

doi:10.1371/journal.pbio.1002372.t004

exposure to HIV and antiretroviral drugs on HIV-uninfected children and to improve monitoring and management of this growing subpopulation of children.

HIV Cure

HIV infection results in latent infection of a select population of memory CD4+ T cells within days post-acquisition [77]. When the integrated HIV proviral DNA is transcriptionally silent, these resting cells are not affected by ARVs and are not recognised by the immune system. HIV can reactivate from these cells, however, and their presence constitutes a major barrier to

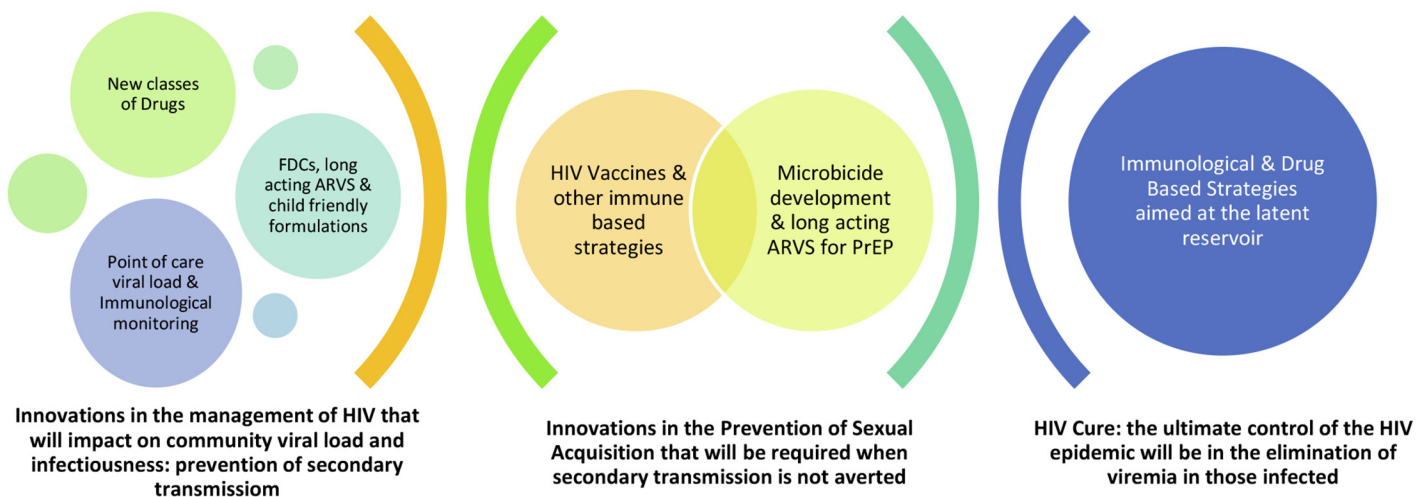


Fig 2. The spectrum of biomedical innovation required to end AIDS. Image credit: Glenda Gray.

doi:10.1371/journal.pbio.1002372.g002

eradicating infection [78,79]. Advances in understanding the basic biology of HIV have provided a framework to conceptualise novel approaches to curing the disease. These include genetic engineering technologies to either protect CD4+ T cells from future infection or deliver probes to inactivate integrated HIV DNA in the cell [80]. Use of genetically engineered T+ cells to provide long-term immune surveillance or to activate latently infected cells and destroy them and any associated HIV virions before they proliferate are being investigated [80]. Whether these strategies can either singly or in combination result in a true cure or what the field has termed a “functional” cure (that is, the ability to maintain viral suppression without ARVs with no risk of sexual or perinatal transmission) remains to be determined [80]. The benefits of such an approach on interrupting the spread of HIV are enormous and warrant a full-scale effort by the international scientific community.

Conclusion

HIV brought an unprecedented shift in the global landscape of public health: it slashed life expectancy and wiped out a generation of economically active adults in their prime in sub-Saharan Africa; it reversed gains in under-five mortality and created a cohort of AIDS orphans; and it exposed the underbelly of poor health systems. It has also revealed the interrelatedness between social behaviour, stigmatization, cultural mores, religious beliefs, and human health. HIV has challenged all societies in our attempts to deal with its economic, societal, and medical aspects.

Even with all the biomedical and behavioural tools available today, HIV continues to be a formidable pathogen, altering the health economics and public health strategies of most countries worldwide. Of the 35 million HIV-infected individuals worldwide in 2014, more than half did not know their HIV status, and over a third were not receiving ARVs [1] despite the availability of affordable point-of-care diagnostics and treatments. Adoption of “universal test and treat” approaches, extension of medical male circumcision programs, universal access to basic harm reduction services for people who inject drugs, and more widespread use of PrEP in targeted populations can do much to “bend the curve” and initiate a process to slow the rate of new HIV infections (Table 4). Such efforts are imperative on a global scale. However, it must also be recognised that true containment of the epidemic requires the development and widespread implementation of a scientific advance that has eluded us to date—a highly effective vaccine. There are potential synergies that will accrue from an integrated approach involving treatment, microbicides, and HIV vaccines (Fig 2). Various mathematical models agree that treatment roll-out on its own will decrease HIV incidence over time. However, when a 30% preventative HIV vaccine was introduced to a model with expanding treatment access in southern Africa, incidence was predicted to be 67% lower over time compared to a scenario with no vaccine introduction [35].

References

1. Centers for Disease Control and Prevention. HIV Surveillance Report Volume 24: 2012. 2012.
2. UNAIDS. The Gap Report 2013. Geneva: UNAIDS, 2014.
3. Granich RM, Gilks CF, Dye C, De Cock KM, Williams BG. Universal voluntary HIV testing with immediate antiretroviral therapy as a strategy for elimination of HIV transmission: a mathematical model. *Lancet*. 2009; 373(9657):48–57. Epub 2008/11/29. doi: [10.1016/S0140-6736\(08\)61697-9](https://doi.org/10.1016/S0140-6736(08)61697-9) PMID: [19038438](https://pubmed.ncbi.nlm.nih.gov/19038438/).
4. World Health Organisation. Guideline on when to start antiretroviral therapy and on pre-exposure prophylaxis for HIV. Geneva: WHO, 2015.
5. Padian NS, van der Straten A, Ramjee G, Chipato T, de Bruyn G, Blanchard K, et al. Diaphragm and lubricant gel for prevention of HIV acquisition in southern African women: a randomised controlled trial. *The Lancet*. 370(9583):251–61. doi: [10.1016/S0140-6736\(07\)60950-7](https://doi.org/10.1016/S0140-6736(07)60950-7)

6. Van Damme L, Corneli A, Ahmed K, Agot K, Lombaard J, Kapiga S, et al. Preexposure Prophylaxis for HIV Infection among African Women. *New England Journal of Medicine*. 2012; 367(5):411–22. doi: [10.1056/NEJMoa1202614](https://doi.org/10.1056/NEJMoa1202614) PMID: [22784040](https://pubmed.ncbi.nlm.nih.gov/22784040/).
7. Auvert B, Taljaard D, Lagarde E, Sobngwi-Tambekou J, Sitta R, Puren A. Randomized, Controlled Intervention Trial of Male Circumcision for Reduction of HIV Infection Risk: The ANRS 1265 Trial. *PLoS Med*. 2005; 2(11):e298. doi: [10.1371/journal.pmed.0020298](https://doi.org/10.1371/journal.pmed.0020298) PMID: [16231970](https://pubmed.ncbi.nlm.nih.gov/16231970/)
8. Goosby E. The President's Emergency Plan for AIDS Relief: marshalling all tools at our disposal toward an AIDS-free generation. *Health Aff (Millwood)*. 2012; 31(7):1593–8. Epub 2012/07/11. doi: [10.1377/hlthaff.2012.0241](https://doi.org/10.1377/hlthaff.2012.0241) PMID: [22778350](https://pubmed.ncbi.nlm.nih.gov/22778350/).
9. Abdool Karim SS. An AIDS-free generation? *Science*. 2012; 337(6091):133. Epub 2012/07/17. doi: [10.1126/science.1226535](https://doi.org/10.1126/science.1226535) PMID: [22798569](https://pubmed.ncbi.nlm.nih.gov/22798569/).
10. Beyrer C, Baral SD, van Griensven F, Goodreau SM, Chariyalertsak S, Wirtz AL, et al. Global epidemiology of HIV infection in men who have sex with men. *Lancet*. 2012; 380(9839):367–77. doi: [10.1016/S0140-6736\(12\)60821-6](https://doi.org/10.1016/S0140-6736(12)60821-6) PMID: [22819660](https://pubmed.ncbi.nlm.nih.gov/22819660/); PubMed Central PMCID: PMC3805037.
11. Siegfried N, Muller M, Deeks JJ, Volmink J. Male circumcision for prevention of heterosexual acquisition of HIV in men. *Cochrane Database Syst Rev*. 2009;(2):Cd003362. Epub 2009/04/17. doi: [10.1002/14651858.CD003362.pub2](https://doi.org/10.1002/14651858.CD003362.pub2) PMID: [19370585](https://pubmed.ncbi.nlm.nih.gov/19370585/).
12. World Health Organisation. Use of antiretroviral drugs for treating pregnant women and preventing HIV infection in infants: Programmatic update. Geneva: WHO, 2012.
13. Baeten JM, Donnell D, Ndase P, Mugo NR, Campbell JD, Wangisi J, et al. Antiretroviral prophylaxis for HIV prevention in heterosexual men and women. *The New England journal of medicine*. 2012; 367(5):399–410. Epub 2012/07/13. PMID: [22784037](https://pubmed.ncbi.nlm.nih.gov/22784037/). doi: [10.1056/NEJMoa1108524](https://doi.org/10.1056/NEJMoa1108524)
14. Grant RM, Lama JR, Anderson PL, McMahan V, Liu AY, Vargas L, et al. Preexposure chemoprophylaxis for HIV prevention in men who have sex with men. *The New England journal of medicine*. 2010; 363(27):2587–99. Epub 2010/11/26. doi: [10.1056/NEJMoa1011205](https://doi.org/10.1056/NEJMoa1011205) PMID: [21091279](https://pubmed.ncbi.nlm.nih.gov/21091279/).
15. Thigpen MC, Kebaabetswe PM, Paxton LA, Smith DK, Rose CE, Segolodi TM, et al. Antiretroviral pre-exposure prophylaxis for heterosexual HIV transmission in Botswana. *The New England journal of medicine*. 2012; 367(5):423–34. Epub 2012/07/13. PMID: [22784038](https://pubmed.ncbi.nlm.nih.gov/22784038/). doi: [10.1056/NEJMoa1110711](https://doi.org/10.1056/NEJMoa1110711)
16. Abdool Karim SS. HIV pre-exposure prophylaxis in injecting drug users. *Lancet*. 2013; 381(9883):2060–2. doi: [10.1016/S0140-6736\(13\)61140-X](https://doi.org/10.1016/S0140-6736(13)61140-X) PMID: [23769217](https://pubmed.ncbi.nlm.nih.gov/23769217/).
17. Choopanya K, Martin M, Suntharasamai P, Sangkum U, Mock PA, Leethochawalit M, et al. Antiretroviral prophylaxis for HIV infection in injecting drug users in Bangkok, Thailand (the Bangkok Tenofovir Study): a randomised, double-blind, placebo-controlled phase 3 trial. *The Lancet*. 2013; 381(9883):2083–90.
18. Buchbinder SP, Mehrotra DV, Duerr A, Fitzgerald DW, Mogg R, Li D, et al. Efficacy assessment of a cell-mediated immunity HIV-1 vaccine (the Step Study): a double-blind, randomised, placebo-controlled, test-of-concept trial. *The Lancet*. 372(9653):1881–93. doi: [10.1016/S0140-6736\(08\)61591-3](https://doi.org/10.1016/S0140-6736(08)61591-3)
19. Gray GE, Allen M, Moodie Z, Churchyard G, Bekker L-G, Nchabeleng M, et al. Safety and efficacy of the HVTN 503/Phambili Study of a clade-B-based HIV-1 vaccine in South Africa: a double-blind, randomised, placebo-controlled test-of-concept phase 2b study. *The Lancet infectious diseases*. 11(7):507–15. doi: [10.1016/S1473-3099\(11\)70098-6](https://doi.org/10.1016/S1473-3099(11)70098-6) PMID: [21570355](https://pubmed.ncbi.nlm.nih.gov/21570355/)
20. Pitisuttithum P, Gilbert P, Gurwith M, Heyward W, Martin M, van Griensven F, et al. Randomized, Double-Blind, Placebo-Controlled Efficacy Trial of a Bivalent Recombinant Glycoprotein 120 HIV-1 Vaccine among Injection Drug Users in Bangkok, Thailand. *Journal of Infectious Diseases*. 2006; 194(12):1661–71. doi: [10.1086/508748](https://doi.org/10.1086/508748) PMID: [17109337](https://pubmed.ncbi.nlm.nih.gov/17109337/)
21. Russell ND, Graham BS, Keefer MC, McElrath MJ, Self SG, Weinhold KJ, et al. Phase 2 Study of an HIV-1 Canarypox Vaccine (vCP1452) Alone and in Combination With rgp120: Negative Results Fail to Trigger a Phase 3 Correlates Trial. *JAIDS Journal of Acquired Immune Deficiency Syndromes*. 2007; 44(2):203–12 doi: [10.1097/01.qai.0000248356.48501.ff](https://doi.org/10.1097/01.qai.0000248356.48501.ff) PMID: [17106277](https://pubmed.ncbi.nlm.nih.gov/17106277/)
22. Hammer SM, Sobieszczyk ME, Janes H, Karuna ST, Mulligan MJ, Grove D, et al. Efficacy Trial of a DNA/rAd5 HIV-1 Preventive Vaccine. *New England Journal of Medicine*. 2013; 369(22):2083–92. doi: [10.1056/NEJMoa1310566](https://doi.org/10.1056/NEJMoa1310566) PMID: [24099601](https://pubmed.ncbi.nlm.nih.gov/24099601/).
23. AVAC. AVAC REPORT 2013: Research & Reality. New York: AVAC, 2013.
24. Rerks-Ngarm S, Pitisuttithum P, Nitayaphan S, Kaewkungwal J, Chiu J, Paris R, et al. Vaccination with ALVAC and AIDSVAX to prevent HIV-1 infection in Thailand. *The New England journal of medicine*. 2009; 361(23):2209–20. Epub 2009/10/22. doi: [10.1056/NEJMoa0908492](https://doi.org/10.1056/NEJMoa0908492) PMID: [19843557](https://pubmed.ncbi.nlm.nih.gov/19843557/).
25. Haynes BF, Gilbert PB, McElrath MJ, Zolla-Pazner S, Tomaras GD, Alam SM, et al. Immune-correlates analysis of an HIV-1 vaccine efficacy trial. *The New England journal of medicine*. 2012; 366(14):1275–86. doi: [10.1056/NEJMoa1113425](https://doi.org/10.1056/NEJMoa1113425) PMID: [22475592](https://pubmed.ncbi.nlm.nih.gov/22475592/); PubMed Central PMCID: PMC3371689.

26. Yates NL, Liao HX, Fong Y, deCamp A, Vandergriff NA, Williams WT, et al. Vaccine-induced Env V1-V2 IgG3 correlates with lower HIV-1 infection risk and declines soon after vaccination. *Science translational medicine*. 2014; 6(228):228ra39. doi: [10.1126/scitranslmed.3007730](https://doi.org/10.1126/scitranslmed.3007730) PMID: [24648342](https://pubmed.ncbi.nlm.nih.gov/24648342/); PubMed Central PMCID: PMC4116665.
27. Liao HX, Bonsignori M, Alam SM, McLellan JS, Tomaras GD, Moody MA, et al. Vaccine induction of antibodies against a structurally heterogeneous site of immune pressure within HIV-1 envelope protein variable regions 1 and 2. *Immunity*. 2013; 38(1):176–86. doi: [10.1016/j.immuni.2012.11.011](https://doi.org/10.1016/j.immuni.2012.11.011) PMID: [23313589](https://pubmed.ncbi.nlm.nih.gov/23313589/); PubMed Central PMCID: PMC3569735.
28. Rolland M, Edlefsen PT, Larsen BB, Tovnanubutra S, Sanders-Buell E, Hertz T, et al. Increased HIV-1 vaccine efficacy against viruses with genetic signatures in Env V2. *Nature*. 2012; 490(7420):417–20. doi: [10.1038/nature11519](https://doi.org/10.1038/nature11519) PMID: [22960785](https://pubmed.ncbi.nlm.nih.gov/22960785/); PubMed Central PMCID: PMC3551291.
29. Lin L, Finak G, Ushey K, Seshadri C, Hawn TR, Frahm N, et al. Combinatorial polyfunctionality analysis of antigen-specific T-cell subsets identifies novel cellular subsets correlated with clinical outcomes. *Nature biotechnology*. 2015; In Press.
30. Hansen SG, Piatak M Jr., Ventura AB, Hughes CM, Gilbride RM, Ford JC, et al. Immune clearance of highly pathogenic SIV infection. *Nature*. 2013; 502(7469):100–4. Epub 2013/09/13. doi: [10.1038/nature12519](https://doi.org/10.1038/nature12519) PMID: [24025770](https://pubmed.ncbi.nlm.nih.gov/24025770/); PubMed Central PMCID: PMC3849456.
31. Moore PL, Gray ES, Wibmer CK, Bhiman JN, Nonyane M, Sheward DJ, et al. Evolution of an HIV glycan-dependent broadly neutralizing antibody epitope through immune escape. *Nature medicine*. 2012; 18(11):1688–92. doi: [10.1038/nm.2985](https://doi.org/10.1038/nm.2985) <http://www.nature.com/nm/journal/v18/n11/abs/nm.2985.html#supplementary-information>. PMID: [23086475](https://pubmed.ncbi.nlm.nih.gov/23086475/)
32. Doria-Rose NA, Schramm CA, Gorman J, Moore PL, Bhiman JN, DeKosky BJ, et al. Developmental pathway for potent V1V2-directed HIV-neutralizing antibodies. *Nature*. 2014; 509(7498):55–62. doi: [10.1038/nature13036](https://doi.org/10.1038/nature13036) <http://www.nature.com/nature/journal/v509/n7498/abs/nature13036.html#supplementary-information>. PMID: [24590074](https://pubmed.ncbi.nlm.nih.gov/24590074/)
33. Kong R, Louder MK, Wagh K, Bailer RT, deCamp A, Greene K, et al. Improving Neutralization Potency and Breadth by Combining Broadly Reactive HIV-1 Antibodies Targeting Major Neutralization Epitopes. *Journal of virology*. 2014. Epub 2014/12/19. doi: [10.1128/jvi.03136-14](https://doi.org/10.1128/jvi.03136-14) PMID: [25520506](https://pubmed.ncbi.nlm.nih.gov/25520506/).
34. Gray RH, Li X, Wawer MJ, Gange SJ, Serwadda D, Sewankambo NK, et al. Stochastic simulation of the impact of antiretroviral therapy and HIV vaccines on HIV transmission; Rakai, Uganda. *AIDS*. 2003; 17(13):1941–51. Epub 2003/09/10. doi: [10.1097/01.aids.0000076313.76477.fc](https://doi.org/10.1097/01.aids.0000076313.76477.fc) PMID: [12960827](https://pubmed.ncbi.nlm.nih.gov/12960827/).
35. Phillips AN, Cambiano V, Nakagawa F, Ford D, Lundgren JD, Roset-Bahmanyar E, et al. Potential future impact of a partially effective HIV vaccine in a southern African setting. *PLoS ONE*. 2014; 9(9): e107214. Epub 2014/09/11. doi: [10.1371/journal.pone.0107214](https://doi.org/10.1371/journal.pone.0107214) PMID: [25207973](https://pubmed.ncbi.nlm.nih.gov/25207973/); PubMed Central PMCID: PMC4160197.
36. Shingai M, Donau OK, Plishka RJ, Buckler-White A, Mascola JR, Nabel GJ, et al. Passive transfer of modest titers of potent and broadly neutralizing anti-HIV monoclonal antibodies block SHIV infection in macaques. *The Journal of experimental medicine*. 2014; 211(10):2061–74. Epub 2014/08/27. doi: [10.1084/jem.20132494](https://doi.org/10.1084/jem.20132494) PMID: [25155019](https://pubmed.ncbi.nlm.nih.gov/25155019/); PubMed Central PMCID: PMC4172223.
37. Mascola JR, Lewis MG, Stiegler G, Harris D, VanCott TC, Hayes D, et al. Protection of Macaques against pathogenic simian/human immunodeficiency virus 89.6PD by passive transfer of neutralizing antibodies. *Journal of virology*. 1999; 73(5):4009–18. Epub 1999/04/10. PMID: [10196297](https://pubmed.ncbi.nlm.nih.gov/10196297/); PubMed Central PMCID: PMC4104180.
38. Baba TW, Liska V, Hofmann-Lehmann R, Vlasak J, Xu W, Ayehunie S, et al. Human neutralizing monoclonal antibodies of the IgG1 subtype protect against mucosal simian-human immunodeficiency virus infection. *Nature medicine*. 2000; 6(2):200–6. Epub 2000/02/02. doi: [10.1038/72309](https://doi.org/10.1038/72309) PMID: [10655110](https://pubmed.ncbi.nlm.nih.gov/10655110/).
39. Parren PW, Marx PA, Hessel AJ, Luckay A, Harouse J, Cheng-Mayer C, et al. Antibody protects macaques against vaginal challenge with a pathogenic R5 simian/human immunodeficiency virus at serum levels giving complete neutralization in vitro. *Journal of virology*. 2001; 75(17):8340–7. Epub 2001/08/03. PMID: [11483779](https://pubmed.ncbi.nlm.nih.gov/11483779/); PubMed Central PMCID: PMC115078.
40. Kwong PD, Mascola JR, Nabel GJ. Broadly neutralizing antibodies and the search for an HIV-1 vaccine: the end of the beginning. *Nature reviews Immunology*. 2013; 13(9):693–701. doi: [10.1038/nri3516](https://doi.org/10.1038/nri3516) PMID: [23969737](https://pubmed.ncbi.nlm.nih.gov/23969737/).
41. Hinton PR, Johlf MG, Xiong JM, Hanestad K, Ong KC, Bullock C, et al. Engineered Human IgG Antibodies with Longer Serum Half-lives in Primates. *Journal of Biological Chemistry*. 2004; 279(8):6213–6. doi: [10.1074/jbc.C300470200](https://doi.org/10.1074/jbc.C300470200) PMID: [14699147](https://pubmed.ncbi.nlm.nih.gov/14699147/)
42. Niwa R, Hatanaka S, Shoji-Hosaka E, Sakurada M, Kobayashi Y, Uehara A, et al. Enhancement of the antibody-dependent cellular cytotoxicity of low-fucose IgG1 Is independent of FcγRIIIa functional polymorphism. *Clinical cancer research: an official journal of the American Association for Cancer*

- Research. 2004; 10(18 Pt 1):6248–55. Epub 2004/09/28. doi: [10.1158/1078-0432.ccr-04-0850](https://doi.org/10.1158/1078-0432.ccr-04-0850) PMID: [15448014](https://pubmed.ncbi.nlm.nih.gov/15448014/).
43. Chan AC, Carter PJ. Therapeutic antibodies for autoimmunity and inflammation. *Nature reviews Immunology*. 2010; 10(5):301–16. Epub 2010/04/24. doi: [10.1038/nri2761](https://doi.org/10.1038/nri2761) PMID: [20414204](https://pubmed.ncbi.nlm.nih.gov/20414204/).
 44. Abdool Karim Q, Abdool Karim SS, Singh B, Short R, Ngxongo S. Seroprevalence of HIV infection in rural South Africa. *AIDS*. 1992; 6:1535–9. PMID: [1492937](https://pubmed.ncbi.nlm.nih.gov/1492937/)
 45. Shisana O, Rehle T, Simbayi LC, Zuma K, Jooste S, Pillay-van-Wyk V, et al. South African National HIV Prevalence, Incidence, Behaviour and Communication Survey 2008: A turning tide among teenagers? Cape Town: HSRC Press, 2009.
 46. UNAIDS. UNAIDS Report on the global AIDS Epidemic 2010. <http://www.unaids.org/globalreport/> (Accessed 11 February 2014). Geneva: Joint United Nations Programme on HIV/AIDS, 2010.
 47. Abdool Karim Q, Kharsany AB, Frohlich JA, Werner L, Mashego M, Mlotshwa M, et al. Stabilizing HIV prevalence masks high HIV incidence rates amongst rural and urban women in KwaZulu-Natal, South Africa. *International journal of epidemiology*. 2011; 40(4):922–30. doi: [10.1093/ije/dyq176](https://doi.org/10.1093/ije/dyq176) PMID: [21047913](https://pubmed.ncbi.nlm.nih.gov/21047913/); PubMed Central PMCID: PMC3156366.
 48. Abdool Karim Q, Abdool Karim SS, Frohlich JA, Grobler AC, Baxter C, Mansoor LE, et al. Effectiveness and Safety of Tenofovir Gel, an Antiretroviral Microbicide, for the Prevention of HIV Infection in Women. *Science*. 2010; 329:1168–74. doi: [10.1126/science.1193748](https://doi.org/10.1126/science.1193748) PMID: [20643915](https://pubmed.ncbi.nlm.nih.gov/20643915/)
 49. Marrazzo J, Ramjee G, Nair G, Palanee T, Mkhize B, Nakabiito C, et al., editors. Pre-exposure Prophylaxis for HIV in Women: Daily Oral Tenofovir, Oral Tenofovir/Emtricitabine, or Vaginal Tenofovir Gel in the VOICE Study (MTN 003). [Abstract #26LB]. 20th Conference of Retroviruses and Opportunistic Infections; 2013 March 3–6; Atlanta, USA.
 50. Rees H, Delany-Moretlwe S, Lombard C, Baron D, Panchia R, Myer L, et al. FACTS 001 Phase III Trial of Pericoital Tenofovir 1% Gel for HIV Prevention in Women. Conference on Retroviruses and Opportunistic Infections (CROI); Seattle2015.
 51. International Partnership for Microbicides. IPM 027: Safety and Efficacy Trial of a Dapivirine Vaginal Matrix Ring in Healthy HIV-Negative Women; <http://clinicaltrials.gov/show/NCT01539226> (last accessed 2 August 2012). 2012.
 52. International Partnership for Microbicides. MTN-020: Phase 3 Safety and Effectiveness Trial of Dapivirine Vaginal Ring for Prevention of HIV-1 in Women (ASPIRE); <http://clinicaltrials.gov/show/NCT01617096> (last accessed 2 August 2012). 2012.
 53. International Partnership for Microbicides. Safety and Pharmacokinetics of Dapivirine/Maraviroc Vaginal Ring; <http://clinicaltrials.gov/ct2/show/NCT01363037> (last accessed 30 January 2011). 2011.
 54. Dezzutti CS, Russo J, Wang L, Abebe KZ, Li J, Friend DR, et al. Development of HIV-1 rectal-specific microbicides and colonic tissue evaluation. *PLoS ONE*. 2014; 9(7):e102585. doi: [10.1371/journal.pone.0102585](https://doi.org/10.1371/journal.pone.0102585) PMID: [25025306](https://pubmed.ncbi.nlm.nih.gov/25025306/); PubMed Central PMCID: PMC4099179.
 55. Zolopa A, Sax PE, DeJesus E, Mills A, Cohen C, Wohl D, et al. A randomized double-blind comparison of coformulated elvitegravir/cobicistat/emtricitabine/tenofovir disoproxil fumarate versus efavirenz/emtricitabine/tenofovir disoproxil fumarate for initial treatment of HIV-1 infection: analysis of week 96 results. *J Acquir Immune Defic Syndr*. 2013; 63(1):96–100. doi: [10.1097/QAI.0b013e318289545c](https://doi.org/10.1097/QAI.0b013e318289545c) PMID: [23392460](https://pubmed.ncbi.nlm.nih.gov/23392460/).
 56. Walmsley SL, Antela A, Clumeck N, Duiculescu D, Eberhard A, Gutierrez F, et al. Dolutegravir plus abacavir-lamivudine for the treatment of HIV-1 infection. *The New England journal of medicine*. 2013; 369(19):1807–18. doi: [10.1056/NEJMoa1215541](https://doi.org/10.1056/NEJMoa1215541) PMID: [24195548](https://pubmed.ncbi.nlm.nih.gov/24195548/).
 57. Miller CM, Ketlhapile M, Rybasack-Smith H, Rosen S. Why are antiretroviral treatment patients lost to follow-up? A qualitative study from South Africa. *Trop Med Int Health*. 2010; 15 Suppl 1:48–54. Epub 2010/07/14. doi: [10.1111/j.1365-3156.2010.02514.x](https://doi.org/10.1111/j.1365-3156.2010.02514.x) PMID: [20586960](https://pubmed.ncbi.nlm.nih.gov/20586960/); PubMed Central PMCID: PMC3060335.
 58. NIH-sponsored study identifies superior drug regimen for preventing mother-to-child HIV transmission [Internet]. Maryland: NIH; 2014. <http://www.nih.gov/news/health/nov2014/niaid-17.htm>
 59. Nagot N, Kankasa C, Tumwine JK, Meda N, Hofmeyr GJ, Vallo R, et al. Extended pre-exposure prophylaxis with lopinavir-ritonavir versus lamivudine to prevent HIV-1 transmission through breastfeeding up to 50 weeks in infants in Africa (ANRS 12174): a randomised controlled trial. *Lancet*. 2015. doi: [10.1016/S0140-6736\(15\)00984-8](https://doi.org/10.1016/S0140-6736(15)00984-8) PMID: [26603917](https://pubmed.ncbi.nlm.nih.gov/26603917/).
 60. Coovadia H, Moodley D. Improving HIV pre-exposure prophylaxis for infants. *Lancet*. 2015. doi: [10.1016/S0140-6736\(15\)00983-6](https://doi.org/10.1016/S0140-6736(15)00983-6) PMID: [26603916](https://pubmed.ncbi.nlm.nih.gov/26603916/).
 61. Goga AE, Dinh TH, Jackson DJ, Lombard C, Delaney KP, Puren A, et al. First population-level effectiveness evaluation of a national programme to prevent HIV transmission from mother to child, South

- Africa. *J Epidemiol Community Health*. 2014. Epub 2014/11/06. doi: [10.1136/jech-2014-204535](https://doi.org/10.1136/jech-2014-204535) PMID: [25371480](https://pubmed.ncbi.nlm.nih.gov/25371480/).
62. Goga AE, Jackson DJ, Singh M, Lombard C. Early (4–8 weeks post-delivery) Population-level Effectiveness of WHO PMTCT Option A, South Africa, 2012–2013. Pretoria: South African Medical Research Council and National Department of Health of South Africa., 2014.
 63. Massyn N, Peer N, Padarath A, Barron P, Day C, editors. *District Health Barometer 2014/15*. Durban: Health Systems Trust, 2015.
 64. Singh P, Sachs JD. 1 million community health workers in sub-Saharan Africa by 2015. *Lancet*. 2013. Epub 2013/04/02. doi: [10.1016/S0140-6736\(12\)62002-9](https://doi.org/10.1016/S0140-6736(12)62002-9) PMID: [23541538](https://pubmed.ncbi.nlm.nih.gov/23541538/).
 65. Tomlinson M, Doherty T, Ijumba P, Jackson D, Lawn J, Persson LA, et al. Goodstart: a cluster randomised effectiveness trial of an integrated, community-based package for maternal and newborn care, with prevention of mother-to-child transmission of HIV in a South African township. *Trop Med Int Health*. 2014; 19(3):256–66. doi: [10.1111/tmi.12257](https://doi.org/10.1111/tmi.12257) PMID: [24433230](https://pubmed.ncbi.nlm.nih.gov/24433230/).
 66. Woldesenbet SA, Jackson D, Goga AE, Crowley S, Doherty T, Mogashoa MM, et al. Missed Opportunities for Early Infant HIV Diagnosis: Results of a National Study in South Africa. *J Acquir Immune Defic Syndr*. 2014. Epub 2014/12/04. doi: [10.1097/qai.0000000000000460](https://doi.org/10.1097/qai.0000000000000460) PMID: [25469521](https://pubmed.ncbi.nlm.nih.gov/25469521/).
 67. Violari A, Cotton MF, Gibb DM, Babiker AG, Steyn J, Madhi SA, et al. Early antiretroviral therapy and mortality among HIV-infected infants. *The New England journal of medicine*. 2008; 359(21):2233–44. doi: [10.1056/NEJMoa0800971](https://doi.org/10.1056/NEJMoa0800971) PMID: [19020325](https://pubmed.ncbi.nlm.nih.gov/19020325/); PubMed Central PMCID: [PMC2950021](https://pubmed.ncbi.nlm.nih.gov/PMC2950021/).
 68. Johnson LF, Stinson K, Newell ML, Bland RM, Moultrie H, Davies MA, et al. The contribution of maternal HIV seroconversion during late pregnancy and breastfeeding to mother-to-child transmission of HIV. *J Acquir Immune Defic Syndr*. 2012; 59(4):417–25. Epub 2011/12/24. doi: [10.1097/QAI.0b013e3182432f27](https://doi.org/10.1097/QAI.0b013e3182432f27) PMID: [22193774](https://pubmed.ncbi.nlm.nih.gov/22193774/); PubMed Central PMCID: [PMC3378499](https://pubmed.ncbi.nlm.nih.gov/PMC3378499/).
 69. Tenthani L, Haas AD, Tweya H, Jahn A, van Oosterhout JJ, Chimbandira F, et al. Retention in care under universal antiretroviral therapy for HIV-infected pregnant and breastfeeding women ('Option B+') in Malawi. *Aids*. 2014; 28(4):589–98. Epub 2014/01/29. doi: [10.1097/qad.0000000000000143](https://doi.org/10.1097/qad.0000000000000143) PMID: [24468999](https://pubmed.ncbi.nlm.nih.gov/24468999/); PubMed Central PMCID: [PMC34009400](https://pubmed.ncbi.nlm.nih.gov/PMC34009400/).
 70. Hodgson I, Plummer ML, Konopka SN, Colvin CJ, Jonas E, Albertini J, et al. A Systematic Review of Individual and Contextual Factors Affecting ART Initiation, Adherence, and Retention for HIV-Infected Pregnant and Postpartum Women. *PLoS ONE*. 2014; 9(11):e111421. Epub 2014/11/06. doi: [10.1371/journal.pone.0111421](https://doi.org/10.1371/journal.pone.0111421) PMID: [25372479](https://pubmed.ncbi.nlm.nih.gov/25372479/); PubMed Central PMCID: [PMC34221025](https://pubmed.ncbi.nlm.nih.gov/PMC34221025/).
 71. Sibiude J, Mandelbrot L, Blanche S, Le Chenadec J, Boullag-Bonnet N, Faye A, et al. Association between Prenatal Exposure to Antiretroviral Therapy and Birth Defects: An Analysis of the French Perinatal Cohort Study (ANRS CO1/CO11). *PLoS Med*. 2014; 11(4):e1001635. doi: [10.1371/journal.pmed.1001635](https://doi.org/10.1371/journal.pmed.1001635) PMID: [24781315](https://pubmed.ncbi.nlm.nih.gov/24781315/)
 72. Ndirangu J, Newell M-L, Bland RM, Thorne C. Maternal HIV infection associated with small-for-gestational age infants but not preterm births: evidence from rural South Africa. *Human Reproduction*. 2012; 27(6):1846–56. doi: [10.1093/humrep/des090](https://doi.org/10.1093/humrep/des090) PMID: [22442245](https://pubmed.ncbi.nlm.nih.gov/22442245/)
 73. McGrath CJ, Nduati R, Richardson BA, Kristal AR, Mbori-Ngacha D, Farquhar C, et al. The Prevalence of Stunting Is High in HIV-1–Exposed Uninfected Infants in Kenya. *The Journal of nutrition*. 2012; 142(4):757–63. doi: [10.3945/jn.111.148874](https://doi.org/10.3945/jn.111.148874) PMID: [22378334](https://pubmed.ncbi.nlm.nih.gov/22378334/)
 74. Le Doaré K, Bland R, Newell M-L. Neurodevelopment in Children Born to HIV-Infected Mothers by Infection and Treatment Status. *Pediatrics*. 2012; 130(5):e1326–e44. doi: [10.1542/peds.2012-0405](https://doi.org/10.1542/peds.2012-0405) PMID: [23118140](https://pubmed.ncbi.nlm.nih.gov/23118140/)
 75. Gaensbauer JT, Rakhola JT, Onyango-Makumbi C, Mubiru M, Westcott JE, Krebs NF, et al. Impaired haemophilus influenzae type b transplacental antibody transmission and declining antibody avidity through the first year of life represent potential vulnerabilities for HIV-exposed but -uninfected infants. *Clinical and vaccine immunology: CVI*. 2014; 21(12):1661–7. doi: [10.1128/CVI.00356-14](https://doi.org/10.1128/CVI.00356-14) PMID: [25298109](https://pubmed.ncbi.nlm.nih.gov/25298109/); PubMed Central PMCID: [PMC4248779](https://pubmed.ncbi.nlm.nih.gov/PMC4248779/).
 76. Kidzeru EB, Hesselting AC, Passmore J-AS, Myer L, Gamielidien H, Tchakoute CT, et al. In-utero exposure to maternal HIV infection alters T-cell immune responses to vaccination in HIV-uninfected infants. *AIDS*. 2014; 28(10):1421–30. doi: [10.1097/qad.0000000000000292](https://doi.org/10.1097/qad.0000000000000292) 00002030-201406190-00005.
 77. Donahue D, Wainberg M. Cellular and molecular mechanisms involved in the establishment of HIV-1 latency. *Retrovirology*. 2013; 10(1):11. doi: [10.1186/1742-4690-10-11](https://doi.org/10.1186/1742-4690-10-11)
 78. Siliciano JD, Kajdas J, Finzi D, Quinn TC, Chadwick K, Margolick JB, et al. Long-term follow-up studies confirm the stability of the latent reservoir for HIV-1 in resting CD4+ T cells. *Nature medicine*. 2003; 9(6):727–8. doi: [10.1038/nm880](https://doi.org/10.1038/nm880) http://www.nature.com/nm/journal/v9/n6/supinfo/nm880_S1.html. PMID: [12754504](https://pubmed.ncbi.nlm.nih.gov/12754504/)
 79. Deng K, Pertea M, Rongvaux A, Wang L, Durand CM, Ghiaur G, et al. Broad CTL response is required to clear latent HIV-1 due to dominance of escape mutations. *Nature*. 2015; 517(7534):381–5. doi: [10.1038/nature13444](https://doi.org/10.1038/nature13444)

[1038/nature14053](http://www.nature.com/nature/journal/v517/n7534/abs/nature14053.html#supplementary-information) <http://www.nature.com/nature/journal/v517/n7534/abs/nature14053.html#supplementary-information>. PMID: [25561180](https://pubmed.ncbi.nlm.nih.gov/25561180/)

80. Anderson JL, Fromentin R, Corbelli GM, Ostergaard L, Ross AL. Progress Towards an HIV Cure: Update from the 2014 International AIDS Society Symposium. *AIDS research and human retroviruses*. 2014. doi: [10.1089/AID.2014.0236](https://doi.org/10.1089/AID.2014.0236) PMID: [25257573](https://pubmed.ncbi.nlm.nih.gov/25257573/).